

Package: MiscMetabar (via r-universe)

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Type Package

Title Miscellaneous Functions for Metabarcoding Analysis

Version 0.9.2

Description Facilitate the description, transformation, exploration, and reproducibility of metabarcoding analyses. 'MiscMetabar' is mainly built on top of the 'phyloseq', 'dada2' and 'targets' R packages. It helps to build reproducible and robust bioinformatics pipelines in R. 'MiscMetabar' makes ecological analysis of alpha and beta-diversity easier, more reproducible and more powerful by integrating a large number of tools. Important features are described in Taudière A. (2023) <[doi:10.21105/joss.06038](https://doi.org/10.21105/joss.06038)>.

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Encoding UTF-8

LazyData true

Depends R (>= 3.5.0), phyloseq, ggplot2 (>= 3.5.0), dada2, dplyr, purrr

Suggests adespatial, ANCOMBC, BiocManager, Biostrings, circlize, ComplexUpset, DECIPHER, DESeq2, devtools, DT, edgeR, formattable, ggalluvial, ggfittext, gghalves, ggh4x, ggstatsplot, ggridges, ggVennDiagram, glmulti, gtsummary, grDevices, grid, gridExtra, here, httr, iNEXT, indicpecies, jsonlite, knitr, magrittr, metacoder, methods, mia, mixtools, multcompView, networkD3, pak, patchwork, permute, phangorn, phyloseqGraphTest, pbapply, plotly, plyr, reshape2, rmarkdown, rotl, Rtsne, scales, seqinr, SRS, stringr, SummarizedExperiment, testthat (>= 3.0.0), tibble, tidyr, treemapify, vegan, venneuler, vctrs, viridis, withr

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URL <https://github.com/adrientaudiere/MiscMetabar>,
<https://adrientaudiere.github.io/MiscMetabar/>

biocViews Sequencing, Microbiome, Metagenomics, Clustering, Classification, Visualization

BugReports <https://github.com/adrientaudiere/MiscMetabar/issues>

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Imports ape, lifecycle, rlang, stats

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MiscMetabar-package	MiscMetabar <i>package</i>
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Description

Functions to help analyze and visualize metabarcoding data. Mainly based on the phyloseq and dada2 packages.

accu_plot	<i>Plot accumulation curves for phyloseq-class object</i>
-----------	---

Description

Note that as most bioinformatic pipeline discard singleton, accumulation curves from metabarcoding cannot be interpreted in the same way as with conventional biodiversity sampling techniques.

Usage

```
accu_plot(
  physeq,
  fact = NULL,
  add_nb_seq = TRUE,
  step = NULL,
  by.fact = FALSE,
  ci_col = NULL,
  col = NULL,
  lwd = 3,
  leg = TRUE,
  print_sam_names = FALSE,
  ci = 2,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor in physeq@sam_data used to plot different lines
add_nb_seq	(default: TRUE, logical) Either plot accumulation curves using sequences or using samples
step	(Integer) distance among points calculated to plot lines. A low value give better plot but is more time consuming. Only used if add_nb_seq = TRUE.
by.fact	(default: FALSE, logical) First merge the OTU table by factor to plot only one line by factor
ci_col	Color vector for confidence interval. Only use if add_nb_seq = FALSE. If add_nb_seq = TRUE, you can use ggplot to modify the plot.
col	Color vector for lines. Only use if add_nb_seq = FALSE. If add_nb_seq = TRUE, you can use ggplot to modify the plot.
lwd	(default: 3) thickness for lines. Only use if add_nb_seq = FALSE.
leg	(default: TRUE, logical) Plot legend or not. Only use if add_nb_seq = FALSE.
print_sam_names	(default: FALSE, logical) Print samples names or not? Only use if add_nb_seq = TRUE.
ci	(default: 2, integer) Confidence interval value used to multiply the standard error to plot confidence interval
...	Additional arguments passed on to ggplot if add_nb_seq = TRUE or to plot if add_nb_seq = FALSE

Value

A [ggplot2](#) plot representing the richness accumulation plot if add_nb_seq = TRUE, else, if add_nb_seq = FALSE return a base plot.

Author(s)

Adrien Taudière

See Also

[specaccum accu_samp_threshold\(\)](#)

Examples

```
data("GlobalPatterns", package = "phyloseq")
GP <- subset_taxa(GlobalPatterns, GlobalPatterns@tax_table[, 1] == "Archaea")
GP <- rarefy_even_depth(subset_samples_pq(GP, sample_sums(GP) > 3000))
p <- accu_plot(GP, "SampleType", add_nb_seq = TRUE, by.fact = TRUE, step = 10)
p <- accu_plot(GP, "SampleType", add_nb_seq = TRUE, step = 10)

p + theme(legend.position = "none")

p + xlim(c(0, 400))
```

 accu_plot_balanced_modality

Plot accumulation curves with balanced modality and depth rarefaction

Description

This function (i) rarefy (equalize) the number of samples per modality of a factor and (ii) rarefy the number of sequences per sample (depth). The seed is set to 1:nperm. Thus, with exactly the same parameter, including nperm values, results must be identical.

Usage

```
accu_plot_balanced_modality(
  physeq,
  fact,
  nperm = 99,
  step = 2000,
  by.fact = TRUE,
  progress_bar = TRUE,
  quantile_prob = 0.975,
  rarefy_by_sample_before_merging = TRUE,
  sample.size = 1000,
  verbose = FALSE,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) The variable to rarefy. Must be present in the sam_data slot of the physeq object.
nperm	(int) The number of permutations to perform.
step	(int) distance among points calculated to plot lines. A low value give better plot but is more time consuming.
by.fact	(logical, default TRUE) First merge the OTU table by factor to plot only one line by factor
progress_bar	(logical, default TRUE) Do we print progress during the calculation?
quantile_prob	(float, [0:1]) the value to compute the quantile. Minimum quantile is compute using 1-quantile_prob.
rarefy_by_sample_before_merging	(logical, default TRUE): rarefy_by_sample_before_merging = FALSE is buggy for the moment.Please only use rarefy_by_sample_before_merging = TRUE
sample.size	(int) A single integer value equal to the number of reads being simulated, also known as the depth. See phyloseq::rarefy_even_depth() .

verbose (logical). If TRUE, print additional informations.
 ... Other params for be passed on to `accu_plot()` function

Value

A ggplot2 plot representing the richness accumulation plot

Author(s)

Adrien Taudière

See Also

`accu_plot()`, `rarefy_sample_count_by_modality()`, `phyloseq::rarefy_even_depth()`

Examples

```
data_fungi_woNA4Time <-
  subset_samples(data_fungi, !is.na(Time))
data_fungi_woNA4Time@sam_data$Time <- paste0("time-", data_fungi_woNA4Time@sam_data$Time)
accu_plot_balanced_modality(data_fungi_woNA4Time, "Time", nperm = 3)

data_fungi_woNA4Height <-
  subset_samples(data_fungi, !is.na(Height))
accu_plot_balanced_modality(data_fungi_woNA4Height, "Height", nperm = 3)
```

accu_samp_threshold *Compute the number of sequence to obtain a given proportion of ASV in accumulation curves*

Description

Note that as most bioinformatic pipeline discard singleton, accumulation curves from metabarcoding cannot be interpreted in the same way as with conventional biodiversity sampling techniques.

Usage

```
accu_samp_threshold(res_accuplot, threshold = 0.95)
```

Arguments

res_accuplot the result of the function `accu_plot()`
 threshold the proportion of ASV to obtain in each samples

Value

a value for each sample of the number of sequences needed to obtain threshold proportion of the ASV

Author(s)

Adrien Taudière

See Also[accu_plot\(\)](#)**Examples**

```

data("GlobalPatterns", package = "phyloseq")
GP <- subset_taxa(GlobalPatterns, GlobalPatterns@tax_table[, 1] == "Archaea")
GP <- rarefy_even_depth(subset_samples_pq(GP, sample_sums(GP) > 3000))
p <- accu_plot(GP, "SampleType", add_nb_seq = TRUE, by.fact = TRUE, step = 10)

val_threshold <- accu_samp_threshold(p)

summary(val_threshold)

# Plot the number of sequences needed to accumulate 0.95% of ASV in 50%, 75%
# and 100% of samples
p + geom_vline(xintercept = quantile(val_threshold, probs = c(0.50, 0.75, 1)))

```

add_blast_info	<i>Add information from blast_pq() to the tax_table slot of a phyloseq object</i>
----------------	---

Description

Basically a wrapper of [blast_pq\(\)](#) with option `unique_per_seq = TRUE` and `score_filter = FALSE`.

Add the information to the taxtable

Usage

```
add_blast_info(physeq, fasta_for_db, silent = FALSE, ...)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fasta_for_db	path to a fasta file to make the blast database
silent	(logical) If true, no message are printing.
...	Other arguments passed on to blast_pq() function.

Value

A new [phyloseq-class](#) object with more information in tax_table based on a blast on a given database

Author(s)

Adrien Taudière

add_dna_to_phyloseq *Add dna in refseq slot of a physeq object using taxa names and re-names taxa using ASV_1, ASV_2, ...*

Description

Useful in targets bioinformatic pipeline.

Usage

```
add_dna_to_phyloseq(physeq)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

Value

A new [phyloseq-class](#) object with refseq slot and new taxa names

add_funguild_info *Add information about Guild for FUNGI the FUNGuild databse*

Description

Please cite Nguyen et al. 2016 ([doi:10.1016/j.funeco.2015.06.006](https://doi.org/10.1016/j.funeco.2015.06.006))

Usage

```
add_funguild_info(
  physeq,
  taxLevels = c("Kingdom", "Phylum", "Class", "Order", "Family", "Genus", "Species")
)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.
 taxLevels Name of the 7 columns in tax_table required by funguild

Details

This function is mainly a wrapper of the work of others. Please make a reference to FUNGuildR package and the associate [publication](#) if you use this function.

Value

A new object of class physeq with Guild information added to tax_table slot

Author(s)

Adrien Taudière

See Also

[plot_guild_pq\(\)](#)

Examples

```
if (requireNamespace("httr")) {
  d_fung_mini <- add_funguild_info(data_fungi_mini,
    taxLevels = c(
      "Domain",
      "Phylum",
      "Class",
      "Order",
      "Family",
      "Genus",
      "Species"
    )
  )
  sort(table(d_fung_mini@tax_table[, "guild"]), decreasing = TRUE)
}
```

add_info_to_sam_data *Add information to sample_data slot of a phyloseq-class object*

Description

Warning: The value nb_seq and nb_otu may be outdated if you transform your phyloseq object, e.g. using the [subset_taxa_pq\(\)](#) function

Usage

```
add_info_to_sam_data(
  physeq,
  df_info = NULL,
  add_nb_seq = TRUE,
  add_nb_otu = TRUE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
df_info	: A dataframe with rownames matching for sample names of the phyloseq object
add_nb_seq	(Logical, default TRUE) Does we add a column nb_seq collecting the number of sequences per sample?
add_nb_otu	(Logical, default TRUE) Does we add a column nb_otu collecting the number of OTUs per sample?

Value

A phyloseq object with an updated sam_data slot

Author(s)

Adrien Taudière

Examples

```
data_fungi <- add_info_to_sam_data(data_fungi)
boxplot(data_fungi@sam_data$nb_otu ~ data_fungi@sam_data$Time)

new_df <- data.frame(
  variable_1 = runif(n = nsamples(data_fungi), min = 1, max = 20),
  variable_2 = runif(n = nsamples(data_fungi), min = 1, max = 2)
)
rownames(new_df) <- sample_names(data_fungi)
data_fungi <- add_info_to_sam_data(data_fungi, new_df)
plot(data_fungi@sam_data$nb_otu ~ data_fungi@sam_data$variable_1)
```

add_new_taxonomy_pq *Add new taxonomic rank to a phyloseq object.*

Description

One of main use of this function is to add taxonomic assignment from a new database.

Usage

```
add_new_taxonomy_pq(physeq, ref_fasta, suffix = NULL, ...)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
ref_fasta	(required) A link to a database. Pass on to dada2::assignTaxonomy.
suffix	(character) The suffix to name the new columns. If set to NULL (the default), the basename of the file reFasta is used.
...	Other arguments pass on to dada2::assignTaxonomy.

Value

A new [phyloseq-class](#) object with a larger slot `tax_table`"

Author(s)

Adrien Taudière

adonis_pq

Permanova on a phyloseq object

Description

A wrapper for the [vegan::adonis2\(\)](#) function in the case of physeq object.

Usage

```
adonis_pq(
  physeq,
  formula,
  dist_method = "bray",
  merge_sample_by = NULL,
  na_remove = FALSE,
  correction_for_sample_size = FALSE,
  rarefy_nb_seqs = FALSE,
  verbose = TRUE,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
formula	(required) the right part of a formula for vegan::adonis2() . Variables must be present in the <code>physeq@sam_data</code> slot.
dist_method	(default "bray") the distance used. See phyloseq::distance() for all available distances or run phyloseq::distanceMethodList() . For <code>aitchison</code> and <code>robust.aitchison</code> distance, vegan::vegdist() function is directly used.
merge_sample_by	a vector to determine which samples to merge using the merge_samples2() function. Need to be in <code>physeq@sam_data</code>
na_remove	(logical, default FALSE) If set to TRUE, remove samples with NA in the variables set in formula.
correction_for_sample_size	(logical, default FALSE) If set to TRUE, the sample size (number of sequences by samples) is added to formula in the form <code>y~Library_Size + Biological_Effect</code> following recommendation of Weiss et al. 2017 . <code>correction_for_sample_size</code> overcome <code>rarefy_nb_seqs</code> if both are TRUE.

rarefy_nb_seqs (logical, default FALSE) Rarefy each sample (before merging if merge_sample_by is set) using phyloseq::rarefy_even_depth(). If correction_for_sample_size is TRUE, rarefy_nb_seqs will have no effect.

verbose (logical, default TRUE) If TRUE, prompt some messages.

... Other arguments passed on to [vegan::adonis2\(\)](#) function.

Details

This function is mainly a wrapper of the work of others. Please make a reference to [vegan::adonis2\(\)](#) if you use this function.

Value

The function returns an anova.cca result object with a new column for partial R^2 . See help of [vegan::adonis2\(\)](#) for more information.

Author(s)

Adrien Taudière

Examples

```
data(enterotype)

adonis_pq(enterotype, "SeqTech*Enterotype", na_remove = TRUE)
adonis_pq(enterotype, "SeqTech", dist_method = "jaccard")
adonis_pq(enterotype, "SeqTech", dist_method = "robust.aitchison")
```

adonis_rarperm_pq *Permanova (adonis) on permutations of rarefaction even depth*

Description

Permanova are computed on a given number of rarefaction with different seed.number. This reduce the risk of a random drawing of a exceptional situation of an unique rarefaction.

Usage

```
adonis_rarperm_pq(
  physeq,
  formula,
  dist_method = "bray",
  merge_sample_by = NULL,
  na_remove = FALSE,
  rarefy_nb_seqs = FALSE,
  verbose = TRUE,
  nperm = 99,
```

```

    progress_bar = TRUE,
    quantile_prob = 0.975,
    sample.size = min(sample_sums(physeq)),
    ...
  )

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
formula	(required) the right part of a formula for vegan::adonis2() . Variables must be present in the physeq@sam_data slot.
dist_method	(default "bray") the distance used. See phyloseq::distance() for all available distances or run phyloseq::distanceMethodList() . For aitchison and robust.aitchison distance, vegan::vegdist() function is directly used.
merge_sample_by	a vector to determine which samples to merge using the merge_samples2() function. Need to be in physeq@sam_data
na_remove	(logical, default FALSE) If set to TRUE, remove samples with NA in the variables set in formula.
rarefy_nb_seqs	(logical, default FALSE) Rarefy each sample (before merging if merge_sample_by is set) using phyloseq::rarefy_even_depth() . if correction_for_sample_size is TRUE, rarefy_nb_seqs will have no effect.
verbose	(logical, default TRUE) If TRUE, prompt some messages.
nperm	(int, default = 99) The number of permutations to perform.
progress_bar	(logical, default TRUE) Do we print progress during the calculation.
quantile_prob	(float, [0:1]) the value to compute the quantile. Minimum quantile is computed using 1-quantile_prob.
sample.size	(int) A single integer value equal to the number of reads being simulated, also known as the depth. See phyloseq::rarefy_even_depth() .
...	Other params to be passed on to adonis_pq() function

Value

A list of three dataframe representing the mean, the minimum quantile and the maximum quantile value for adonis results. See [adonis_pq\(\)](#).

Author(s)

Adrien Taudière

See Also

[adonis_pq\(\)](#)

Examples

```
if (requireNamespace("vegan")) {
  data_fungi_woNA <-
    subset_samples(data_fungi, !is.na(Time) & !is.na(Height))
  adonis_rarperm_pq(data_fungi_woNA, "Time*Height", na_remove = TRUE, nperm = 3)
}
```

all_object_size	<i>List the size of all objects of the GlobalEnv.</i>
-----------------	---

Description

Code from <https://tolstoy.newcastle.edu.au/R/e6/help/09/01/1121.html>

Usage

```
all_object_size()
```

Value

a list of size

ancombc_pq	<i>Run ANCOMBC2 on phyloseq object</i>
------------	--

Description

A wrapper for the [ANCOMBC: : ancombc2\(\)](#) function

Usage

```
ancombc_pq(physeq, fact, levels_fact = NULL, tax_level = "Class", ...)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor in physeq@sam_data used to plot different lines
levels_fact	(default NULL) The order of the level in the factor. Used for reorder levels and select levels (filter out levels not present en levels_fact)
tax_level	The taxonomic level passed on to ANCOMBC: : ancombc2()
...	Other arguments passed on to ANCOMBC: : ancombc2() function.

Details

This function is mainly a wrapper of the work of others. Please make a reference to [ANCOMBC: : ancombc2\(\)](#) if you use this function.

Value

The result of `ANCOMBC::ancombc2()` function

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("mia")) {
  data_fungi_mini@tax_table <- phyloseq::tax_table(cbind(
    data_fungi_mini@tax_table,
    "taxon" = taxa_names(data_fungi_mini)
  ))
  res_height <- ancombc_pq(
    data_fungi_mini,
    fact = "Height",
    levels_fact = c("Low", "High"),
    verbose = TRUE
  )

  ggplot(
    res_height$res,
    aes(
      y = reorder(taxon, lfc_HeightHigh),
      x = lfc_HeightHigh,
      color = diff_HeightHigh
    )
  ) +
  geom_vline(xintercept = 0) +
  geom_segment(aes(
    xend = 0, y = reorder(taxon, lfc_HeightHigh),
    yend = reorder(taxon, lfc_HeightHigh)
  ), color = "darkgrey") +
  geom_point()

  res_time <- ancombc_pq(
    data_fungi_mini,
    fact = "Time",
    levels_fact = c("0", "15"),
    tax_level = "Family",
    verbose = TRUE
  )
}
```

are_modality_even_depth

Test if the mean number of sequences by samples is link to the modality of a factor

Description

The aim of this function is to provide a warnings if samples depth significantly vary among the modalities of a factor present in the `sam_data` slot.

This function apply a Kruskal-Wallis rank sum test to the number of sequences per samples in function of the factor `fact`.

Usage

```
are_modality_even_depth(physeq, fact, boxplot = FALSE)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the phyloseq package.
<code>fact</code>	(required): Name of the factor to cluster samples by modalities. Need to be in <code>physeq@sam_data</code> .
<code>boxplot</code>	(logical) Do you want to plot boxplot?

Value

The result of a Kruskal-Wallis rank sum test

Author(s)

Adrien Taudière

Examples

```
are_modality_even_depth(data_fungi_mini, "Time")$p.value  
are_modality_even_depth(rarefy_even_depth(data_fungi_mini), "Time")$p.value  
are_modality_even_depth(data_fungi_mini, "Height", boxplot = TRUE)
```

asv2otu

Recluster sequences of an object of class physeq or a list of DNA sequences

Description

This function use the `merge_taxa_vec` function to merge taxa into clusters.

Usage

```

asv2otu(
  physeq = NULL,
  dna_seq = NULL,
  nproc = 1,
  method = "clusterize",
  id = 0.97,
  vsearchpath = "vsearch",
  tax_adjust = 0,
  vsearch_cluster_method = "--cluster_size",
  vsearch_args = "--strand both",
  keep_temporary_files = FALSE,
  swarnpath = "swarm",
  d = 1,
  swarm_args = "--fastidious",
  method_clusterize = "overlap",
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
dna_seq	You may directly use a character vector of DNA sequences in place of physeq args. When physeq is set, dna sequences take the value of physeq@refseq
nproc	(default: 1) Set to number of cpus/processors to use for the clustering
method	(default: clusterize) Set the clustering method. <ul style="list-style-type: none"> • clusterize use the DECIPHER::Clusterize() fonction, • vsearch use the vsearch software (https://github.com/torognes/vsearch) with arguments <code>--cluster_size</code> by default (see args <code>vsearch_cluster_method</code>) and <code>-strand both</code> (see args <code>vsearch_args</code>) • swarm use the swarm
id	(default: 0.97) level of identity to cluster
vsearchpath	(default: vsearch) path to vsearch
tax_adjust	(Default 0) See the man page of merge_taxa_vec() for more details. To conserved the taxonomic rank of the most abundant ASV, set tax_adjust to 0 (default). For the moment only tax_adjust = 0 is robust
vsearch_cluster_method	(default: <code>--cluster_size</code>) See other possible methods in the vsearch manual (e.g. <code>--cluster_size</code> or <code>--cluster_smallmem</code>) <ul style="list-style-type: none"> • <code>--cluster_fast</code> : Clusterize the fasta sequences in filename, automatically sort by decreasing sequence length beforehand. • <code>--cluster_size</code> : Clusterize the fasta sequences in filename, automatically sort by decreasing sequence abundance beforehand. • <code>--cluster_smallmem</code> : Clusterize the fasta sequences in filename without automatically modifying their order beforehand. Sequence are expected to

	be sorted by decreasing sequence length, unless <code>-usersort</code> is used. In that case you may set <code>vsearch_args</code> to <code>vsearch_args = "-strand both -usersort"</code>
<code>vsearch_args</code>	(default : <code>"-strand both"</code>) a one length character element defining other parameters to be passed on to <code>vsearch</code> .
<code>keep_temporary_files</code>	(logical, default: FALSE) Do we keep temporary files <ul style="list-style-type: none"> • <code>temp.fasta</code> (refseq in <code>fasta</code> or <code>dna_seq</code> sequences) • <code>cluster.fasta</code> (centroid if <code>method = "vsearch"</code>) • <code>temp.uc</code> (clusters if <code>method = "vsearch"</code>)
<code>swarmpath</code>	(default: <code>swarm</code>) path to <code>swarm</code>
<code>d</code>	(default: 1) maximum number of differences allowed between two amplicons, meaning that two amplicons will be grouped if they have <code>d</code> (or less) differences
<code>swarm_args</code>	(default : <code>"-fastidious"</code>) a one length character element defining other parameters to be passed on to <code>swarm</code> See other possible methods in the SWARM pdf manual
<code>method_clusterize</code>	(default "overlap") the method for the <code>DECIPHER::Clusterize()</code> method
<code>...</code>	Other arguments passed on to <code>DECIPHER::Clusterize()</code>

Details

This function use the `merge_taxa_vec` function to merge taxa into clusters. By default `tax_adjust = 0`. See the man page of [merge_taxa_vec\(\)](#).

Value

A new object of class `physeq` or a list of cluster if `dna_seq` args was used.

Author(s)

Adrien Taudière

References

VSEARCH can be downloaded from <https://github.com/torognes/vsearch>. More information in the associated publication <https://pubmed.ncbi.nlm.nih.gov/27781170>.

See Also

[vsearch_clustering\(\)](#) and [swarm_clustering\(\)](#)

Examples

```
if (requireNamespace("DECIPHER")) {
  asv2otu(data_fungi_mini)
}

if (requireNamespace("DECIPHER")) {
```

```
asv2otu(data_fungi_mini, method_clusterize = "longest")

if (MiscMetabar::is_swarm_installed()) {
  d_swarm <- asv2otu(data_fungi_mini, method = "swarm")
}
if (MiscMetabar::is_vsearch_installed()) {
  d_vs <- asv2otu(data_fungi_mini, method = "vsearch")
}
}
```

as_binary_otu_table *Transform the otu_table of a [phyloseq-class](#) object into a [phyloseq-class](#) object with a binary otu_table.*

Description

Useful to test if the results are not biased by sequences bias that appended during PCR or NGS pipeline.

Usage

```
as_binary_otu_table(physeq, min_number = 1)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.
min_number (int) the minimum number of sequences to put a 1 in the OTU table.

Value

A physeq object with only 0/1 in the OTU table

Author(s)

Adrien Taudière

Examples

```
data(enterotype)
enterotype_bin <- as_binary_otu_table(enterotype)
```

 biplot_pq

Visualization of two samples for comparison

Description

Graphical representation of distribution of taxa across two samples.

Usage

```
biplot_pq(
  physeq,
  fact = NULL,
  merge_sample_by = NULL,
  rarefy_after_merging = FALSE,
  inverse_side = FALSE,
  left_name = NULL,
  left_name_col = "#4B3E1E",
  left_fill = "#4B3E1E",
  left_col = "#f3f2d9",
  right_name = NULL,
  right_name_col = "#1d2949",
  right_fill = "#1d2949",
  right_col = "#1d2949",
  log10trans = TRUE,
  nudge_y = c(0.3, 0.3),
  geom_label = FALSE,
  text_size = 3,
  size_names = 5,
  y_names = NA,
  ylim_modif = c(1, 1),
  nb_samples_info = TRUE,
  plotly_version = FALSE,
  ...
)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

fact (default: NULL) Name of the factor in `physeq@sam_data`. If left to NULL use the `left_name` and `right_name` parameter as modality.

merge_sample_by (default: NULL) if not NULL samples of `physeq` are merged using the vector set by `merge_sample_by`. This merging used the [merge_samples2\(\)](#). In the case of [biplot_pq\(\)](#) this must be a factor with two levels only.

rarefy_after_merging Rarefy each sample after merging by the modalities `merge_sample_by`

<code>inverse_side</code>	Inverse the side (put the right modality in the left side).
<code>left_name</code>	Name fo the left sample.
<code>left_name_col</code>	Color for the left name
<code>left_fill</code>	Fill fo the left sample.
<code>left_col</code>	Color fo the left sample.
<code>right_name</code>	Name fo the right sample.
<code>right_name_col</code>	Color for the right name
<code>right_fill</code>	Fill fo the right sample.
<code>right_col</code>	Color fo the right sample.
<code>log10trans</code>	(logical) Does abundancy is log10 transformed ?
<code>nudge_y</code>	A parameter to control the y position of abundancy values. If a vector of two values are set. The first value is for the left side. and the second value for the right one. If one value is set, this value is used for both side.
<code>geom_label</code>	(default: FALSE, logical) if TRUE use the <code>ggplot2::geom_label()</code> function instead of <code>ggplot2::geom_text()</code> to indicate the numbers of sequences.
<code>text_size</code>	size for the number of sequences
<code>size_names</code>	size for the names of the 2 samples
<code>y_names</code>	y position for the names of the 2 samples. If NA (default), computed using the maximum abundances values.
<code>ylim_modif</code>	vector of two values. Modificator (by a multiplication) of ylim. If one value is set, this value is used for both limits.
<code>nb_samples_info</code>	(default: TRUE, logical) if TRUE and <code>merge_sample_by</code> is set, add the number of samples merged for both levels.
<code>plotly_version</code>	If TRUE, use <code>plotly::ggplotly()</code> to return a interactive ggplot.
<code>...</code>	Other arguments for the ggplot function

Value

A plot

Author(s)

Adrien Taudière

Examples

```
data_fungi_2Height <- subset_samples(data_fungi_mini, Height %in% c("Low", "High"))
biplot_pq(data_fungi_2Height, "Height", merge_sample_by = "Height")
```

blast_pq	<i>Blast all sequence of refseq slot of a phyloseq-class object against a custom database.</i>
----------	--

Description

Use the blast software.

Usage

```
blast_pq(
  physeq,
  fasta_for_db = NULL,
  database = NULL,
  blastpath = NULL,
  id_cut = 90,
  bit_score_cut = 50,
  min_cover_cut = 50,
  e_value_cut = 1e-30,
  unique_per_seq = FALSE,
  score_filter = TRUE,
  nproc = 1,
  args_makedb = NULL,
  args_blastn = NULL,
  keep_temporary_files = FALSE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fasta_for_db	path to a fasta file to make the blast database
database	path to a blast database
blastpath	path to blast program
id_cut	(default: 90) cut of in identity percent to keep result
bit_score_cut	(default: 50) cut of in bit score to keep result The higher the bit-score, the better the sequence similarity. The bit-score is the requires size of a sequence database in which the current match could be found just by chance. The bit-score is a log2 scaled and normalized raw-score. Each increase by one doubles the required database size (2bit-score).
min_cover_cut	(default: 50) cut of in query cover (%) to keep result
e_value_cut	(default: 1e-30) cut of in e-value (%) to keep result The BLAST E-value is the number of expected hits of similar quality (score) that could be found just by chance.
unique_per_seq	(logical, default FALSE) if TRUE only return the better match (higher bit score) for each sequence

score_filter	(logical, default TRUE) does results are filter by score? If FALSE, id_cut, bit_score_cut, e_value_cut and min_cover_cut are ignored
nproc	(default: 1) Set to number of cpus/processors to use for blast (args -num_threads for blastn command)
args_makedb	Additional parameters parse to makeblastdb command
args_blastn	Additional parameters parse to blastn command
keep_temporary_files	(logical, default: FALSE) Do we keep temporary files <ul style="list-style-type: none"> • db.fasta (refseq transformed into a database) • dbase list of files (output of blastn) • blast_result.txt the summary result of blastn using -outfmt "6 qseqid qlen sseqid slen length p

Value

a blast table

See Also

[blast_to_phyloseq\(\)](#) to use refseq slot as a database

blast_to_derep

Blast some sequence against sequences from of a derep-class object.

Description

Use the blast software.

Usage

```
blast_to_derep(
  derep,
  seq2search,
  blastpath = NULL,
  id_cut = 90,
  bit_score_cut = 50,
  min_cover_cut = 50,
  e_value_cut = 1e-30,
  unique_per_seq = FALSE,
  score_filter = FALSE,
  list_no_output_query = FALSE,
  min_length_seq = 200,
  args_makedb = NULL,
  args_blastn = NULL,
  nproc = 1,
  keep_temporary_files = FALSE
)
```

Arguments

derep	The result of <code>dada2::derepFastq()</code> . A list of derep-class object.
seq2search	(required) path to a fasta file defining the sequences you want to blast against the ASV sequences from the physeq object.
blastpath	path to blast program
id_cut	(default: 90) cut of in identity percent to keep result
bit_score_cut	(default: 50) cut of in bit score to keep result The higher the bit-score, the better the sequence similarity. The bit-score is the requires size of a sequence database in which the current match could be found just by chance. The bit-score is a log2 scaled and normalized raw-score. Each increase by one doubles the required database size (2bit-score).
min_cover_cut	(default: 50) cut of in query cover (%) to keep result
e_value_cut	(default: 1e-30) cut of in e-value (%) to keep result The BLAST E-value is the number of expected hits of similar quality (score) that could be found just by chance.
unique_per_seq	(logical, default FALSE) if TRUE only return the better match (higher bit score) for each sequence
score_filter	(logical, default TRUE) does results are filter by score? If FALSE, <code>id_cut</code> , <code>bit_score_cut</code> , <code>e_value_cut</code> and <code>min_cover_cut</code> are ignored
list_no_output_query	(logical) does the result table include query sequences for which blastn does not find any correspondence?
min_length_seq	(default: 200) Removed sequences with less than <code>min_length_seq</code> from derep before blast. Set to 0 to discard filtering sequences by length.
args_makedb	Additional parameters parse to makeblastdb command
args_blastn	Additional parameters parse to blastn command
nproc	(default: 1) Set to number of cpus/processors to use for blast (<code>args -num_threads</code> for blastn command)
keep_temporary_files	(logical, default: FALSE) Do we keep temporary files : <ul style="list-style-type: none"> • db.fasta (refseq transformed into a database) • dbase list of files (output of blastn) • blast_result.txt the summary result of blastn using <code>-outfmt "6 qseqid qlen sseqid slen length p</code>

Value

A blast table

Author(s)

Adrien Taudière

See Also

[blast_pq\(\)](#) to use refseq slot as query sequences against un custom database and [blast_to_phyloseq\(\)](#) to use refseq slot as a database

blast_to_phyloseq *Blast some sequence against refseq slot of a [phyloseq-class](#) object.*

Description

Use the blast software.

Usage

```
blast_to_phyloseq(
  physeq,
  seq2search,
  blastpath = NULL,
  id_cut = 90,
  bit_score_cut = 50,
  min_cover_cut = 50,
  e_value_cut = 1e-30,
  unique_per_seq = FALSE,
  score_filter = TRUE,
  list_no_output_query = FALSE,
  args_makedb = NULL,
  args_blastn = NULL,
  nproc = 1,
  keep_temporary_files = FALSE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
seq2search	(required) path to a fasta file defining the sequences you want to blast against the ASV sequences from the physeq object.
blastpath	path to blast program
id_cut	(default: 90) cut of in identity percent to keep result
bit_score_cut	(default: 50) cut of in bit score to keep result The higher the bit-score, the better the sequence similarity. The bit-score is the requires size of a sequence database in which the current match could be found just by chance. The bit-score is a log2 scaled and normalized raw-score. Each increase by one doubles the required database size (2bit-score).
min_cover_cut	(default: 50) cut of in query cover (%) to keep result
e_value_cut	(default: 1e-30) cut of in e-value (%) to keep result The BLAST E-value is the number of expected hits of similar quality (score) that could be found just by chance.
unique_per_seq	(logical, default FALSE) if TRUE only return the better match (higher bit score) for each sequence

score_filter (logical, default TRUE) does results are filter by score? If FALSE, id_cut, bit_score_cut, e_value_cut and min_cover_cut are ignored

list_no_output_query (logical) does the result table include query sequences for which blastn does not find any correspondence?

args_makedb Additional parameters parse to makeblastdb command

args_blastn Additional parameters parse to blastn command

nproc (default: 1) Set to number of cpus/processors to use for blast (args -num_threads for blastn command)

keep_temporary_files (logical, default: FALSE) Do we keep temporary files

- db.fasta (refseq transformed into a database)
- dbase list of files (output of blastn)
- blast_result.txt the summary result of blastn using -outfmt "6 qseqid qlen sseqid slen length p

Value

the blast table

See Also

[blast_pq\(\)](#) to use refseq slot as query sequences against un custom database.

Examples

```
## Not run:

blastpath <- "...YOUR_PATH_TO_BLAST..."
blast_to_phyloseq(data_fungi,
  seq2search = system.file("extdata", "ex.fasta",
    package = "MiscMetabar", mustWork = TRUE
  ),
  blastpath = blastpath
)

## End(Not run)
```

Description

This function build tree phylogenetic tree and if `nb_bootstrap` is set, it build also the 3 corresponding bootstrapped tree.

Default parameters are based on [doi:10.12688/f1000research.8986.2](https://doi.org/10.12688/f1000research.8986.2) and phangorn vignette [Estimating phylogenetic trees with phangorn](#). You should understand your data, especially the markers, before using this function.

Note that phylogenetic reconstruction with markers used for metabarcoding are not robust. You must verify the robustness of your phylogenetic tree using taxonomic classification (see vignette [Tree visualization](#)) and bootstrap or multi-tree visualization

Usage

```
build_phytree_pq(
  physeq,
  nb_bootstrap = 0,
  model = "GTR",
  optInv = TRUE,
  optGamma = TRUE,
  rearrangement = "NNI",
  control = phangorn::pml.control(trace = 0),
  optNni = TRUE,
  multicore = FALSE,
  ...
)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the phyloseq package.
<code>nb_bootstrap</code>	(default 0): If a positive number is set, the function also build 3 bootstrapped trees using <code>nb_bootstrap</code> bootstrap samples
<code>model</code>	allows to choose an amino acid models or nucleotide model, see phangorn::optim.pml() for more details
<code>optInv</code>	Logical value indicating whether topology gets optimized (NNI). See phangorn::optim.pml() for more details
<code>optGamma</code>	Logical value indicating whether gamma rate parameter gets optimized. See phangorn::optim.pml() for more details
<code>rearrangement</code>	type of tree tree rearrangements to perform, one of "NNI", "stochastic" or "ratchet" see phangorn::optim.pml() for more details
<code>control</code>	A list of parameters for controlling the fitting process. see phangorn::optim.pml() for more details
<code>optNni</code>	Logical value indicating whether topology gets optimized (NNI). see phangorn::optim.pml() for more details
<code>multicore</code>	(logical) whether models should estimated in parallel. see phangorn::bootstrap.pml() for more details
<code>...</code>	Other params for be passed on to phangorn::optim.pml() function

Details

This function is mainly a wrapper of the work of others. Please make a reference to phangorn package if you use this function.

Value

A list of phylogenetic tree

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("phangorn")) {
  df <- subset_taxa_pq(data_fungi_mini, taxa_sums(data_fungi_mini) > 9000)
  df_tree <- build_phytree_pq(df, nb_bootstrap = 2)
  plot(df_tree$UPGMA)
  phangorn::plotBS(df_tree$UPGMA, df_tree$UPGMA_bs, main = "UPGMA")
  plot(df_tree$NJ, "unrooted")
  plot(df_tree$ML)

  phangorn::plotBS(df_tree$ML$tree, df_tree$ML_bs, p = 20, frame = "circle")
  phangorn::plotBS(
    df_tree$ML$tree,
    df_tree$ML_bs,
    p = 20,
    frame = "circle",
    method = "TBE"
  )
  plot(phangorn::consensusNet(df_tree$ML_bs))
  plot(phangorn::consensusNet(df_tree$NJ_bs))
  ps_tree <- merge_phyloseq(df, df_tree$ML$tree)
}
```

chimera_detection_vs *Detect for chimera taxa using R*[hrefhttps://github.com/torognes/vsearchvsearch](https://github.com/torognes/vsearchvsearch)

Description

Use the VSEARCH software.

Usage

```
chimera_detection_vs(
  seq2search,
  nb_seq,
  vsearchpath = "vsearch",
  abskew = 2,
  min_seq_length = 100,
  vsearch_args = "--fasta_width 0",
  keep_temporary_files = FALSE
)
```

Arguments

seq2search	(required) a list of DNA sequences coercible by function Biostrings::DNAStringSet()
nb_seq	(required) a numeric vector giving the number of sequences for each DNA sequences
vsearchpath	(default: vsearch) path to vsearch
abskew	(int, default 2) The abundance skew is used to distinguish in a three way alignment which sequence is the chimera and which are the parents. The assumption is that chimeras appear later in the PCR amplification process and are therefore less abundant than their parents. The default value is 2.0, which means that the parents should be at least 2 times more abundant than their chimera. Any positive value equal or greater than 1.0 can be used.
min_seq_length	(int, default 100) Minimum length of sequences to be part of the analysis
vsearch_args	(default "--fasta_width 0") A list of other args for vsearch command
keep_temporary_files	(logical, default: FALSE) Do we keep temporary files ? <ul style="list-style-type: none"> • non_chimeras.fasta • chimeras.fasta • borderline.fasta

Details

This function is mainly a wrapper of the work of others. Please make [vsearch](#).

Value

A list of 3 including non-chimera taxa (`$non_chimera`), chimera taxa (`$chimera`) and borderline taxa (`$borderline`)

Author(s)

Adrien Taudière

Examples

```
chimera_detection_vs(
  seq2search = data_fungi@refseq,
  nb_seq = taxa_sums(data_fungi)
)
```

chimera_removal_vs *Search for a list of sequence in an object to remove chimera taxa using VSEARCH*
[R/hrefhttps://github.com/torognes/vsearch/vsearch](https://github.com/torognes/vsearch/vsearch)

Description

Use the VSEARCH software.

Usage

```
chimera_removal_vs(object, type = "Discard_only_chim", clean_pq = FALSE, ...)
```

Arguments

object	(required) A phyloseq-class object or one of dada, derep, data.frame or list coercible to sequences table using the function <code>dada2::makeSequenceTable()</code>
type	(default "Discard_only_chim"). The type define the type of filtering. <ul style="list-style-type: none"> • "Discard_only_chim" will only discard taxa classify as chimera by vsearch • "Select_only_non_chim" will only select taxa classify as non-chimera by vsearch(after filtering taxa based on their sequence length by the parameter <code>min_seq_length</code> from the <code>chimera_detection_vs()</code> function) • "Select_only_chim" will only select taxa classify as chimera by vsearch (after filtering taxa based on their sequence length by the parameter <code>min_seq_length</code> from the <code>chimera_detection_vs()</code> function)
clean_pq	(logical; default FALSE) If TRUE, return the phyloseq object after cleaning using the default parameter of <code>clean_pq()</code> function.
...	Other arguments passed on to <code>chimera_detection_vs()</code> function

Details

This function is mainly a wrapper of the work of others. Please make [vsearch](#).

Value

- I/ a sequences tables if object is of class dada, derep, data.frame or list.
- II/ a phyloseq object without (or with if type = 'Select_only_chim') chimeric taxa

Author(s)

Adrien Taudière

See Also[chimera_detection_vs\(\)](#)**Examples**

```
data_fungi_nochim <- chimera_removal_vs(data_fungi)
data_fungi_nochim_16 <- chimera_removal_vs(data_fungi,
  abskew = 16,
  min_seq_length = 10
)
data_fungi_nochim2 <-
  chimera_removal_vs(data_fungi, type = "Select_only_non_chim")
data_fungi_chimera <-
  chimera_removal_vs(data_fungi, type = "Select_only_chim")
```

`circle_pq`*Plot OTU circle for [phyloseq-class](#) object*

Description

Graphical representation of distribution of taxa across a factor.

Usage

```
circle_pq(
  physeq = NULL,
  fact = NULL,
  taxa = "Order",
  nproc = 1,
  add_nb_seq = TRUE,
  rarefy = FALSE,
  min_prop_tax = 0.01,
  min_prop_mod = 0.1,
  gap_degree = NULL,
  start_degree = NULL,
  row_col = NULL,
  grid_col = NULL,
  log10trans = FALSE,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
taxa	(default: 'Order') Name of the taxonomic rank of interest
nproc	(default 1) Set to number of cpus/processors to use for parallelization
add_nb_seq	(default: TRUE) Represent the number of sequences or the number of OTUs (add_nb_seq = FALSE)
rarefy	(logical) Does each samples modalities need to be rarefy in order to compare them with the same amount of sequences?
min_prop_tax	(default: 0.01) The minimum proportion for taxon to be plotted
min_prop_mod	(default: 0.1) The minimum proportion for modalities to be plotted
gap_degree	Gap between two neighbour sectors. It can be a single value or a vector. If it is a vector, the first value corresponds to the gap after the first sector.
start_degree	The starting degree from which the circle begins to draw. Note this degree is measured in the standard polar coordinate which means it is always reverse-clockwise.
row_col	Color vector for row
grid_col	Grid colors which correspond to sectors. The length of the vector should be either 1 or the number of sectors. It's preferred that grid_col is a named vector of which names correspond to sectors. If it is not a named vector, the order of grid_col corresponds to order of sectors.
log10trans	(logical) Should sequence be log10 transformed (more precisely by log10(1+x))?
...	Additional arguments passed on to chordDiagram or circos.par

Value

A [chordDiagram](#) plot representing the distribution of OTUs or sequences in the different modalities of the factor fact

Author(s)

Adrien Taudière

See Also

[chordDiagram](#)

[circos.par](#)

Examples

```
if (requireNamespace("pbapply")) {
  data("GlobalPatterns", package = "phyloseq")
  GP <- subset_taxa(GlobalPatterns, GlobalPatterns@tax_table[, 1] == "Archaea")
  circle_pq(GP, "SampleType")
}
```

```

circle_pq(GP, "SampleType", add_nb_seq = FALSE)
circle_pq(GP, "SampleType", taxa = "Class")
}

```

clean_pq

Clean phyloseq object by removing empty samples and taxa

Description

In addition, this function check for discrepancy (and rename) between (i) taxa names in refseq, taxonomy table and otu_table and between (ii) sample names in sam_data and otu_table.

Usage

```

clean_pq(
  physeq,
  remove_empty_samples = TRUE,
  remove_empty_taxa = TRUE,
  clean_samples_names = TRUE,
  silent = FALSE,
  verbose = FALSE,
  force_taxa_as_columns = FALSE,
  force_taxa_as_rows = FALSE,
  reorder_asv = FALSE,
  rename_asv = FALSE,
  simplify_taxo = FALSE
)

```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

remove_empty_samples (logical) Do you want to remove samples without sequences (this is done after removing empty taxa)

remove_empty_taxa (logical) Do you want to remove taxa without sequences (this is done before removing empty samples)

clean_samples_names (logical) Do you want to clean samples names?

silent (logical) If true, no message are printing.

verbose (logical) Additional informations in the message the verbose parameter overwrite the silent parameter.

force_taxa_as_columns (logical) If true, if the taxa are rows transpose the otu_table and set taxa_are_rows to false

force_taxa_as_rows	(logical) If true, if the taxa are columns transpose the otu_table and set taxa_are_rows to true
reorder_asv	(logical) if TRUE the otu_table is ordered by the number of sequences of ASV (descending order). Default to FALSE.
rename_asv	(logical) if TRUE, ASV are renamed by their position in the OTU_table (asv_1, asv_2, ...). Default to FALSE. If rename ASV is true, the ASV names in verbose information can be misleading.
simplify_taxo	(logical) if TRUE, correct the taxonomy_table using the MiscMetabar::simplify_taxo() function

Value

A new [phyloseq-class](#) object

compare_pairs_pq	<i>Compare samples in pairs using diversity and number of ASV including shared ASV.</i>
------------------	---

Description

For the moment refseq slot need to be not Null.

Usage

```
compare_pairs_pq(
  physeq = NULL,
  bifactor = NULL,
  modality = NULL,
  merge_sample_by = NULL,
  nb_min_seq = 0,
  veg_index = "shannon",
  na_remove = TRUE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
bifactor	(required) a factor (present in the sam_data slot of the physeq object) presenting the pair names
modality	the name of the column in the sam_data slot of the physeq object to split samples by pairs
merge_sample_by	a vector to determine which samples to merge using the merge_samples2() function. Need to be in physeq@sam_data

nb_min_seq	minimum number of sequences per sample to count the ASV/OTU
veg_index	(default: "shannon") index for the <code>vegan::diversity</code> function
na_remove	(logical, default TRUE) If set to TRUE, remove samples with NA in the variables set in <code>bifactor</code> , <code>modality</code> and <code>merge_sample_by</code> . NA in variables are well managed even if <code>na_remove = FALSE</code> , so <code>na_remove</code> may be useless.

Value

A tibble with information about the number of shared ASV, shared number of sequences and diversity

Examples

```
data_fungi_low_high <- subset_samples(data_fungi, Height %in% c("Low", "High"))
compare_pairs_pq(data_fungi_low_high, bifactor = "Height", merge_sample_by = "Height")
compare_pairs_pq(data_fungi_low_high,
  bifactor = "Height",
  merge_sample_by = "Height", modality = "Time"
)
```

count_seq	<i>Count sequences in fasta or fastq file</i>
-----------	---

Description

Use `grep` to count the number of line with only one '+' (fastq, fastq.gz) or lines starting with a '>' (fasta) to count sequences.

Usage

```
count_seq(file_path = NULL, folder_path = NULL, pattern = NULL)
```

Arguments

file_path	The path to a fasta, fastq or fastq.gz file
folder_path	The path to a folder with fasta, fastq or fastq.gz files
pattern	A pattern to filter files in a folder. E.g. <i>R2</i>

Value

the number of sequences

Author(s)

Adrien Taudière

Examples

```
count_seq(file_path = system.file(
  "extdata",
  "ex.fasta",
  package = "MiscMetabar",
  mustWork = TRUE
))
count_seq(
  folder_path = system.file("extdata", package = "MiscMetabar"),
  pattern = "*.fasta"
)
```

cutadapt_remove_primers

Remove primers using [Rhrefhttps://github.com/marcelm/cutadapt/cutadapt](https://github.com/marcelm/cutadapt/cutadapt)

Description

You need to install **Cutadapt**. See also <https://github.com/VascoElbrecht/JAMP/blob/master/JAMP/R/Cutadapt.R> for another call to cutadapt from R

Usage

```
cutadapt_remove_primers(
  path_to_fastq,
  primer_fw = NULL,
  primer_rev = NULL,
  folder_output = "wo_primers",
  nproc = 1,
  pattern = "fastq.gz",
  pattern_R1 = "_R1",
  pattern_R2 = "_R2",
  nb_files = Inf,
  cmd_is_run = TRUE,
  args_before_cutadapt =
    "source ~/miniconda3/etc/profile.d/conda.sh && conda activate cutadaptenv && "
)
```

Arguments

path_to_fastq	(Required) A path to a folder with fastq files. See list_fastq_files() for help.
primer_fw	(Required, String) The forward primer DNA sequence.
primer_rev	(String) The reverse primer DNA sequence.
folder_output	The path to a folder for output files

nproc	(default 1) Set to number of cpus/processors to use for the clustering
pattern	a pattern to filter files (passed on to list.files function).
pattern_R1	a pattern to filter R1 files (default "R1")
pattern_R2	a pattern to filter R2 files (default "R2")
nb_files	the number of fastq files to list (default FALSE)
cmd_is_run	(logical, default TRUE) Do the cutadapt command is run. If set to FALSE, the only effect of the function is to return a list of command to manually run in a terminal.
args_before_cutadapt	(String) A one line bash command to run before to run cutadapt. For examples, "source ~/miniconda3/etc/profile.d/conda.sh && conda activate cutadaptenv &&" allow to bypass the conda init which asks to restart the shell

Details

This function is mainly a wrapper of the work of others. Please cite cutadapt ([doi:10.14806/ej.17.1.200](https://doi.org/10.14806/ej.17.1.200)).

Value

a list of command

Author(s)

Adrien Taudière

Examples

```
## Not run:
cutadapt_remove_primers("inst/extdata", "TTC", "GAA",
  folder_output = tempdir()
)

cutadapt_remove_primers(
  system.file("extdata",
    package = "dada2"
  ),
  pattern_R1 = "F.fastq.gz",
  pattern_R2 = "R.fastq.gz",
  primer_fw = "TTC",
  primer_rev = "GAA",
  folder_output = tempdir()
)

cutadapt_remove_primers(
  system.file("extdata",
    package = "dada2"
  ),
  pattern_R1 = "F.fastq.gz",
  primer_fw = "TTC",
```

```
    folder_output = tempdir(),
    cmd_is_run = FALSE
  )

  unlink(tempdir(), recursive = TRUE)

  ## End(Not run)
```

data_fungi	<i>Fungal OTU in phyloseq format</i>
------------	--------------------------------------

Description

Fungal OTU in phyloseq format

Usage

```
data(data_fungi)
```

Format

A physeq object containing 1420 taxa with references sequences described by 14 taxonomic ranks and 185 samples described by 7 sample variables:

- *X*: the name of the fastq-file
- *Sample_names*: the names of ... the samples
- *Treename*: the name of an tree
- *Sample_id*: identifier for each sample
- *Height*: height of the sample in the tree
- *Diameter*: diameter of the trunk
- *Time*: time since the dead of the tree

data_fungi_mini	<i>Fungal OTU in phyloseq format</i>
-----------------	--------------------------------------

Description

It is a subset of the data_fungi dataset including only Basidiomycota with more than 5000 sequences.

Usage

```
data(data_fungi_mini)
```

```
data(data_fungi_mini)
```


Format

A physeq object containing 45 taxa with references sequences described by 14 taxonomic ranks and 137 samples described by 7 sample variables:

- *X*: the name of the fastq-file
- *Sample_names*: the names of ... the samples
- *Treename*: the name of an tree
- *Sample_id*: identifier for each sample
- *Height*: height of the sample in the tree
- *Diameter*: diameter of the trunk
- *Time*: time since the dead of the tree

A physeq object containing 45 taxa with references sequences described by 14 taxonomic ranks and 137 samples described by 7 sample variables:

- *X*: the name of the fastq-file
- *Sample_names*: the names of ... the samples
- *Treename*: the name of an tree
- *Sample_id*: identifier for each sample
- *Height*: height of the sample in the tree
- *Diameter*: diameter of the trunk
- *Time*: time since the dead of the tree

Details

Obtain using `data_fungi_mini <- subset_taxa(data_fungi, Phylum == "Basidiomycota")` and then `data_fungi_mini <- subset_taxa_pq(data_fungi_mini, colSums(data_fungi_mini@otu_table) > 5000)`

data_fungi_sp_known *Fungal OTU in phyloseq format*

Description

It is a subset of the data_fungi dataset including only taxa with information at the species level

Usage

```
data(data_fungi_sp_known)
```

Format

A physeq object containing 651 taxa with references sequences described by 14 taxonomic ranks and 185 samples described by 7 sample variables:

- *X*: the name of the fastq-file
- *Sample_names*: the names of ... the samples
- *Treename*: the name of an tree
- *Sample_id*: identifier for each sample
- *Height*: height of the sample in the tree
- *Diameter*: diameter of the trunk
- *Time*: time since the dead of the tree

Details

Obtain using `data_fungi_sp_known <- subset_taxa(data_fungi, !is.na(data_fungi@tax_table[, "Species"]))`

`diff_fct_diff_class` *Compute different functions for different class of vector.*

Description

Mainly an internal function useful in "sapply(..., tapply)" methods

Usage

```
diff_fct_diff_class(
  x,
  numeric_fonction = mean,
  logical_method = "TRUE_if_one",
  character_method = "unique_or_na",
  ...
)
```

Arguments

`x` : a vector

`numeric_fonction` : a function for numeric vector. For ex. sum or mean

`logical_method` : A method for logical vector. One of :

- TRUE_if_one (default)
- NA_if_not_all_TRUE
- FALSE_if_not_all_TRUE

`character_method` : A method for character vector (and factor). One of :

- unique_or_na (default)
 - more_frequent
 - more_frequent_without_equality
- ... Other arguments passed on to the numeric function (ex. na.rm=TRUE)

Value

a single value

Author(s)

Adrien Taudière

Examples

```
diff_fct_diff_class(
  data_fungi@sam_data$Sample_id,
  numeric_fonction = sum,
  na.rm = TRUE
)
diff_fct_diff_class(
  data_fungi@sam_data$Time,
  numeric_fonction = mean,
  na.rm = TRUE
)
diff_fct_diff_class(
  data_fungi@sam_data$Height == "Low",
  logical_method = "TRUE_if_one"
)
diff_fct_diff_class(
  data_fungi@sam_data$Height == "Low",
  logical_method = "NA_if_not_all_TRUE"
)
diff_fct_diff_class(
  data_fungi@sam_data$Height == "Low",
  logical_method = "FALSE_if_not_all_TRUE"
)
diff_fct_diff_class(
  data_fungi@sam_data$Height,
  character_method = "unique_or_na"
)
diff_fct_diff_class(
  c("IE", "IE"),
  character_method = "unique_or_na"
)
diff_fct_diff_class(
  c("IE", "IE", "TE", "TE"),
  character_method = "more_frequent"
)
diff_fct_diff_class(
  c("IE", "IE", "TE", "TE"),
  character_method = "more_frequent_without_equality"
```

)

distri_1_taxa	<i>Distribution of sequences across a factor for one taxon</i>
---------------	--

Description

Focus on one taxon and one factor.

Usage

```
distri_1_taxa(physeq, fact, taxa_name, digits = 2)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor in physeq@sam_data used to plot different lines
taxa_name	(required): the name of the taxa
digits	(default = 2) integer indicating the number of decimal places to be used (see ?round for more information)

Value

a dataframe with levels as rows and information as column :

- the number of sequences of the taxa (nb_seq)
- the number of samples of the taxa (nb_samp)
- the mean (mean_nb_seq) and standard deviation (sd_nb_seq) of the *nb_seq*
- the mean (mean_nb_seq_when_present) *nb_seq* excluding samples with zero
- the total number of samples (nb_total_samp)
- the proportion of samples with the taxa

Author(s)

Adrien Taudière

Examples

```
distri_1_taxa(data_fungi, "Height", "ASV2")
distri_1_taxa(data_fungi, "Time", "ASV81", digits = 1)
```

dist_bycol	<i>Compute paired distances among matrix (e.g. otu_table)</i>
------------	---

Description

May be used to verify ecological distance among samples.

Usage

```
dist_bycol(x, y, method = "bray", nperm = 99, ...)
```

Arguments

x	(required) A first matrix.
y	(required) A second matrix.
method	(default: 'bray') the method to use internally in the vegdist function.
nperm	(int) The number of permutations to perform.
...	Others argument for <code>vegan::vegdist</code> function

Value

A list of length two : (i) a vector of observed distance (`$obs`) and (ii) a matrix of the distance after randomization (`$null`)

Note

the first column of the first matrix is compare to the first column of the second matrix, the second column of the first matrix is compare to the second column of the second matrix and so on.

Author(s)

Adrien Taudière

See Also

[vegdist](#)

dist_pos_control	<i>Calculate ecological distance among positive controls vs distance for all samples</i>
------------------	--

Description

Compute distance among positive controls, i.e. samples which are duplicated to test for variation, for example in (i) a step in the sampling, (ii) a step in the extraction, (iii) a step in the sequencing.

Usage

```
dist_pos_control(physeq, samples_names, method = "bray")
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

samples_names (required) a vector of names for samples with positives controls of the same samples having the same name

method (default: "bray") a method to calculate the distance, parsed to [vegan::vegdist\(\)](#). See [?vegdist](#) for a list of possible values.

Value

A list of two data-frames with (i) the distance among positive controls and (ii) the distance among all samples

Author(s)

Adrien Taudière

Examples

```
data("enterotype")
sam_name_factice <- gsub("TS1_V2", "TS10_V2", sample_names(enterotype))
res_dist_cont <- dist_pos_control(enterotype, sam_name_factice)
hist(unlist(res_dist_cont$distAllSamples))
abline(
  v = mean(unlist(res_dist_cont$dist_controlontrolSamples), na.rm = TRUE),
  col = "red", lwd = 3
)
```

fac2col *Translates a factor into colors.*

Description

Translates a factor into colors.

Usage

```
fac2col(x, col.pal = funky_color, na.col = "grey", seed = NULL)
```

Arguments

x a numeric vector (for num2col) or a vector converted to a factor (for fac2col).
col.pal (default funky_color) a function generating colors according to a given palette.
na.col (default grey) the color to be used for missing values (NAs)
seed (default NULL) a seed for R's random number generated, used to fix the random permutation of colors in the palette used; if NULL, no randomization is used and the colors are taken from the palette according to the ordering of the levels

Value

a color vector

Author(s)

Thibaut Jombart in adegenet package

See Also

The R package RColorBrewer, proposing a nice selection of color palettes. The viridis package, with many excellent palettes

filter_asv_blast *Filter undesirable taxa using blast against a custom database.*

Description

Use the blast software.

Usage

```

filter_asv_blast(
  physeq,
  fasta_for_db = NULL,
  database = NULL,
  clean_pq = TRUE,
  add_info_to_taxtable = TRUE,
  id_filter = 90,
  bit_score_filter = 50,
  min_cover_filter = 50,
  e_value_filter = 1e-30,
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fasta_for_db	path to a fasta file to make the blast database
database	path to a blast database
clean_pq	(logical) If set to TRUE, empty samples and empty ASV are discarded after filtering.
add_info_to_taxtable	(logical, default TRUE) Does the blast information are added to the taxtable ?
id_filter	(default: 90) cut of in identity percent to keep result
bit_score_filter	(default: 50) cut of in bit score to keep result The higher the bit-score, the better the sequence similarity. The bit-score is the requires size of a sequence database in which the current match could be found just by chance. The bit-score is a log2 scaled and normalized raw-score. Each increase by one doubles the required database size (2bit-score).
min_cover_filter	(default: 50) cut of in query cover (%) to keep result
e_value_filter	(default: 1e-30) cut of in e-value (%) to keep result The BLAST E-value is the number of expected hits of similar quality (score) that could be found just by chance.
...	Others options for the blast_pq() function. See ?blast_pq. Note that params unique_per_seq must be lft to TRUE and score_filter must be left to FALSE.

Value

A new [phyloseq-class](#) object.

filter_trim	<i>A wrapper of the function <code>dada2::filterAndTrim()</code> to use in R</i> <i>hrefhttps://books.ropensci.org/targets/targets_pipeline</i>
-------------	--

Description

This function filter and trim (with parameters passed on to `dada2::filterAndTrim()` function) forward sequences or paired end sequence if 'rev' parameter is set. It return the list of files to subsequent analysis in a targets pipeline.

Usage

```
filter_trim(  
  fw = NULL,  
  rev = NULL,  
  output_fw = paste(getwd(), "/output/filterAndTrim_fwd", sep = ""),  
  output_rev = paste(getwd(), "/output/filterAndTrim_rev", sep = ""),  
  ...  
)
```

Arguments

fw	(required) a list of forward fastq files
rev	a list of reverse fastq files for paired end trimming
output_fw	Path to output folder for forward files. By default, this function will create a folder "output/filterAndTrim_fwd" in the current working directory.
output_rev	Path to output folder for reverse files. By default, this function will create a folder "output/filterAndTrim_fwd" in the current working directory.
...	Other parameters passed on to <code>dada2::filterAndTrim()</code> function.

Value

A list of files. If rev is set, will return a list of two lists. The first list is a list of forward files, and the second one is a list of reverse files.

Author(s)

Adrien Taudière

See Also

`dada2::filterAndTrim()`

Examples

```

testFastqs_fw <- c(
  system.file("extdata", "sam1F.fastq.gz", package = "dada2"),
  system.file("extdata", "sam2F.fastq.gz", package = "dada2")
)
testFastqs_rev <- c(
  system.file("extdata", "sam1R.fastq.gz", package = "dada2"),
  system.file("extdata", "sam2R.fastq.gz", package = "dada2")
)

filt_fastq_fw <- filter_trim(testFastqs_fw, output_fw = tempdir())
derep_fw <- derepFastq(filt_fastq_fw[1])
derep_fw

filt_fastq_pe <- filter_trim(testFastqs_fw,
  testFastqs_rev,
  output_fw = tempdir("fw"),
  output_rev = tempdir("rev")
)
derep_fw_pe <- derepFastq(filt_fastq_pe[[1]])
derep_rv_pe <- derepFastq(filt_fastq_pe[[2]])
derep_fw_pe
derep_rv_pe

```

formattable_pq	<i>Create a visualization table to describe taxa distribution across a modality</i>
----------------	---

Description

Allow to visualize a table with graphical input.

Usage

```

formattable_pq(
  physeq,
  modality,
  taxonomic_levels = c("Phylum", "Order", "Family", "Genus"),
  min_nb_seq_taxa = 1000,
  log10trans = FALSE,
  void_style = FALSE,
  lev_col_taxa = "Phylum",
  arrange_by = "nb_seq",
  descending_order = TRUE,
  na_remove = TRUE,
  formattable_args = NULL
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
modality	(required) The name of a column present in the @sam_data slot of the physeq object. Must be a character vector or a factor.
taxonomic_levels	(default = c("Phylum", "Order", "Family", "Genus")) The taxonomic levels (must be present in the @sam_data slot) you want to see and/or used (for example to compute a color) in the table.
min_nb_seq_taxa	(default = 1000) filter out taxa with less than min_nb_seq_taxa sequences
log10trans	(logical, default TRUE) Do sequences count is log10 transformed (using log10(x + 1) to allow 0)
void_style	(logical, default FALSE) Do the default style is discard ?
lev_col_taxa	Taxonomic level used to plot the background color of taxa names
arrange_by	The column used to sort the table. Can take the values NULL, "proportion_samp", "nb_seq" (default), , "nb_sam" "OTU", or a column names from the levels of modality or from taxonomic levels
descending_order	(logical, default TRUE) Do we use descending order when sort the table (if arrange_by is not NULL) ?
na_remove	(logical, default TRUE) if TRUE remove all the samples with NA in the split_by variable of the physeq@sam_data slot
formattable_args	Other args to the formattable function. See examples and formattable::formattable()

Details

This function is mainly a wrapper of the work of others. Please make a reference to formattable::formattable() if you use this function.

Value

A datatable

Author(s)

Adrien Taudière

See Also

formattable::formattable()

Examples

```

if (requireNamespace("formattable")) {
  ## Distribution of the nb of sequences per OTU across Height
  ## modality (nb of sequences are log-transformed).
  ## Only OTU with more than 10000 sequences are taking into account
  ## The Phylum column is discarded
  formattable_pq(
    data_fungi,
    "Height",
    min_nb_seq_taxa = 10000,
    formattable_args = list("Phylum" = FALSE),
    log10trans = TRUE
  )

  ## Distribution of the nb of samples per OTU across Height modality
  ## Only OTU present in more than 50 samples are taking into account
  formattable_pq(
    as_binary_otu_table(data_fungi),
    "Height",
    min_nb_seq_taxa = 50,
    formattable_args = list("nb_seq" = FALSE),
  )

  ## Distribution of the nb of sequences per OTU across Time modality
  ## arranged by Family Name in ascending order.
  ## Only OTU with more than 10000 sequences are taking into account
  ## The Phylum column is discarded
  formattable_pq(
    data_fungi,
    "Time",
    min_nb_seq_taxa = 10000,
    taxonomic_levels = c("Order", "Family", "Genus", "Species"),
    formattable_args = list(
      Order = FALSE,
      Species = formattable::formatter(
        "span",
        style = x ~ formattable::style(
          "font-style" = "italic",
          `color` = ifelse(is.na(x), "white", "grey")
        )
      )
    ),
    arrange_by = "Family",
    descending_order = FALSE
  )
}

if (requireNamespace("formattable")) {
  ## Distribution of the nb of sequences per OTU across Height modality
  ## (nb of sequences are log-transformed).
  ## OTU name background is light gray for Basidiomycota
  ## and dark grey otherwise (Ascomycota)

```

```

## A different color is defined for each modality level
formattable_pq(
  data_fungi,
  "Height",
  taxonomic_levels = c("Phylum", "Family", "Genus"),
  void_style = TRUE,
  formattable_args = list(
    OTU = formattable::formatter(
      "span",
      style = ~ formattable::style(
        "display" = "block",
        `border-radius` = "5px",
        `background-color` = ifelse(Phylum == "Basidiomycota", transp("gray"), "gray")
      ),
      `padding-right` = "2px"
    ),
    High = formattable::formatter(
      "span",
      style = x ~ formattable::style(
        "font-size" = "80%",
        "display" = "inline-block",
        direction = "rtl",
        `border-radius` = "0px",
        `padding-right` = "2px",
        `background-color` = formattable::csscolor(formattable::gradient(
          as.numeric(x), transp("#1a91ff"), "#1a91ff"
        ))
      ),
      width = formattable::percent(formattable::proportion(as.numeric(x), na.rm = TRUE))
    ),
    Low = formattable::formatter(
      "span",
      style = x ~ formattable::style(
        "font-size" = "80%",
        "display" = "inline-block",
        direction = "rtl",
        `border-radius` = "0px",
        `padding-right` = "2px",
        `background-color` = formattable::csscolor(formattable::gradient(
          as.numeric(x),
          transp("green"), "green"
        ))
      ),
      width = formattable::percent(formattable::proportion(as.numeric(x), na.rm = TRUE))
    ),
    Middle = formattable::formatter(
      "span",
      style = x ~ formattable::style(
        "font-size" = "80%",
        "display" = "inline-block",
        direction = "rtl",
        `border-radius` = "0px",
        `padding-right` = "2px",

```

```

    `background-color` = formattable::csscolor(formattable::gradient(
      as.numeric(x), transp("orange"), "orange"
    )),
    width = formattable::percent(formattable::proportion(as.numeric(x), na.rm = TRUE))
  )
)
}

```

funguild_assign

Assign Guilds to Organisms Based on Taxonomic Classification

Description

The original function and documentation was written by Brendan Furneaux in the [FUNGuildR](#) package.

These functions have identical behavior if supplied with a database; however they download the database corresponding to their name by default.

Taxa present in the database are matched to the taxa present in the supplied `otu_table` by exact name. In the case of multiple matches, the lowest (most specific) rank is chosen. No attempt is made to check or correct the classification in `otu_table$Taxonomy`.

Usage

```

funguild_assign(
  otu_table,
  db_funguild = get_funguild_db(),
  tax_col = "Taxonomy"
)

```

Arguments

<code>otu_table</code>	A <code>data.frame</code> with a character column named "Taxonomy" (or another name as specified in <code>tax_col</code>), as well as any other columns. Each entry in <code>otu_table\$Taxonomy</code> should be a comma-, colon-, underscore-, or semicolon-delimited classification of an organism. Rank indicators as given by Sntax ("k:", "p:...") or Unite ("k_", "p_", ...) are also allowed. A character vector, representing only the taxonomic classification, is also accepted.
<code>db_funguild</code>	A <code>data.frame</code> representing the FUNGuild as returned by get_funguild_db() . If not supplied, the default database will be downloaded.
<code>tax_col</code>	A character string, optionally giving an alternate column name in <code>otu_table</code> to use instead of <code>otu_table\$Taxonomy</code> .

Value

A `tibble::tibble` containing all columns of `otu_table`, plus relevant columns of information from the FUNGuild

Author(s)

Brendan Furneaux (orcid: [0000-0003-3522-7363](https://orcid.org/0000-0003-3522-7363)), modified by Adrien Taudière

References

Nguyen NH, Song Z, Bates ST, Branco S, Tedersoo L, Menke J, Schilling JS, Kennedy PG. 2016. *FUNGuild: An open annotation tool for parsing fungal community datasets by ecological guild*. Fungal Ecology 20:241-248.

funky_color

Funky palette color

Description

Funky palette color

Usage

```
funky_color(n)
```

Arguments

n a number of colors

Value

a color palette

Author(s)

Thibaut Jombart in `adeget` package

See Also

The R package `RColorBrewer`, proposing a nice selection of color palettes. The `viridis` package, with many excellent palettes

get_file_extension *Get the extension of a file*

Description

Internally used in `count_seq()`.

Usage

```
get_file_extension(file_path)
```

Arguments

file_path (required): path to a file

Value

The extension of a file.

Author(s)

Adrien Taudière

get_funguild_db *Retrieve the FUNGuild database*

Description

The original function and documentation was written by Brendan Furneaux in the [FUNGuildR](#) package.

Please cite this [publication](#).

Usage

```
get_funguild_db(db_url = "http://www.stbates.org/funguild_db_2.php")
```

Arguments

db_url a length 1 character string giving the URL to retrieve the database from

Value

a `tibble::tibble` containing the database, which can be passed to the db argument of `funguild_assign()`

Author(s)

Brendan Furneaux (orcid: [0000-0003-3522-7363](#)), modified by Adrien Taudière

References

Nguyen NH, Song Z, Bates ST, Branco S, Tedersoo L, Menke J, Schilling JS, Kennedy PG. 2016. *FUNGuild: An open annotation tool for parsing fungal community datasets by ecological guild*. *Fungal Ecology* 20:241-248.

 ggaluv_pq

Alluvial plot for taxonomy and samples factor visualization

Description

Basically a wrapper of **ggalluvial** package

Usage

```
ggaluv_pq(
  physeq,
  taxa_ranks = c("Phylum", "Class", "Order", "Family"),
  wrap_factor = NULL,
  by_sample = FALSE,
  rarefy_by_sample = FALSE,
  fact = NULL,
  type = "nb_seq",
  width = 1.2,
  min.size = 3,
  na_remove = FALSE,
  use_ggfitttext = FALSE,
  use_geom_label = FALSE,
  size_lab = 2,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
taxa_ranks	A vector of taxonomic ranks. For examples c("Family","Genus"). If taxa ranks is not set (default value = c("Phylum", "Class", "Order", "Family")).
wrap_factor	A name to determine which samples to merge using merge_samples2() function. Need to be in physeq@sam_data. Need to be use when you want to wrap by factor the final plot with the number of taxa (type="nb_asv")
by_sample	(logical) If FALSE (default), sample information is not taking into account, so the taxonomy is studied globally. If fact is not NULL, by_sample is automatically set to TRUE.
rarefy_by_sample	(logical, default FALSE) If TRUE, rarefy samples using phyloseq::rarefy_even_depth() function.

fact	(required) Name of the factor in physeq@sam_data used to plot the last column
type	If "nb_seq" (default), the number of sequences is used in plot. If "nb_asv", the number of ASV is plotted.
width	(passed on to <code>ggalluvial::geom_flow()</code>) the width of each stratum, as a proportion of the distance between axes. Defaults to 1/3.
min.size	(passed on to <code>ggfittext::geom_fit_text()</code>) Minimum font size, in points. Text that would need to be shrunk below this size to fit the box will be hidden. Defaults to 4 pt.
na_remove	(logical, default FALSE) If set to TRUE, remove samples with NA in the variables set in formula.
use_ggfittext	(logical, default FALSE) Do we use ggfittext to plot labels?
use_geom_label	(logical, default FALSE) Do we use geom_label to plot labels?
size_lab	Size for label if use_ggfittext is FALSE
...	Other arguments passed on to <code>ggalluvial::geom_flow()</code> function.

Details

This function is mainly a wrapper of the work of others. Please make a reference to ggalluvial package if you use this function.

Value

A ggplot object

Author(s)

Adrien Taudière

See Also

[sankey_pq\(\)](#)

Examples

```
if (requireNamespace("ggalluvial")) {
  ggaluv_pq(data_fungi_mini)
}

if (requireNamespace("ggalluvial")) {
  ggaluv_pq(data_fungi_mini, type = "nb_asv")

  ggaluv_pq(data_fungi_mini, wrap_factor = "Height", by_sample = TRUE, type = "nb_asv") +
    facet_wrap("Height")

  ggaluv_pq(data_fungi_mini,
    width = 0.9, min.size = 10,
    type = "nb_asv", taxa_ranks = c("Phylum", "Class", "Order", "Family", "Genus")
  ) +
```

```

    coord_flip() + scale_x_discrete(limits = rev)
  }

```

ggbetween_pq

Box/Violin plots for between-subjects comparisons of Hill Number

Description

Note that contrary to `hill_pq()`, this function does not take into account for difference in the number of sequences per samples/modalities. You may use `rarefy_by_sample = TRUE` if the mean number of sequences per samples differs among modalities.

Basically a wrapper of function `ggstatsplot::ggbetweenstats()` for object of class `phyloseq`

Usage

```
ggbetween_pq(physeq, fact, one_plot = FALSE, rarefy_by_sample = FALSE, ...)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the <code>phyloseq</code> package.
<code>fact</code>	(required): The variable to test. Must be present in the <code>sam_data</code> slot of the <code>physeq</code> object.
<code>one_plot</code>	(logical, default FALSE) If TRUE, return a unique plot with the three plot inside using the <code>patchwork</code> package.
<code>rarefy_by_sample</code>	(logical, default FALSE) If TRUE, rarefy samples using phyloseq::rarefy_even_depth() function
<code>...</code>	Other arguments passed on to ggstatsplot::ggbetweenstats() function.

Details

This function is mainly a wrapper of the work of others. Please make a reference to `ggstatsplot::ggbetweenstats()` if you use this function.

Value

Either an unique `ggplot2` object (if `one_plot` is TRUE) or a list of 3 `ggplot2` plot:

- `plot_Hill_0` : the `ggbetweenstats` of Hill number 0 (= species richness) against the variable `fact`
- `plot_Hill_1` : the `ggbetweenstats` of Hill number 1 (= Shannon index) against the variable `fact`
- `plot_Hill_2` : the `ggbetweenstats` of Hill number 2 (= Simpson index) against the variable `fact`

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("ggstatsplot")) {
  p <- ggbetween_pq(data_fungi, fact = "Time", p.adjust.method = "BH")
  p[[1]]
  ggbetween_pq(data_fungi, fact = "Height", one_plot = TRUE)
  ggbetween_pq(data_fungi, fact = "Height", one_plot = TRUE, rarefy_by_sample = TRUE)
}
```

ggscatt_pq

Scatterplot with marginal distributions and statistical results against Hill diversity of phyloseq object

Description

Basically a wrapper of function `ggstatsplot::ggscatterstats()` for object of class `phyloseq` and Hill number.

Usage

```
ggscatt_pq(
  physeq,
  num_modality,
  hill_scales = c(0, 1, 2),
  rarefy_by_sample = FALSE,
  one_plot = TRUE,
  ...
)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the <code>phyloseq</code> package.
<code>num_modality</code>	(required) Name of the numeric column in <code>physeq@sam_data</code> to plot and test against hill numberk
<code>hill_scales</code>	(a vector of integer) The list of q values to compute the hill number H^q . If Null, no hill number are computed. Default value compute the Hill number 0 (Species richness), the Hill number 1 (exponential of Shannon Index) and the Hill number 2 (inverse of Simpson Index).
<code>rarefy_by_sample</code>	(logical, default FALSE) If TRUE, rarefy samples using phyloseq::rarefy_even_depth() function.
<code>one_plot</code>	(logical, default FALSE) If TRUE, return a unique plot with the three plot inside using the patchwork package.
<code>...</code>	Other arguments passed on to ggstatsplot::ggscatterstats() function.

Details

This function is mainly a wrapper of the work of others. Please make a reference to `ggstatsplot::ggscatterstats()` if you use this function.

Value

Either an unique `ggplot2` object (if `one_plot` is `TRUE`) or a list of `ggplot2` plot for each `hill_scales`.

Author(s)

Adrien Taudière

See Also

[ggbetween_pq\(\)](#)

Examples

```
if (requireNamespace("ggstatsplot")) {
  ggscatt_pq(data_fungi_mini, "Time", type = "non-parametric")
  ggscatt_pq(data_fungi_mini, "Time", hill_scales = 1:4, type = "parametric")
  ggscatt_pq(data_fungi_mini, "Sample_id",
    hill_scales = c(0, 0.5),
    one_plot = FALSE
  )
}
```

ggvenn_pq

Venn diagram of [phyloseq-class](#) object using `ggVennDiagram::ggVennDiagram` function

Description

Note that you can use `ggplot2` function to customize the plot for ex. `+ scale_fill_distiller(palette = "BuPu", direction = 1)` and `+ scale_x_continuous(expand = expansion(mult = 0.5))`. See examples.

Usage

```
ggvenn_pq(
  physeq = NULL,
  fact = NULL,
  min_nb_seq = 0,
  taxonomic_rank = NULL,
  split_by = NULL,
  add_nb_samples = TRUE,
  add_nb_seq = FALSE,
  rarefy_before_merging = FALSE,
```

```

  rarefy_after_merging = FALSE,
  ...
)

```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the phyloseq package.
<code>fact</code>	(required): Name of the factor to cluster samples by modalities. Need to be in <code>physeq@sam_data</code> .
<code>min_nb_seq</code>	minimum number of sequences by OTUs by samples to take into count this OTUs in this sample. For example, if <code>min_nb_seq=2</code> , each value of 2 or less in the OTU table will not count in the venn diagram
<code>taxonomic_rank</code>	Name (or number) of a taxonomic rank to count. If set to Null (the default) the number of OTUs is counted.
<code>split_by</code>	Split into multiple plot using variable <code>split_by</code> . The name of a variable must be present in <code>sam_data</code> slot of the physeq object.
<code>add_nb_samples</code>	(logical, default TRUE) Add the number of samples to levels names
<code>add_nb_seq</code>	(logical, default FALSE) Add the number of sequences to levels names
<code>rarefy_before_merging</code>	Rarefy each sample before merging by the modalities of args <code>fact</code> . Use <code>phyloseq::rarefy_even_depth</code> function
<code>rarefy_after_merging</code>	Rarefy each sample after merging by the modalities of args <code>fact</code> .
<code>...</code>	Other arguments for the <code>ggVennDiagram::ggVennDiagram</code> function for ex. <code>category.names</code> .

Value

A [ggplot2](#) plot representing Venn diagram of modalities of the argument factor or if `split_by` is set a list of plots.

Author(s)

Adrien Taudière

See Also

[upset_pq\(\)](#)

Examples

```

if (requireNamespace("ggVennDiagram")) {
  ggvenn_pq(data_fungi, fact = "Height")
}

if (requireNamespace("ggVennDiagram")) {
  ggvenn_pq(data_fungi, fact = "Height") +
  ggplot2::scale_fill_distiller(palette = "BuPu", direction = 1)
  pl <- ggvenn_pq(data_fungi, fact = "Height", split_by = "Time")
}

```

```

for (i in seq_along(pl)) {
  p <- pl[[i]] +
    scale_fill_distiller(palette = "BuPu", direction = 1) +
    theme(plot.title = element_text(hjust = 0.5, size = 22))
  print(p)
}

data_fungi2 <- subset_samples(data_fungi, data_fungi@sam_data$Tree_name == "A10-005" |
  data_fungi@sam_data$Height %in% c("Low", "High"))
ggvenn_pq(data_fungi2, fact = "Height")

ggvenn_pq(data_fungi, fact = "Height", add_nb_seq = TRUE, set_size = 4)
ggvenn_pq(data_fungi, fact = "Height", rarefy_before_merging = TRUE)
ggvenn_pq(data_fungi, fact = "Height", rarefy_after_merging = TRUE) +
  scale_x_continuous(expand = expansion(mult = 0.5))
}

```

glmutli_pq

*Automated model selection and multimodel inference with (G)LMs for phyloseq***Description**

See `glmulti::glmulti()` for more information.

Usage

```

glmutli_pq(
  physeq,
  formula,
  fitfunction = "lm",
  hill_scales = c(0, 1, 2),
  aic_step = 2,
  confsetsize = 100,
  plotty = FALSE,
  level = 1,
  method = "h",
  crit = "aicc",
  ...
)

```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`formula` (required) a formula for `glmulti::glmulti()` Variables must be present in the `physeq@sam_data` slot or be one of hill number defined in `hill_scales` or the variable `Abundance` which refer to the number of sequences per sample.

fitfunction	(default "lm")
hill_scales	(a vector of integer) The list of q values to compute the hill number H^q . If Null, no hill number are computed. Default value compute the Hill number 0 (Species richness), the Hill number 1 (exponential of Shannon Index) and the Hill number 2 (inverse of Simpson Index).
aic_step	The value between AIC scores to cut for.
confsetsize	The number of models to be looked for, i.e. the size of the returned confidence set.
plotty	(logical) Whether to plot the progress of the IC profile when running.
level	If 1, only main effects (terms of order 1) are used to build the candidate set. If 2, pairwise interactions are also used (higher order interactions are currently ignored)
method	The method to be used to explore the candidate set of models. If "h" (default) an exhaustive screening is undertaken. If "g" the genetic algorithm is employed (recommended for large candidate sets). If "l", a very fast exhaustive branch-and-bound algorithm is used. Package leaps must then be loaded, and this can only be applied to linear models with covariates and no interactions. If "d", a simple summary of the candidate set is printed, including the number of candidate models.
crit	The Information Criterion to be used. Default is the small-sample corrected AIC (aicc). This should be a function that accepts a fitted model as first argument. Other provided functions are the classic AIC, the Bayes IC (bic), and QAIC/QAICc (qaic and qaicc).
...	Other arguments passed on to <code>glmulti::glmulti()</code> function

Details

This function is mainly a wrapper of the work of others. Please make a reference to `glmulti::glmulti()` if you use this function.

Value

A data.frame summarizing the glmulti results with columns
 -estimates -unconditional_interval -nb_model" -importance -alpha

See Also

`glmulti::glmulti()`

Examples

```
if (requireNamespace("glmulti")) {
  res_glmulti <-
    glmutli_pq(data_fungi, "Hill_0 ~ Hill_1 + Abundance + Time + Height", level = 1)
  res_glmulti
  res_glmulti_interaction <-
    glmutli_pq(data_fungi, "Hill_0 ~ Abundance + Time + Height", level = 2)
```



```

    res_glmulti
  }

```

graph_test_pq

Performs graph-based permutation tests on phyloseq object

Description

A wrapper of `phyloseqGraphTest::graph_perm_test()` for quick plot with important statistics

Usage

```

graph_test_pq(
  physeq,
  fact,
  merge_sample_by = NULL,
  nperm = 999,
  return_plot = TRUE,
  title = "Graph Test",
  na_remove = FALSE,
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor to cluster samples by modalities. Need to be in <code>physeq@sam_data</code> . This should be a factor with two or more levels.
merge_sample_by	a vector to determine which samples to merge using merge_samples2() function. Need to be in <code>physeq@sam_data</code>
nperm	(int) The number of permutations to perform.
return_plot	(logical) Do we return only the result of the test, or do we plot the result?
title	The title of the Graph.
na_remove	(logical, default FALSE) If set to TRUE, remove samples with NA in the variables set in formula.
...	Other params for be passed on to phyloseqGraphTest::graph_perm_test() function

Details

This function is mainly a wrapper of the work of others. Please cite `phyloseqGraphTest` package.

Value

A [ggplot2](#) plot with a subtitle indicating the pvalue and the number of permutations

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("phyloseqGraphTest")) {  
  data(enterotype)  
  graph_test_pq(enterotype, fact = "SeqTech")  
  graph_test_pq(enterotype, fact = "Enterotype", na_remove = TRUE)  
}
```

heat_tree_pq

Heat tree from metacoder package using tax_table slot

Description

Note that the number of ASV is store under the name `n_obs` and the number of sequences under the name `nb_sequences`

Usage

```
heat_tree_pq(physeq, taxonomic_level = NULL, ...)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`taxonomic_level` (default: NULL): a vector of selected taxonomic level using their column numbers (e.g. `taxonomic_level = 1:7`)

`...` Arguments passed on to [heat_tree](#)

Value

A plot

Author(s)

Adrien Taudière

Examples

```

if (requireNamespace("metacoder")) {
  library("metacoder")
  data("GlobalPatterns", package = "phyloseq")

  GPsubset <- subset_taxa(
    GlobalPatterns,
    GlobalPatterns@tax_table[, 1] == "Bacteria"
  )

  GPsubset <- subset_taxa(
    GPsubset,
    rowSums(GPsubset@otu_table) > 5000
  )

  GPsubset <- subset_taxa(
    GPsubset,
    rowSums(is.na(GPsubset@tax_table)) == 0
  )

  heat_tree_pq(GPsubset,
    node_size = n_obs,
    node_color = n_obs,
    node_label = taxon_names,
    tree_label = taxon_names,
    node_size_trans = "log10 area"
  )

  heat_tree_pq(GPsubset,
    node_size = nb_sequences,
    node_color = n_obs,
    node_label = taxon_names,
    tree_label = taxon_names,
    node_size_trans = "log10 area"
  )
}

```

hill_pq

Graphical representation of hill number 0, 1 and 2 across a factor

Description

Hill numbers are the number of equiprobable species giving the same diversity value as the observed distribution. The Hill number 0 correspond to Species richness), the Hill number 1 to the exponential of Shannon Index and the Hill number 2 to the inverse of Simpson Index)

Note that (if `correction_for_sample_size` is TRUE, default behavior) this function use a sqrt of the read numbers in the linear model in order to correct for uneven sampling depth. This correction is only done before tuckey HSD plot and do not change the hill number computed.

Usage

```
hill_pq(
  physeq,
  fact = NULL,
  variable = NULL,
  hill_scales = c(0, 1, 2),
  color_fac = NA,
  letters = FALSE,
  add_points = FALSE,
  add_info = TRUE,
  kruskal_test = TRUE,
  one_plot = FALSE,
  plot_with_tuckey = TRUE,
  correction_for_sample_size = TRUE,
  na_remove = TRUE,
  vioplot = FALSE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required): The variable to test. Must be present in the sam_data slot of the physeq object.
variable	: Alias for factor. Kept only for backward compatibility.
hill_scales	(a vector of integer) The list of q values to compute the hill number H^q . If Null, no hill number are computed. Default value compute the Hill number 0 (Species richness), the Hill number 1 (exponential of Shannon Index) and the Hill number 2 (inverse of Simpson Index).
color_fac	(optional): The variable to color the barplot. For ex. same as fact. Not very useful because ggplot2 plot colors can be change using <code>scale_color_XXX()</code> function.
letters	(optional, default FALSE): If set to TRUE, the plot show letters based on p-values for comparison. Use the multcompLetters function from the package multcompLetters. BROKEN for the moment. Note that na values in The variable param need to be removed (see examples) to use letters.
add_points	(logical, default FALSE): add jitter point on boxplot
add_info	(logical, default TRUE) Do we add a subtitle with information about the number of samples per modality ?
kruskal_test	(logical, default TRUE) Do we test for global effect of our factor on each hill scales values? When kruskal_test is TRUE, the resulting test value are add in each plot in subtitle (unless add_info is FALSE). Moreover, if at least one hill scales is not significantly link to fact ($pval > 0.05$), a message is prompt saying that Tuckey HSD plot is not informative for those Hill scales and letters are not printed.
one_plot	(logical, default FALSE) If TRUE, return a unique plot with the four plot inside using the patchwork package. Note that if letters and one_plot are both TRUE,

	tuckey HSD results are discarded from the unique plot. In that case, use <code>one_plot = FALSE</code> to see the tuckey HSD results in the fourth plot of the resulting list.
<code>plot_with_tuckey</code>	(logical, default TRUE). If <code>one_plot</code> is set to TRUE and <code>letters</code> to FALSE, allow to discard the tuckey plot part with <code>plot_with_tuckey = FALSE</code>
<code>correction_for_sample_size</code>	(logical, default TRUE) This function use a sqrt of the read numbers in the linear model in order to correct for uneven sampling depth in the Tuckey TEST. This params do not change value of Hill number but only the test associated values (including the pvalues). To rarefy samples, you may use the function phyloseq::rarefy_even_depth() .
<code>na_remove</code>	(logical, default TRUE) Do we remove samples with NA in the factor fact ? Note that <code>na_remove</code> is always TRUE when using <code>letters = TRUE</code>
<code>vioplot</code>	(logical, default FALSE) Do we plot violin plot instead of boxplot ?

Value

Either an unique `ggplot2` object (if `one_plot` is TRUE) or a list of `n+1` `ggplot2` plot (with `n` the number of hill scale value). For example, with the default scale value:

- `plot_Hill_0` : the boxplot of Hill number 0 (= species richness) against the variable
- `plot_Hill_1` : the boxplot of Hill number 1 (= Shannon index) against the variable
- `plot_Hill_2` : the boxplot of Hill number 2 (= Simpson index) against the variable
- `plot_tuckey` : plot the result of the Tuckey HSD test

Author(s)

Adrien Taudière

See Also

[psmelt_samples_pq\(\)](#) and [ggbetween_pq\(\)](#)

Examples

```
p <- hill_pq(data_fungi_mini, "Height", hill_scales = 1:2)
p_h1 <- p[[1]] + theme(legend.position = "none")
p_h2 <- p[[2]] + theme(legend.position = "none")
multiplot(plotlist = list(p_h1, p_h2, p[[3]]), cols = 4)

if (requireNamespace("multcompView")) {
  p2 <- hill_pq(data_fungi, "Time",
    correction_for_sample_size = FALSE,
    letters = TRUE, add_points = TRUE,
    plot_with_tuckey = FALSE
  )
  if (requireNamespace("patchwork")) {
    patchwork::wrap_plots(p2, guides = "collect")
  }
}
```

```

# Artificially modify data_fungi to force alpha-diversity effect
data_fungi_modif <- clean_pq(subset_samples_pq(data_fungi, !is.na(data_fungi@sam_data$Height)))
data_fungi_modif@otu_table[data_fungi_modif@sam_data$Height == "High", ] <-
  data_fungi_modif@otu_table[data_fungi_modif@sam_data$Height == "High", ] +
  sample(c(rep(0, ntaxa(data_fungi_modif) / 2), rep(100, ntaxa(data_fungi_modif) / 2)))
p3 <- hill_pq(data_fungi_modif, "Height",
  letters = TRUE, vioplot = TRUE,
  add_points = TRUE
)
}

```

hill_test_rarperm_pq *Test multiple times effect of factor on Hill diversity with different rarefaction even depth*

Description

This reduce the risk of a random drawing of a exceptional situation of an unique rarefaction.

Usage

```

hill_test_rarperm_pq(
  physeq,
  fact,
  hill_scales = c(0, 1, 2),
  nperm = 99,
  sample.size = min(sample_sums(physeq)),
  verbose = FALSE,
  progress_bar = TRUE,
  p_val_signif = 0.05,
  type = "non-parametrique",
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor in physeq@sam_data used to plot different lines
hill_scales	(a vector of integer) The list of q values to compute the hill number H^q . If Null, no hill number are computed. Default value compute the Hill number 0 (Species richness), the Hill number 1 (exponential of Shannon Index) and the Hill number 2 (inverse of Simpson Index).
nperm	(int) The number of permutations to perform.
sample.size	(int) A single integer value equal to the number of reads being simulated, also known as the depth. See phyloseq::rarefy_even_depth() .
verbose	(logical). If TRUE, print additional informations.

progress_bar	(logical, default TRUE) Do we print progress during the calculation?
p_val_signif	(float, [0:1]) The minimum value of p-value to count a test as significant in the prop_signif result.
type	A character specifying the type of statistical approach (See ggstatsplot::ggbetweenstats() for more details): <ul style="list-style-type: none"> • "parametric" • "nonparametric" • "robust" • "bayes"
...	Other arguments passed on to ggstatsplot::ggbetweenstats() function

Value

A list of 6 components :

- method
- expressions
- plots
- pvals
- prop_signif
- statistics

Author(s)

Adrien Taudière

See Also

[ggstatsplot::ggbetweenstats\(\)](#), [hill_pq\(\)](#)

Examples

```
if (requireNamespace("ggstatsplot")) {
  hill_test_rarperm_pq(data_fungi, "Time", nperm = 2)
  res <- hill_test_rarperm_pq(data_fungi, "Height", nperm = 9, p.val = 0.9)
  patchwork::wrap_plots(res$plots[[1]])
  res$plots[[1]][[1]] + res$plots[[2]][[1]] + res$plots[[3]][[1]]
  res$prop_signif
  res_para <- hill_test_rarperm_pq(data_fungi, "Height", nperm = 9, type = "parametrique")
  res_para$plots[[1]][[1]] + res_para$plots[[2]][[1]] + res_para$plots[[3]][[1]]
  res_para$pvals
  res_para$method
  res_para$expressions[[1]]
}
```

hill_tuckey_pq	<i>Calculate hill number and compute Tuckey post-hoc test</i>
----------------	---

Description

Note that, by default, this function use a sqrt of the read numbers in the linear model in order to correct for uneven sampling depth.

Usage

```
hill_tuckey_pq(  
  physeq,  
  modality,  
  hill_scales = c(0, 1, 2),  
  silent = TRUE,  
  correction_for_sample_size = TRUE  
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
modality	(required) the variable to test
hill_scales	(a vector of integer) The list of q values to compute the hill number H^q . If Null, no hill number are computed. Default value compute the Hill number 0 (Species richness), the Hill number 1 (exponential of Shannon Index) and the Hill number 2 (inverse of Simpson Index).
silent	(logical) If TRUE, no message are printing.
correction_for_sample_size	(logical, default TRUE) This function use a sqrt of the read numbers in the linear model in order to correct for uneven sampling depth.

Value

A ggplot2 object

Author(s)

Adrien Taudière

Examples

```
data("GlobalPatterns", package = "phyloseq")  
GlobalPatterns@sam_data[, "Soil_logical"] <-  
  ifelse(GlobalPatterns@sam_data[, "SampleType"] == "Soil", "Soil", "Not Soil")  
hill_tuckey_pq(GlobalPatterns, "Soil_logical")  
hill_tuckey_pq(GlobalPatterns, "Soil_logical", hill_scales = 1:2)
```


iNEXT_pq

*iNterpolation and EXTrapolation of Hill numbers (with iNEXT)***Description**

Note that this function is quite time-consuming due to high dimensionality in metabarcoding community matrix.

Usage

```
iNEXT_pq(physeq, merge_sample_by = NULL, ...)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`merge_sample_by` (default: NULL) if not NULL samples of physeq are merged using the vector set by `merge_sample_by`. This merging used the [merge_samples2\(\)](#). In the case of [biplot_pq\(\)](#) this must be a factor with two levels only.

... Other arguments for the `iNEXT::iNEXT()` function

Value

see [iNEXT::iNEXT\(\)](#) documentation

Author(s)

Adrien Taudière This function is mainly a wrapper of the work of others. Please make a reference to `iNEXT::iNEXT()` if you use this function.

Examples

```
if (requireNamespace("iNEXT")) {
  data("GlobalPatterns", package = "phyloseq")
  GPsubset <- subset_taxa(
    GlobalPatterns,
    GlobalPatterns@tax_table[, 1] == "Bacteria"
  )
  GPsubset <- subset_taxa(
    GPsubset,
    rowSums(GPsubset@otu_table) > 20000
  )
  GPsubset <- subset_taxa(
    GPsubset,
    rowSums(is.na(GPsubset@tax_table)) == 0
  )
  GPsubset@sam_data$human <- GPsubset@sam_data$SampleType %in%
    c("Skin", "Feces", "Tong")
}
```

```
res_iNEXT <- iNEXT_pq(  
  GPsubset,  
  merge_sample_by = "human",  
  q = 1,  
  datatype = "abundance",  
  nboot = 2  
)  
iNEXT::ggiNEXT(res_iNEXT)  
iNEXT::ggiNEXT(res_iNEXT, type = 2)  
iNEXT::ggiNEXT(res_iNEXT, type = 3)  
}
```

is_cutadapt_installed *Test if cutadapt is installed.*

Description

Useful for testthat and examples compilation for R CMD CHECK and test coverage

Usage

```
is_cutadapt_installed(  
  args_before_cutadapt =  
    "source ~/miniconda3/etc/profile.d/conda.sh && conda activate cutadaptenv && "  
)
```

Arguments

args_before_cutadapt
: (String) A one line bash command to run before to run cutadapt. For examples,
"source ~/miniconda3/etc/profile.d/conda.sh && conda activate cutadaptenv &&"
allow to bypass the conda init which asks to restart the shell

Value

A logical that say if cutadapt is install in

Author(s)

Adrien Taudière

Examples

```
MiscMetabar::is_cutadapt_installed()
```

is_falco_installed *Test if falco is installed.*

Description

Useful for testthat and examples compilation for R CMD CHECK and test coverage

Usage

```
is_falco_installed(path = "falco")
```

Arguments

path (default: falco) Path to falco

Value

A logical that say if falco is install in

Author(s)

Adrien Taudière

Examples

```
MiscMetabar::is_falco_installed()
```

is_krona_installed *Test if krona is installed.*

Description

Useful for testthat and examples compilation for R CMD CHECK and test coverage

Usage

```
is_krona_installed(path = "ktImportKrona")
```

Arguments

path (default: krona) Path to krona

Value

A logical that say if krona is install in

Author(s)

Adrien Taudière

Examples

```
MiscMetabar::is_krona_installed()
```

is_mumu_installed *Test if mumu is installed.*

Description

Useful for testthat and examples compilation for R CMD CHECK and test coverage

Usage

```
is_mumu_installed(path = "mumu")
```

Arguments

path (default: mumu) Path to mumu

Value

A logical that say if mumu is install in

Author(s)

Adrien Taudière

Examples

```
MiscMetabar::is_mumu_installed()
```

is_swarm_installed *Test if swarm is installed.*

Description

Useful for testthat and examples compilation for R CMD CHECK and test coverage

Usage

```
is_swarm_installed(path = "swarm")
```

Arguments

path (default: swarm) Path to falco

Value

A logical that say if swarm is install in

Author(s)

Adrien Taudière

Examples

```
MiscMetabar::is_swarm_installed()
```

is_vsearch_installed *Test if vsearch is installed.*

Description

Useful for testthat and examples compilation for R CMD CHECK and test coverage

Usage

```
is_vsearch_installed(path = "vsearch")
```

Arguments

path (default: vsearch) Path to vsearch

Value

A logical that say if vsearch is install in

Author(s)

Adrien Taudière

Examples

```
MiscMetabar::is_vsearch_installed()
```

krona

Make Krona files using R[href="https://github.com/marbl/Krona/wiki/KronaTools"](https://github.com/marbl/Krona/wiki/KronaTools).

Description

Need the installation of kronatools on the computer ([installation instruction](#)).

Usage

```
krona(  
  physeq,  
  file = "krona.html",  
  nb_seq = TRUE,  
  ranks = "All",  
  add_unassigned_rank = 0,  
  name = NULL  
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
file	(required) the location of the html file to save
nb_seq	(logical) If true, Krona set the distribution of sequences in the taxonomy. If False, Krona set the distribution of ASVs in the taxonomy.
ranks	Number of the taxonomic ranks to plot (num of the column in tax_table slot of your physeq object). Default setting plot all the ranks (argument 'All').
add_unassigned_rank	(int) Add unassigned for rank inferior to 'add_unassigned_rank' when necessary.
name	A name for intermediary files, Useful to name your krona result files before merging using merge_krona()

Details

This function is mainly a wrapper of the work of others. Please cite [Krona](#) if you use this function.

Value

A html file

Author(s)

Adrien Taudière

See Also

[merge_krona](#)

Examples

```
data("GlobalPatterns", package = "phyloseq")
GA <- subset_taxa(GlobalPatterns, Phylum == "Acidobacteria")
## Not run:
krona(GA, "Number.of.sequences.html")
krona(GA, "Number.of.ASVs.html", nb_seq = FALSE)
merge_krona(c("Number.of.sequences.html", "Number.of.ASVs.html"))

## End(Not run)
```

LCBD_pq

Compute and test local contributions to beta diversity (LCBD) of samples

Description

A wrapper for the [adespatial::beta.div\(\)](#) function in the case of phyloseq object.

Usage

```
LCBD_pq(phyloseq, p_adjust_method = "BH", ...)
```

Arguments

`phyloseq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.
`p_adjust_method` (chr, default "BH"): the method used to adjust p-value
... Other arguments passed on to [adespatial::beta.div\(\)](#) function

Value

An object of class `beta.div` see [adespatial::beta.div\(\)](#) function for more information

Author(s)

Adrien Taudière This function is mainly a wrapper of the work of others. Please make a reference to [adespatial::beta.div\(\)](#) if you use this function.

See Also

[plot_LCBD_pq, adespatial::beta.div\(\)](#)

Examples

```
if (requireNamespace("adespatial")) {
  res <- LCBD_pq(data_fungi_sp_known, nperm = 5)
  str(res)
  length(res$LCBD)
  length(res$SCBD)
}

if (requireNamespace("adespatial")) {
  LCBD_pq(data_fungi_sp_known, nperm = 5, method = "jaccard")
}
```

list_fastq_files	<i>List fastq files</i>
------------------	-------------------------

Description

Useful for targets bioinformatic pipeline.

Usage

```
list_fastq_files(
  path,
  paired_end = TRUE,
  pattern = "fastq",
  pattern_R1 = "_R1_",
  pattern_R2 = "_R2_",
  nb_files = Inf
)
```

Arguments

path	path to files (required)
paired_end	do you have paired_end files? (default TRUE)
pattern	a pattern to filter files (passed on to list.files function).
pattern_R1	a pattern to filter R1 files (default "R1")
pattern_R2	a pattern to filter R2 files (default "R2")
nb_files	the number of fastq files to list (default FALSE)

Value

a list of one (single end) or two (paired end) list of files files are sorted by names (default behavior of `list.files()`)

Author(s)

Adrien Taudière

Examples

```
list_fastq_files("extdata")
list_fastq_files("extdata", paired_end = FALSE, pattern_R1 = "")
```

lulu

Post Clustering Curation of Amplicon Data.

Description

The original function and documentation was written by Tobias Guldborg Frøslev in the **lulu** package.

This algorithm lulu consumes an OTU table and a matchlist, and evaluates cooccurrence of 'daughters' (potential analytical artefacts) and their 'parents' (~= real biological species/OTUs). The algorithm requires an OTU table (species/site matrix), and a match list. The OTU table can be made with various r-packages (e.g. DADA2) or external pipelines (VSEARCH, USEARCH, QIIME, etc.), and the match-list can be made with external bioinformatic tools like VSEARCH, USEARCH, BLASTN or another algorithm for pair-wise sequence matching.

Usage

```
lulu(
  otu_table,
  matchlist,
  minimum_ratio_type = "min",
  minimum_ratio = 1,
  minimum_match = 84,
  minimum_relative_cooccurrence = 0.95,
  progress_bar = TRUE,
  log_conserved = FALSE
)
```

Arguments

`otu_table` a data.frame with with an OTU table that has sites/samples as columns and OTUs (unique OTU id's) as rows, and observations as read counts.

matchlist	a data.frame containing three columns: (1) OTU id of potential child, (2) OTU id of potential parent, (3) match - % identity between the sequences of the potential parent and potential child OTUs. NB: The matchlist is the product of a mapping of OTU sequences against each other. This is currently carried out by an external script in e.g. Blastn or VSEARCH, prior to running lulu!
minimum_ratio_type	sets whether a potential error must have lower abundance than the parent in all samples min (default), or if an error just needs to have lower abundance on average avg. Choosing lower abundance on average over globally lower abundance will greatly increase the number of designated errors. This option was introduced to make it possible to account for non-sufficiently clustered intraspecific variation, but is not generally recommended, as it will also increase the potential of cluster well-separated, but co-occurring, sequence similar species.
minimum_ratio	sets the minimum abundance ratio between a potential error and a potential parent to be identified as an error. If the minimum_ratio_type is set to min (default), the minimum_ratio applies to the lowest observed ratio across the samples. If the minimum_ratio_type is set to avg (default), the minimum_ratio applies to the mean of observed ratio across the samples. avg. (default is 1).
minimum_match	minimum threshold of sequence similarity for considering any OTU as an error of another can be set (default 84%).
minimum_relative_cooccurrence	minimum co-occurrence rate, i.e. the lower rate of occurrence of the potential error explained by co-occurrence with the potential parent for considering error state.
progress_bar	(Logical, default TRUE) print progress during the calculation or not.
log_conserved	(Logical, default FALSE) conserved log files written in the disk

Details

Please cite the lulu original paper: <https://www.nature.com/articles/s41467-017-01312-x>

Value

Function lulu returns a list of results based on the input OTU table and match list.

- curated_table - a curated OTU table with daughters merged with their matching parents.
- curated_count - number of curated (parent) OTUs.
- curated_otus - ids of the OTUs that were accepted as valid OTUs.
- discarded_count - number of discarded (merged with parent) OTUs.
- discarded_otus - ids of the OTUs that were identified as errors (daughters) and merged with respective parents.
- runtime - time used by the script.
- minimum_match - the id threshold (minimum match \ by user).
- minimum_relative_cooccurrence - minimum ratio of daughter-occurrences explained by co-occurrence with parent (set by user).

- `otu_map` - information of which daughters were mapped to which parents.
- `original_table` - original OTU table.

The matchlist is the product of a mapping of OTU sequences against each other. This is currently carried out by an external script in e.g. BLASTN or VSEARCH, prior to running lulu! Producing the match list requires a file with all the OTU sequences (centroids) - e.g. OTUcentroids.fasta. The matchlist can be produced by mapping all OTUs against each other with an external algorithm like VSEARCH or BLASTN. In VSEARCH a matchlist can be produced e.g. with the following command: `vsearch --usearch_global OTUcentroids.fasta --db OTUcentroids.fasta --strand plus --self --id .80 --iddef 1 --userout matchlist.txt --userfields query+target+id --maxaccepts 0 --query_cov .9 --maxhits 10`. In BLASTN a matchlist can be produced e.g. with the following commands. First we produce a blast-database from the fasta file: `makeblastdb -in OTUcentroids.fasta -parse_seqids -dbtype nucl`, then we match the centroids against that database: `blastn -db OTUcentroids.fasta -num_threads 10 -outfmt '6 qseqid sseqid pident' -out matchlist.txt -qcov_hsp_perc .90 -perc_identity .84 -query OTUcentroids.fasta`

Author(s)

Tobias Guldberg Frøslev (orcid: [0000-0002-3530-013X](https://orcid.org/0000-0002-3530-013X)), modified by Adrien Taudière

lulu_pq

Lulu reclustering of class physeq

Description

See <https://www.nature.com/articles/s41467-017-01312-x> for more information on the method.

Usage

```
lulu_pq(
  physeq,
  nproc = 1,
  id = 0.84,
  vsearchpath = "vsearch",
  verbose = FALSE,
  clean_pq = FALSE,
  keep_temporary_files = FALSE,
  ...
)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the phyloseq package.
<code>nproc</code>	(default 1) Set to number of cpus/processors to use for the clustering
<code>id</code>	(default: 0.84) id for <code>--usearch_global</code> .
<code>vsearchpath</code>	(default: vsearch) path to vsearch.

verbose (logical) if true, print some additional messages.
clean_pq (logical) if true, empty samples and empty ASV are discarded before clustering.
keep_temporary_files (logical, default: FALSE) Do we keep temporary files
... Others args for function `lulu()`

Details

The version of LULU is a fork of Adrien Taudière (<https://github.com/adrietaudiere/lulu>) from <https://github.com/tobiasgf/lulu>

Value

a list of for object

- "new_physeq": The new phyloseq object (class physeq)
- "discrepancy_vector": A vector of discrepancy showing for each taxonomic level the proportion of identic value before and after lulu reclustering. A value of 0.6 stands for 60% of ASV before re-clustering have identical value after re-clustering. In other word, 40% of ASV are assigned to a different taxonomic value. NA value are not counted as discrepancy.
- "res_lulu": A list of the result from the lulu function
- "merged_ASV": the data.frame used to merged ASV

Author(s)

Tobias Guldborg Frøslev <tobiasgf@snm.ku.dk> & Adrien Taudière <adrien.taudiere@zaclys.net>

References

- LULU : <https://github.com/adrietaudiere/lulu> forked from <https://github.com/tobiasgf/lulu>.
- VSEARCH can be downloaded from <https://github.com/torognes/vsearch>.

Examples

```
lulu_pq(data_fungi_sp_known)
```

`merge_krona`*Merge Krona files using [R](https://github.com/marbl/Krona/wiki/KronaTools)<https://github.com/marbl/Krona/wiki/KronaTools>.*

Description

Need the installation of kronatools on the computer ([installation instruction](#)).

Function `merge_krona` allows merging multiple html files in one interactive krona file

Note that you need to use the name args in `krona()` function before `merge_krona()` in order to give good name to each krona pie in the output.

Usage

```
merge_krona(files = NULL, output = "mergeKrona.html")
```

Arguments

<code>files</code>	(required) path to html files to merged
<code>output</code>	path to the output file

Details

This function is mainly a wrapper of the work of others. Please cite [Krona](#) if you use this function.

Value

A html file

Author(s)

Adrien Taudière

See Also

[krona](#)

Examples

```
## Not run:
data("GlobalPatterns", package = "phyloseq")
GA <- subset_taxa(GlobalPatterns, Phylum == "Acidobacteria")
krona(GA, "Number.of.sequences.html", name = "Nb_seq_GP_acidobacteria")
krona(GA, "Number.of.ASVs.html", nb_seq = FALSE, name = "Nb_asv_GP_acidobacteria")
merge_krona(c("Number.of.sequences.html", "Number.of.ASVs.html"), "mergeKrona.html")
unlink(c("Number.of.sequences.html", "Number.of.ASVs.html", "mergeKrona.html"))

## End(Not run)
```

`merge_samples2`*Merge samples by a sample variable or factor*

Description

Firstly release in the `speedyseq` R package by Michael R. McLaren.

This function provides an alternative to `phyloseq::merge_samples()` that better handles sample variables of different types, especially categorical sample variables. It combines the samples in `x` defined by the sample variable or factor `group` by summing the abundances in `otu_table(x)` and combines sample variables by the summary functions in `funs`. The default summary function, `unique_or_na()`, collapses the values within a group to a single unique value if it exists and otherwise returns NA. The new (merged) samples are named by the values in `group`.

Usage

```
merge_samples2(  
  x,  
  group,  
  fun_otu = sum,  
  funs = list(),  
  reorder = FALSE,  
  default_fun = unique_or_na  
)  
  
## S4 method for signature 'phyloseq'  
merge_samples2(  
  x,  
  group,  
  fun_otu = sum,  
  funs = list(),  
  reorder = FALSE,  
  default_fun = unique_or_na  
)  
  
## S4 method for signature 'otu_table'  
merge_samples2(  
  x,  
  group,  
  fun_otu = sum,  
  reorder = FALSE,  
  default_fun = unique_or_na  
)  
  
## S4 method for signature 'sample_data'  
merge_samples2(  
  x,
```

```

    group,
    funs = list(),
    reorder = FALSE,
    default_fun = unique_or_na
  )

```

Arguments

x	A phyloseq, otu_table, or sample_data object
group	A sample variable or a vector of length nsamples(x) defining the sample grouping. A vector must be supplied if x is an otu_table
fun_otu	Function for combining abundances in the otu_table; default is sum. Can be a formula to be converted to a function by <code>purrr::as_mapper()</code>
funs	Named list of merge functions for sample variables; default is unique_or_na
reorder	Logical specifying whether to reorder the new (merged) samples by name
default_fun	Default functions if funs is not set. Per default the function unique_or_na is used. See <code>diff_fct_diff_class()</code> for a useful alternative.

Value

A new phyloseq-class, otu_table or sam_data object depending on the class of the x param

Author(s)

Michael R. McLaren (orcid: [0000-0003-1575-473X](https://orcid.org/0000-0003-1575-473X)) modified by Adrien Taudiere

Examples

```

data(enterotype)

# Merge samples with the same project and clinical status
ps <- enterotype
sample_data(ps) <- sample_data(ps) %>%
  transform(Project.ClinicalStatus = Project:ClinicalStatus)
sample_data(ps) %>% head()
ps0 <- merge_samples2(ps, "Project.ClinicalStatus",
  fun_otu = mean,
  funs = list(Age = mean)
)
sample_data(ps0) %>% head()

```

merge_taxa_vec	<i>Merge taxa in groups (vectorized version)</i>
----------------	--

Description

Firstly release in the `speedyseq` R package by Michael R. McLaren.

Merge taxa in `x` into a smaller set of taxa defined by the vector `group`. Taxa whose value in `group` is NA will be dropped. New taxa will be named according to the most abundant taxon in each group (phyloseq and `otu_table` objects) or the first taxon in each group (all other phyloseq component objects).

If `x` is a phyloseq object with a phylogenetic tree, then the new taxa will be ordered as they are in the tree. Otherwise, the taxa order can be controlled by the `reorder` argument, which behaves like the `reorder` argument in `base::rowsum()`. `reorder = FALSE` will keep taxa in the original order determined by when the member of each group first appears in `taxa_names(x)`; `reorder = TRUE` will order new taxa according to their corresponding value in `group`.

The `tax_adjust` argument controls the handling of taxonomic disagreements within groups. Setting `tax_adjust == 0` causes no adjustment; the taxonomy of the new group is set to the archetype taxon (see below). Otherwise, disagreements within a group at a given rank cause the values at lower ranks to be set to NA. If `tax_adjust == 1` (the default), then a rank where all taxa in the group are already NA is not counted as a disagreement, and lower ranks may be kept if the taxa agree. This corresponds to the original phyloseq behavior. If `tax_adjust == 2`, then these NAs are treated as a disagreement; all ranks are set to NA after the first disagreement or NA.

Usage

```
merge_taxa_vec(x, group, reorder = FALSE, tax_adjust = 1L)

## S4 method for signature 'phyloseq'
merge_taxa_vec(x, group, reorder = FALSE, tax_adjust = 1L)

## S4 method for signature 'otu_table'
merge_taxa_vec(x, group, reorder = FALSE)

## S4 method for signature 'taxonomyTable'
merge_taxa_vec(x, group, reorder = FALSE, tax_adjust = 1L)

## S4 method for signature 'phylo'
merge_taxa_vec(x, group)

## S4 method for signature 'XStringSet'
merge_taxa_vec(x, group, reorder = FALSE)
```

Arguments

x	A phyloseq object or component object
---	---------------------------------------

group	A vector with one element for each taxon in physeq that defines the new groups. see <code>base::rowsum()</code> .
reorder	Logical specifying whether to reorder the taxa by their group values. Ignored if <code>x</code> has (or is) a phylogenetic tree.
tax_adjust	0: no adjustment; 1: phyloseq-compatible adjustment; 2: conservative adjustment

Value

A new phyloseq-class, `otu_table`, `tax_table`, `XStringset` or `sam_data` object depending on the class of the `x` param

Author(s)

Michael R. McLaren (orcid: [0000-0003-1575-473X](https://orcid.org/0000-0003-1575-473X)) modified by Adrien Taudiere

See Also

Function in MiscMetabar that use this function: [asv2otu\(\)](#)

[base::rowsum\(\)](#)

[phyloseq::merge_taxa\(\)](#)

MiscMetabar-deprecated

Deprecated function(s) in the MiscMetabar package

Description

These functions are provided for compatibility with older version of the MiscMetabar package. They may eventually be completely removed.

Usage

```
physeq_graph_test(...)
```

Arguments

... Parameters to be passed on to the modern version of the function

Value

Depend on the functions.

Details

graph_test_pq	now a synonym for physeq_graph_test
adonis_pq	now a synonym for adonis_phyloseq
clean_pq	now a synonym for clean_physeq
lulu_pq	now a synonym for lulu_phyloseq
circle_pq	now a synonym for otu_circle
biplot_pq	now a synonym for biplot_physeq
read_pq	now a synonym for read_phyloseq
write_pq	now a synonym for write_phyloseq
sankey_pq	now a synonym for sankey_phyloseq
summary_plot_pq	now a synonym for summary_plot_phyloseq
plot_edgeR_pq	now a synonym for plot_edgeR_phyloseq
plot_deseq2_pq	now a synonym for plot_deseq2_phyloseq
venn_pq	now a synonym for venn_phyloseq
ggvenn_pq	now a synonym for ggVenn_phyloseq
hill_tuckey_pq	now a synonym for hill_tuckey_phyloseq
hill_pq	now a synonym for hill_phyloseq
heat_tree_pq	now a synonym for physeq_heat_tree
compare_pairs_pq	now a synonym for multiple_share_bisamples

multipatt_pq

Test and plot multipatt result

Description

A wrapper for the `indicspecies::multipatt()` function in the case of physeq object.

Usage

```

multipatt_pq(
  physeq,
  fact,
  p_adjust_method = "BH",
  pval = 0.05,
  control = permute::how(nperm = 999),
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor in physeq@sam_data used to plot different lines

p_adjust_method (chr, default "BH"): the method used to adjust p-value
pval (int, default 0.05): the value to determine the significance of LCBD
control see ?indicspecies::multipatt()
... Other arguments passed on to [indicspecies::multipatt\(\)](#) function

Details

This function is mainly a wrapper of the work of others. Please make a reference to [indicspecies::multipatt\(\)](#) if you use this function.

Value

A ggplot2 object

Author(s)

Adrien Taudière

Examples

```

if (requireNamespace("indicspecies")) {
  data(data_fungi)
  data_fungi_ab <- subset_taxa_pq(data_fungi, taxa_sums(data_fungi) > 10000)
  multipatt_pq(subset_samples(data_fungi_ab, !is.na(Time)), fact = "Time")
}

if (requireNamespace("indicspecies")) {
  multipatt_pq(subset_samples(data_fungi_ab, !is.na(Time)),
    fact = "Time",
    max.order = 1, control = permute::how(nperm = 99)
  )
}

```

multiplot

Multiple plot function

Description

ggplot objects can be passed in ..., or to plotlist (as a list of ggplot objects)

If the layout is something like `matrix(c(1,2,3,3), nrow=2, byrow=TRUE)`, then plot 1 will go in the upper left, 2 will go in the upper right, and 3 will go all the way across the bottom.

Usage

```
multiplot(..., plotlist = NULL, cols = 1, layout = NULL)
```

Arguments

...	list of ggplot objects
plotlist	list of ggplot objects
cols	number of columns
layout	A matrix specifying the layout. If present, 'cols' is ignored.

Value

Nothing. Print the list of ggplot objects

multitax_bar_pq	<i>Plot taxonomic distribution across 3 taxonomic levels and optionally one sample factor</i>
-----------------	---

Description

Note that lv13 need to be nested in lv12 which need to be nested in lv11

Usage

```
multitax_bar_pq(
  physeq,
  lv11,
  lv12,
  lv13,
  fact = NULL,
  nb_seq = TRUE,
  log10trans = TRUE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
lv11	(required) Name of the first (higher) taxonomic rank of interest
lv12	(required) Name of the second (middle) taxonomic rank of interest
lv13	(required) Name of the first (lower) taxonomic rank of interest
fact	Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data. If not set, the taxonomic distribution is plot for all samples together.
nb_seq	(logical; default TRUE) If set to FALSE, only the number of ASV is count. Concretely, physeq otu_table is transformed in a binary otu_table (each value different from zero is set to one)
log10trans	(logical, default TRUE) If TRUE, the number of sequences (or ASV if nb_seq = FALSE) is log10 transformed.

Value

A ggplot2 graphic

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("ggh4x")) {
  multitax_bar_pq(data_fungi_sp_known, "Phylum", "Class", "Order", "Time")
  multitax_bar_pq(data_fungi_sp_known, "Phylum", "Class", "Order")
  multitax_bar_pq(data_fungi_sp_known, "Phylum", "Class", "Order",
    nb_seq = FALSE, log10trans = FALSE
  )
}
```

multi_biplot_pq

Visualization of a collection of couples of samples for comparison

Description

This allow to plot all the possible `biplot_pq()` combination using one factor.

Usage

```
multi_biplot_pq(physeq, split_by = NULL, pairs = NULL, na_remove = TRUE, ...)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
split_by	(required if pairs is NULL) the name of the factor to make all combination of couples of values
pairs	(required if pairs is NULL) the name of the factor in physeq@sam_data slot to make plot by pairs of samples. Each level must be present only two times. Note that if you set pairs, you also must set fact arguments to pass on to biplot_pq() .
na_remove	(logical, default TRUE) if TRUE remove all the samples with NA in the split_by variable of the physeq@sam_data slot
...	Other parameters passed on to biplot_pq()

Value

a list of ggplot object

Author(s)

Adrien Taudière

Examples

```
data_fungi_abun <- subset_taxa_pq(data_fungi, taxa_sums(data_fungi) > 10000)
p <- multi_biplot_pq(data_fungi_abun, "Height")
lapply(p, print)
```

mumu_pq

*MUMU reclustering of class physeq***Description**

See <https://www.nature.com/articles/s41467-017-01312-x> for more information on the original method LULU. This is a wrapper of **mumu** a C++ re-implementation of LULU by Frédéric Mahé

Usage

```
mumu_pq(
  physeq,
  nproc = 1,
  id = 0.84,
  vsearchpath = "vsearch",
  mumupath = "mumu",
  verbose = FALSE,
  clean_pq = TRUE,
  keep_temporary_files = FALSE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
nproc	(default 1) Set to number of cpus/processors to use for the clustering
id	(default: 0.84) id for <code>-usearch_global</code> .
vsearchpath	(default: vsearch) path to vsearch.
mumupath	path to mumu. See mumu for installation instruction
verbose	(logical) if true, print some additional messages.
clean_pq	(logical) if true, empty samples and empty ASV are discarded before clustering.
keep_temporary_files	(logical, default: FALSE) Do we keep temporary files

Details

This function is mainly a wrapper of the work of others. Please cite **mumu** and **lulu** if you use this function for your work.

Value

a list of for object

- "new_physeq": The new phyloseq object (class phyloseq)
- "mumu_results": The log file of the mumu software. Run `man mumu` into bash to obtain details about columns' signification.

Author(s)

Frédéric Mahé & Adrien Taudière <adrien.taudiere@zaclys.net>

References

- MUMU: <https://github.com/frederic-mahe/mumu>
- VSEARCH can be downloaded from <https://github.com/torognes/vsearch>.

Examples

```
## Not run:  
mumu_pq(data_fungi_sp_known)  
  
## End(Not run)
```

normalize_prop_pq *Normalize OTU table using samples depth*

Description

This function implement the method proposed by McKnight et al. 2018 ([doi:10.5061/dryad.tn8qs35](https://doi.org/10.5061/dryad.tn8qs35))

Usage

```
normalize_prop_pq(physeq, base_log = 2, constante = 10000, digits = 4)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
base_log	(integer, default 2) the base for log-transformation. If set to NULL or NA, no log-transformation is compute after normalization.
constante	a constante to multiply the otu_table values
digits	(default = 2) integer indicating the number of decimal places to be used (see ?round for more information)

Value

A new [phyloseq-class](#) object with otu_table count normalize and log transformed (if base_log is an integer)

Author(s)

Adrien Taudière

Examples

```
taxa_sums(data_fungi_mini)
data_f_norm <- normalize_prop_pq(data_fungi_mini)
taxa_sums(data_f_norm)
ggplot(data.frame(
  "norm" = scale(taxa_sums(data_f_norm)),
  "raw" = scale(taxa_sums(data_fungi_mini)),
  "name_otu" = taxa_names(data_f_norm)
)) +
  geom_point(aes(x = raw, y = norm))

data_f_norm <- normalize_prop_pq(data_fungi_mini, base_log = NULL)
```

perc

Convert a value (or a fraction x/y) in percentage

Description

Mostly for internal use.

Usage

```
perc(x, y = NULL, accuracy = 0, add_symbol = FALSE)
```

Arguments

x	(required): value
y	if y is set, compute the division of x by y
accuracy	number of digits (number of digits after zero)
add_symbol	if set to TRUE add the % symbol to the value

Value

The percentage value (number or character if add_symbol is set to TRUE)

Author(s)

Adrien Taudière

phyloseq_to_edgeR *Convert phyloseq OTU count data into DGEList for edgeR package*

Description

Convert phyloseq OTU count data into DGEList for edgeR package

Usage

```
phyloseq_to_edgeR(physeq, group, method = "RLE", ...)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
group	(required) A character vector or factor giving the experimental group/condition for each sample/library. Alternatively, you may provide the name of a sample variable. This name should be among the output of <code>sample_variables(physeq)</code> , in which case <code>get_variable(physeq, group)</code> would return either a character vector or factor. This is passed on to DGEList , and you may find further details or examples in its documentation.
method	The label of the edgeR-implemented normalization to use. See calcNormFactors for supported options and details. The default option is "RLE", which is a scaling factor method proposed by Anders and Huber (2010). At time of writing, the edgeR package supported the following options to the method argument: <code>c("TMM", "RLE", "upperquartile", "none")</code> .
...	Additional arguments passed on to DGEList

Value

A DGEList object. See [edgeR::estimateTagwiseDisp\(\)](#) for more details.

physeq_or_string_to_dna
Return a DNASTringSet object from either a character vector of DNA sequences or the refseq slot of a phyloseq-class object

Description

Internally used in [vsearch_clustering\(\)](#), [swarm_clustering\(\)](#) and [asv2otu\(\)](#).

Usage

```
physeq_or_string_to_dna(physeq = NULL, dna_seq = NULL)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

dna_seq You may directly use a character vector of DNA sequences in place of physeq args. When physeq is set, dna sequences take the value of physeq@refseq

Value

An object of class DNASTringSet (see the [Biostrings::DNASTringSet\(\)](#) function)

Author(s)

Adrien Taudière

See Also

[Biostrings::DNASTringSet\(\)](#)

Examples

```
dna <- physeq_or_string_to_dna(data_fungi)
dna

sequences_ex <- c(
  "TACCTATGTTGCCTTGGCGGCTAAACCTACCCGGGATTTGATGGGGCGAATTAATAACGAATTCATTGAATCA",
  "TACCTATGTTGCCTTGGCGGCTAAACCTACCCGGGATTTGATGGGGCGAATTACCTGGTAAGGCCCACTT",
  "TACCTATGTTGCCTTGGCGGCTAAACCTACCCGGGATTTGATGGGGCGAATTACCTGGTAGAGGTG",
  "TACCTATGTTGCCTTGGCGGCTAAACCTACC",
  "CGGGATTTGATGGCGAATTACCTGGTATTTTAGCCCACTTACCCGGTACCATGAGGTG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACCTGG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACAAAG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACAAAG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACAAAG"
)
dna2 <- physeq_or_string_to_dna(dna_seq = sequences_ex)
dna2
```

plot_ancombc_pq

Plot ANCOMBC2 result for phyloseq object

Description

Graphical representation of ANCOMBC2 result.

Usage

```
plot_ancombc_pq(
  physeq,
  ancombc_res,
  filter_passed = TRUE,
  filter_diff = TRUE,
  min_abs_lfc = 0,
  tax_col = "Genus",
  tax_label = "Species",
  add_marginal_violplot = TRUE,
  add_label = TRUE,
  add_hline_cut_lfc = NULL
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
ancombc_res	(required) the result of the ancombc_pq function For the moment only bimodal factors are possible.
filter_passed	(logical, default TRUE) Do we filter using the column passed_ss? The passed_ss value is TRUE if the taxon passed the sensitivity analysis, i.e., adding different pseudo-counts to 0s would not change the results.
filter_diff	(logical, default TRUE) Do we filter using the column diff? The diff value is TRUE if the taxon is significant (has q less than alpha)
min_abs_lfc	(integer, default 0) Minimum absolute value to filter results based on Log Fold Change. For ex. a value of 1 filter out taxa for which the abundance in a given level of the modality is not at least the double of the abundance in the other level.
tax_col	The taxonomic level (must be present in tax_table slot) to color the points
tax_label	The taxonomic level (must be present in tax_table slot) to add label
add_marginal_violplot	(logical, default TRUE) Do we add a marginal violplot representing all the taxa lfc from ancombc_res.
add_label	(logical, default TRUE) Do we add a label?
add_hline_cut_lfc	(logical, default NULL) Do we add two horizontal lines when min_abs_lfc is set (different from zero)?

Details

This function is mainly a wrapper of the work of others. Please make a reference to ANCOMBC: : ancombc2() if you use this function.

Value

A ggplot2 object. If add_marginal_violplot is TRUE, this is a patchworks of plot made using patchwork::plot_layout().

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("mia")) {
  data_fungi_mini@tax_table <- phyloseq::tax_table(cbind(
    data_fungi_mini@tax_table,
    "taxon" = taxa_names(data_fungi_mini)
  ))

  res_time <- ancombc_pq(
    data_fungi_mini,
    fact = "Time",
    levels_fact = c("0", "15"),
    tax_level = "taxon",
    verbose = TRUE
  )

  plot_ancombc_pq(data_fungi_mini, res_time,
    filter_passed = FALSE,
    tax_label = "Genus", tax_col = "Order"
  )
  plot_ancombc_pq(data_fungi_mini, res_time, tax_col = "Genus")
  plot_ancombc_pq(data_fungi_mini, res_time,
    filter_passed = FALSE,
    filter_diff = FALSE, tax_col = "Family", add_label = FALSE
  )
}
```

`plot_deseq2_pq`*Plot DESeq2 results for a phyloseq or a DESeq2 object.*

Description

Graphical representation of DESeq2 analysis.

Usage

```
plot_deseq2_pq(
  data,
  contrast = NULL,
  tax_table = NULL,
  pval = 0.05,
  taxolev = "Genus",
  select_taxa = NULL,
  color_tax = "Phylum",
  tax_depth = NULL,
```

```

    verbose = TRUE,
    jitter_width = 0.1,
    ...
)

```

Arguments

data	(required) a phyloseq-class or a DESeqDataSet-class object.
contrast	(required) contrast specifies what comparison to extract from the object to build a results table. See results man page for more details.
tax_table	Required if data is a DESeqDataSet-class object. The taxonomic table used to find the taxa and color_taxa arguments. If data is a phyloseq-class object, data@tax_table is used.
pval	(default: 0.05) the significance cutoff used for optimizing the independent filtering. If the adjusted p-value cutoff (FDR) will be a value other than 0.05, pval should be set to that value.
taxolev	taxonomic level of interest
select_taxa	Either the name of the taxa (in the form of DESeq2::results()) or a logical vector (length of the results from DESeq2::results()) to select taxa to plot.
color_tax	taxonomic level used for color or a color vector.
tax_depth	Taxonomic depth to test for differential distribution among contrast. If Null the analysis is done at the OTU (i.e. Species) level. If not Null, data need to be a column name in the tax_table slot of the phyloseq-class object.
verbose	whether the function print some information during the computation
jitter_width	width for the jitter positioning
...	Additional arguments passed on to DESeq or ggplot

Details

Please cite DESeq2 package if you use this function.

Value

A [ggplot2](#) plot representing DESeq2 results

Author(s)

Adrien Taudière

See Also

[DESeq](#)
[results](#)
[plot_edgeR_pq](#)

Examples

```

data("GlobalPatterns", package = "phyloseq")
GP <- subset_taxa(GlobalPatterns, GlobalPatterns@tax_table[, 1] == "Archaea")
GP <- subset_samples(GP, SampleType %in% c("Soil", "Skin"))
if (requireNamespace("DESeq2")) {
  res <- DESeq2::DESeq(phyloseq_to_deseq2(GP, ~SampleType),
    test = "Wald", fitType = "local"
  )
  plot_deseq2_pq(res, c("SampleType", "Soil", "Skin"),
    tax_table = GP@tax_table, color_tax = "Kingdom"
  )
  plot_deseq2_pq(res, c("SampleType", "Soil", "Skin"),
    tax_table = GP@tax_table, color_tax = "Kingdom",
    pval = 0.7
  )
  plot_deseq2_pq(res, c("SampleType", "Soil", "Skin"),
    tax_table = GP@tax_table, color_tax = "Class",
    select_taxa = c("522457", "271582")
  )
}

```

plot_edgeR_pq

Plot edgeR results for a phyloseq or a edgeR object.

Description

Graphical representation of edgeR result.

Usage

```

plot_edgeR_pq(
  physeq,
  contrast = NULL,
  pval = 0.05,
  taxolev = "Genus",
  color_tax = "Phylum",
  verbose = TRUE,
  ...
)

```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

contrast (required): This argument specifies what comparison to extract from the object to build a results table. See [results](#) man page for more details.

pval	(default: 0.05): the significance cutoff used for optimizing the independent filtering. If the adjusted p-value cutoff (FDR) will be a value other than 0.05, pval should be set to that value.
taxolev	taxonomic level of interest
color_tax	taxonomic level used for color assignation
verbose	(logical): whether the function print some information during the computation
...	Additional arguments passed on to exactTest or ggplot

Value

A [ggplot2](#) plot representing edgeR results

Author(s)

Adrien Taudière

See Also

[exactTest](#)

[plot_deseq2_pq](#)

Examples

```
data("GlobalPatterns", package = "phyloseq")
GP_archae <- subset_taxa(GlobalPatterns, GlobalPatterns@tax_table[, 1] == "Archaea")

if (requireNamespace("edgeR")) {
  plot_edgeR_pq(GP_archae, c("SampleType", "Soil", "Feces"),
    color_tax = "Kingdom"
  )

  plot_edgeR_pq(GP_archae, c("SampleType", "Soil", "Feces"),
    taxolev = "Class", color_tax = "Kingdom"
  )
}
```

plot_guild_pq

Plot information about Guild from tax_table slot previously created with [add_funguild_info\(\)](#)

Description

Graphical function.

Usage

```
plot_guild_pq(physeq, levels_order = NULL, clean_pq = TRUE, ...)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

levels_order (Default NULL) A character vector to reorder the levels of guild. See examples.

clean_pq (logical, default TRUE): Does the phyloseq object is cleaned using the [clean_pq\(\)](#) function?

... Other params for be passed on to [clean_pq\(\)](#) function

Value

A ggplot2 object

Author(s)

Adrien Taudière

See Also

[add_funguild_info\(\)](#)

Examples

```
if (requireNamespace("httr")) {
  d_fung_mini <- add_funguild_info(data_fungi_mini,
    taxLevels = c(
      "Domain",
      "Phylum",
      "Class",
      "Order",
      "Family",
      "Genus",
      "Species"
    )
  )
  sort(table(d_fung_mini@tax_table[, "guild"]), decreasing = TRUE)

  p <- plot_guild_pq(d_fung_mini)
  if (requireNamespace("patchwork")) {
    (plot_guild_pq(subset_samples(d_fung_mini, Height == "Low"),
      levels_order = p$data$Guild[order(p$data$nb_seq)]
    ) + theme(legend.position = "none")) +
    (plot_guild_pq(subset_samples(d_fung_mini, Height == "High"),
      levels_order = p$data$Guild[order(p$data$nb_seq)]
    ) + ylab("")) + theme(axis.text.y = element_blank())
  }
}
```

`plot_LCBD_pq`*Plot and test local contributions to beta diversity (LCBD) of samples*

Description

A wrapper for the `adespatial::beta.div()` function in the case of physeq object.

Usage

```
plot_LCBD_pq(  
  physeq,  
  p_adjust_method = "BH",  
  pval = 0.05,  
  sam_variables = NULL,  
  only_plot_significant = TRUE,  
  ...  
)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`p_adjust_method` (chr, default "BH"): the method used to adjust p-value

`pval` (int, default 0.05): the value to determine the significance of LCBD

`sam_variables` A vector of variable names present in the `sam_data` slot to plot alongside the LCBD value

`only_plot_significant` (logical, default TRUE) Do we plot all LCBD values or only the significant ones

... Other arguments passed on to `adespatial::beta.div()` function

Details

This function is mainly a wrapper of the work of others. Please make a reference to `vegan::beta.div()` if you use this function.

Value

A `ggplot2` object build with the package `patchwork`

Author(s)

Adrien Taudière

See Also

[LCBD_pq](#), [adespatial::beta.div\(\)](#)

Examples

```

data(data_fungi)
if (requireNamespace("adespatial")) {
  plot_LCBD_pq(data_fungi_mini,
    nperm = 100, only_plot_significant = FALSE,
    pval = 0.2
  )
}

if (requireNamespace("adespatial")) {
  plot_LCBD_pq(data_fungi_mini,
    nperm = 100, only_plot_significant = TRUE,
    pval = 0.2
  )
  if (requireNamespace("patchwork")) {
    plot_LCBD_pq(data_fungi_mini,
      nperm = 100, only_plot_significant = FALSE,
      sam_variables = c("Time", "Height")
    )
    plot_LCBD_pq(data_fungi_mini,
      nperm = 100, only_plot_significant = TRUE, pval = 0.2,
      sam_variables = c("Time", "Height", "Tree_name")
    ) &
    theme(
      legend.key.size = unit(0.4, "cm"),
      legend.text = element_text(size = 10),
      axis.title.x = element_text(size = 6)
    )
  }
}

```

`plot_mt`*Plot the result of a mt test [phyloseq::mt\(\)](#)*

Description

Graphical representation of mt test.

Usage

```
plot_mt(mt = NULL, alpha = 0.05, color_tax = "Class", taxa = "Species")
```

Arguments

<code>mt</code>	(required) Result of a mt test from the function phyloseq::mt() .
<code>alpha</code>	(default: 0.05) Choose the cut off p-value to plot taxa.
<code>color_tax</code>	(default: "Class") A taxonomic level to color the points.
<code>taxa</code>	(default: "Species") The taxonomic level you choose for x-positioning.

Value

a [ggplot2](#) plot of result of a mt test

Author(s)

Adrien Taudière

See Also

[phyloseq::mt\(\)](#)

Examples

```
# Filter samples that don't have Time
data_fungi_mini2 <- subset_samples(data_fungi_mini, !is.na(Time))
res <- mt(data_fungi_mini2, "Time", method = "fdr", test = "f", B = 300)
plot_mt(res)
plot_mt(res, taxa = "Genus", color_tax = "Order")
```

plot_SCBD_pq

Plot species contributions to beta diversity (SCBD) of samples

Description

A wrapper for the [adespatial::beta.div\(\)](#) function in the case of physeq object.

Usage

```
plot_SCBD_pq(
  physeq,
  tax_level = "ASV",
  tax_col = "Order",
  min_SCBD = 0.01,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
tax_level	Taxonomic level to used in y axis
tax_col	Taxonomic level to colored points
min_SCBD	(default 0.01) the minimum SCBD value to plot the taxa
...	Other arguments passed on to adespatial::beta.div() function

Details

This function is mainly a wrapper of the work of others. Please make a reference to `vegan::beta.div()` if you use this function.

Value

A `ggplot2` object build with the package `patchwork`

Author(s)

Adrien Taudière

See Also

[LCBD_pq](#), [adespatial::beta.div\(\)](#)

Examples

```
data(data_fungi)
if (requireNamespace("adespatial")) {
  plot_SCBD_pq(data_fungi) +
    geom_text(aes(label = paste(Genus, Species)), hjust = 1, vjust = 2) +
    xlim(c(0, NA))
}

if (requireNamespace("adespatial")) {
  plot_SCBD_pq(data_fungi, tax_level = "Class", tax_col = "Phylum", min_SCBD = 0) +
    geom_jitter()
}
```

plot_tax_pq

Plot taxonomic distribution in function of a factor with stacked bar in %

Description

An alternative to `phyloseq::plot_bar()` function.

Usage

```
plot_tax_pq(
  physeq,
  fact = NULL,
  merge_sample_by = NULL,
  type = "nb_seq",
  taxa_fill = "Order",
```

```

    print_values = TRUE,
    color_border = "lightgrey",
    linewidth = 0.1,
    prop_print_value = 0.01,
    nb_print_value = NULL,
    add_info = TRUE,
    na_remove = TRUE,
    clean_pq = TRUE
  )

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
merge_sample_by	a vector to determine which samples to merge using the merge_samples2() function. Need to be in physeq@sam_data
type	If "nb_seq" (default), the number of sequences is used in plot. If "nb_asv", the number of ASV is plotted. If both, return a list of two plots, one for nbSeq and one for ASV.
taxa_fill	(default: 'Order'): Name of the taxonomic rank of interest
print_values	(logical, default TRUE): Do we print some values on plot?
color_border	color for the border
linewidth	The line width of geom_bar
prop_print_value	minimal proportion to print value (default 0.01)
nb_print_value	number of higher values to print (replace prop_print_value if both are set).
add_info	(logical, default TRUE) Do we add title and subtitle with information about the total number of sequences and the number of samples per modality.
na_remove	(logical, default TRUE) if TRUE remove all the samples with NA in the split_by variable of the physeq@sam_data slot
clean_pq	(logical) If set to TRUE, empty samples are discarded after subsetting ASV

Value

A ggplot2 graphic

Author(s)

Adrien Taudière

See Also

[tax_bar_pq\(\)](#) and [multitax_bar_pq\(\)](#)

Examples

```

data(data_fungi_sp_known)
plot_tax_pq(data_fungi_sp_known,
  "Time",
  merge_sample_by = "Time",
  taxa_fill = "Class"
)

plot_tax_pq(data_fungi_sp_known,
  "Height",
  merge_sample_by = "Height",
  taxa_fill = "Class",
  na_remove = TRUE,
  color_border = rgb(0, 0, 0, 0)
)

plot_tax_pq(data_fungi_sp_known,
  "Height",
  merge_sample_by = "Height",
  taxa_fill = "Class",
  na_remove = FALSE,
  clean_pq = FALSE
)

```

plot_tsne_pq

Plot a tsne low dimensional representation of a phyloseq object

Description

Partially inspired by `phylosmith::tsne_phyloseq()` function developed by Schuyler D. Smith.

Usage

```

plot_tsne_pq(
  physeq,
  method = "bray",
  dims = 2,
  theta = 0,
  perplexity = 30,
  fact = NA,
  ellipse_level = 0.95,
  plot_dims = c(1, 2),
  na_remove = TRUE,
  force_factor = TRUE,
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
method	A method to calculate distance using <code>vegan::vegdist()</code> function (default: "bray")
dims	(Int) Output dimensionality (default: 2)
theta	(Numeric) Speed/accuracy trade-off (increase for less accuracy), set to 0.0 for exact TSNE (default: 0.0 see details in the man page of <code>Rtsne::Rtsne</code>).
perplexity	(Numeric) Perplexity parameter (should not be bigger than $3 * \text{perplexity} < \text{nrow}(X) - 1$, see details in the man page of <code>Rtsne::Rtsne</code>)
fact	Name of the column in <code>physeq@sam_data</code> used to color points and compute ellipses.
ellipse_level	The level used in <code>stat_ellipse</code> . Set to NULL to discard ellipse (default = 0.95)
plot_dims	A vector of 2 values defining the rank of dimension to plot (default: <code>c(1,2)</code>)
na_remove	(logical, default TRUE) Does the samples with NA values in fact are removed? (default: true)
force_factor	(logical, default TRUE) Force the fact column to be a factor.
...	Other arguments passed on to <code>Rtsne::Rtsne()</code>

Value

A ggplot object

Author(s)

Adrien Taudière

Examples

```
data(data_fungi)
if (requireNamespace("Rtsne")) {
  plot_tsne_pq(data_fungi, fact = "Height", perplexity = 15)
}

if (requireNamespace("Rtsne")) {
  plot_tsne_pq(data_fungi, fact = "Time") + geom_label(aes(label = Sample_id, fill = Time))
  plot_tsne_pq(data_fungi, fact = "Time", na_remove = FALSE, force_factor = FALSE)
}
```

plot_var_part_pq *Plot the partition the variation of a phyloseq object*

Description

Graphical representation of the partition of variation obtain with `var_par_pq()`.

Usage

```
plot_var_part_pq(
  res_varpart,
  cutoff = 0,
  digits = 1,
  digits_quantile = 2,
  fill_bg = c("seagreen3", "mediumpurple", "blue", "orange"),
  show_quantiles = FALSE,
  filter_quantile_zero = TRUE,
  show_dbrda_signif = FALSE,
  show_dbrda_signif_pval = 0.05,
  alpha = 63,
  id.size = 1.2,
  min_prop_pval_signif_dbrda = 0.95
)
```

Arguments

<code>res_varpart</code>	(required) the result of the functions <code>var_par_pq()</code> or <code>var_par_rarperm_pq()</code>
<code>cutoff</code>	The values below cutoff will not be displayed.
<code>digits</code>	The number of significant digits.
<code>digits_quantile</code>	The number of significant digits for quantile.
<code>fill_bg</code>	Fill colours of ellipses.
<code>show_quantiles</code>	Do quantiles are printed ?
<code>filter_quantile_zero</code>	Do we filter out value with quantile encompassing the zero value?
<code>show_dbrda_signif</code>	Do dbrda significance for each component is printed using *?
<code>show_dbrda_signif_pval</code>	(float, [0:1]) The value under which the dbrda is considered significant.
<code>alpha</code>	(int, [0:255]) Transparency of the fill colour.
<code>id.size</code>	A numerical value giving the character expansion factor for the names of circles or ellipses.
<code>min_prop_pval_signif_dbrda</code>	(float, [0:1]) Only used if using the result of <code>var_par_rarperm_pq()</code> function. The * for <code>dbrda_signif</code> is only add if at least <code>min_prop_pval_signif_dbrda</code> of permutations show significance.

Details

This function is mainly a wrapper of the work of others. Please make a reference to `vegan::varpart()` if you use this function.

Value

A plot

Author(s)

Adrien Taudière

See Also

[var_par_rarperm_pq\(\)](#), [var_par_pq\(\)](#)

Examples

```
if (requireNamespace("vegan")) {
  data_fungi_woNA <- subset_samples(data_fungi, !is.na(Time) & !is.na(Height))
  res_var_9 <- var_par_rarperm_pq(
    data_fungi_woNA,
    list_component = list(
      "Time" = c("Time"),
      "Size" = c("Height", "Diameter")
    ),
    nperm = 9,
    dbrda_computation = TRUE
  )
  res_var_2 <- var_par_rarperm_pq(
    data_fungi_woNA,
    list_component = list(
      "Time" = c("Time"),
      "Size" = c("Height", "Diameter")
    ),
    nperm = 2,
    dbrda_computation = TRUE
  )
  res_var0 <- var_par_pq(data_fungi_woNA,
    list_component = list(
      "Time" = c("Time"),
      "Size" = c("Height", "Diameter")
    ),
    dbrda_computation = TRUE
  )
  plot_var_part_pq(res_var0, digits_quantile = 2, show_dbrda_signif = TRUE)
  plot_var_part_pq(res_var_9,
    digits_quantile = 2, show_quantiles = TRUE,
    show_dbrda_signif = TRUE
  )
  plot_var_part_pq(
    res_var_2,
```

```

    digits = 5,
    digits_quantile = 2,
    cutoff = 0,
    show_quantiles = TRUE
  )
}

```

psmelt_samples_pq *Build a sample information tibble from physeq object*

Description

Hill numbers are the number of equiprobable species giving the same diversity value as the observed distribution.

Note that contrary to [hill_pq\(\)](#), this function does not take into account for difference in the number of sequences per samples/modalities. You may use `rarefy_by_sample = TRUE` if the mean number of sequences per samples differs among modalities.

Usage

```

psmelt_samples_pq(
  physeq,
  hill_scales = c(0, 1, 2),
  filter_zero = TRUE,
  rarefy_by_sample = FALSE,
  taxa_ranks = NULL
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
hill_scales	(a vector of integer) The list of q values to compute the hill number H^q . If Null, no hill number are computed. Default value compute the Hill number 0 (Species richness), the Hill number 1 (exponential of Shannon Index) and the Hill number 2 (inverse of Simpson Index).
filter_zero	(logical, default TRUE) Do we filter non present OTU from samples ? For the moment, this has no effect on the result because the dataframe is grouped by samples with abundance summed across OTU.
rarefy_by_sample	(logical, default FALSE) If TRUE, rarefy samples using phyloseq::rarefy_even_depth() function.
taxa_ranks	A vector of taxonomic ranks. For examples <code>c("Family","Genus")</code> . If taxa ranks is not set (default value = NULL), taxonomic information are not present in the resulting tibble.

Value

A tibble with a row for each sample. Columns provide information from `sam_data` slot as well as hill numbers, Abundance (nb of sequences), and `Abundance_log10` ($\log_{10}(1+Abundance)$).

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("ggstatsplot")) {
  psm_tib <- psmelt_samples_pq(data_fungi_mini, hill_scales = c(0, 2, 7))
  ggstatsplot::ggbetweenstats(psm_tib, Height, Hill_0)
  ggstatsplot::ggbetweenstats(psm_tib, Height, Hill_7)

  psm_tib_tax <- psmelt_samples_pq(data_fungi_mini, taxa_ranks = c("Class", "Family"))
  ggplot(filter(psm_tib_tax, Abundance > 2000), aes(y = Family, x = Abundance, fill = Time)) +
    geom_bar(stat = "identity") +
    facet_wrap(~Height)
}
```

`rarefy_sample_count_by_modality`

Rarefy (equalize) the number of samples per modality of a factor

Description

This function randomly draw the same number of samples for each modality of factor. It is usefull to disentangle the effect of different number of samples per modality on diversity. Internally used in [accu_plot_balanced_modality\(\)](#).

Usage

```
rarefy_sample_count_by_modality(physeq, fact, rngseed = FALSE, verbose = TRUE)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the phyloseq package.
<code>fact</code>	(required): The variable to rarefy. Must be present in the <code>sam_data</code> slot of the physeq object.
<code>rngseed</code>	(Optional). A single integer value passed to <code>set.seed</code> , which is used to fix a seed for reproducibly random number generation (in this case, reproducibly random subsampling). If set to <code>FALSE</code> , then no iddling with the RNG seed is performed, and it is up to the user to appropriately call
<code>verbose</code>	(logical). If <code>TRUE</code> , print additional informations.

Value

A new [phyloseq-class](#) object.

Author(s)

Adrien Taudière

See Also

[accu_plot_balanced_modality\(\)](#)

Examples

```
table(data_fungi_mini@sam_data$Height)
data_fungi_mini2 <- rarefy_sample_count_by_modality(data_fungi_mini, "Height")
table(data_fungi_mini2@sam_data$Height)
if (requireNamespace("patchwork")) {
  ggvenn_pq(data_fungi_mini, "Height") + ggvenn_pq(data_fungi_mini2, "Height")
}
```

read_pq

Read phyloseq object from multiple csv tables and a phylogenetic tree in Newick format.

Description

This is the reverse function of [write_pq\(\)](#).

Usage

```
read_pq(
  path = NULL,
  taxa_are_rows = FALSE,
  sam_names = NULL,
  sep_csv = "\t",
  ...
)
```

Arguments

path	(required) a path to the folder to read the phyloseq object
taxa_are_rows	(default to FALSE) see ?phyloseq for details
sam_names	The name of the variable (column) in sam_data.csv to rename samples. Note that if you use write_phyloseq() function to save your physeq object, you may use sam_names = "X" to rename the samples names as before.
sep_csv	(default tabulation) separator for column
...	Other arguments passed on to utils::write.table() function.

Value

One to four csv tables (refseq.csv, otu_table.csv, tax_table.csv, sam_data.csv) and if present a phy_tree in Newick format. At least the otu_table.csv need to be present.

Examples

```
write_pq(data_fungi, path = paste0(tempdir(), "/phyloseq"))
read_pq(path = paste0(tempdir(), "/phyloseq"))
unlink(paste0(tempdir(), "/phyloseq"), recursive = TRUE)
```

rename_samples	<i>Rename the samples of a phyloseq slot</i>
----------------	--

Description

Useful for targets bioinformatic pipeline.

Usage

```
rename_samples(phyloseq_component, names_of_samples, taxa_are_rows = FALSE)
```

Arguments

phyloseq_component
(required) one of otu_table or sam_data slot of a phyloseq-class object

names_of_samples
(required) A vector of samples names

taxa_are_rows (default to FALSE) see ?phyloseq for details

Value

The otu_table or the sam_data slot with new samples names

Author(s)

Adrien Taudière

Examples

```
otutab <- rename_samples(
  data_fungi@otu_table,
  paste0("data_f", sample_names(data_fungi))
)
otutab2 <- rename_samples(
  clean_pq(data_fungi,
    force_taxa_as_rows = TRUE
  )@otu_table,
  paste0("data_f", sample_names(data_fungi))
)
```

```
)  
sanda <- rename_samples(  
  data_fungi@sam_data,  
  paste0("data_f", sample_names(data_fungi))  
)
```

rename_samples_otu_table

Rename samples of an otu_table

Description

Useful for targets bioinformatic pipeline.

Usage

```
rename_samples_otu_table(physeq, names_of_samples)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.
names_of_samples (required) The new names of the samples

Value

the matrix with new colnames (or rownames if taxa_are_rows is true)

Author(s)

Adrien Taudière

Examples

```
rename_samples_otu_table(data_fungi, as.character(seq_along(sample_names(data_fungi))))
```

reorder_taxa_pq	<i>Reorder taxa in otu_table/tax_table/refseq slot of a phyloseq object</i>
-----------------	---

Description

Note that the taxa order in a phyloseq object with a tree is locked by the order of leaf in the phylogenetic tree.

Usage

```
reorder_taxa_pq(phyloseq, names_ordered, remove_phy_tree = FALSE)
```

Arguments

phyloseq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

names_ordered (required): Names of the taxa (must be the same as taxa in `taxa_names(phyloseq)`) in a given order

remove_phy_tree (logical, default FALSE) If TRUE, the phylogenetic tree is removed. It is

Value

A phyloseq object

Author(s)

Adrien Taudière

Examples

```
data_fungi_ordered_by_genus <- reorder_taxa_pq(  
  data_fungi,  
  taxa_names(data_fungi)[order(as.vector(data_fungi@tax_table[, "Genus"]))]  
)  
  
data_fungi_mini_asc_ordered_by_abundance <- reorder_taxa_pq(  
  data_fungi_mini,  
  taxa_names(data_fungi_mini)[order(taxa_sums(data_fungi_mini))]  
)
```

ridges_pq

*Ridge plot of a phyloseq object***Description**

Graphical representation of distribution of taxa across a factor using ridges.

Usage

```
ridges_pq(
  physeq,
  fact,
  nb_seq = TRUE,
  log10trans = TRUE,
  tax_level = "Class",
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required) Name of the factor in physeq@sam_data used to plot different lines
nb_seq	(logical; default TRUE) If set to FALSE, only the number of ASV is count. Concretely, physeq otu_table is transformed in a binary otu_table (each value different from zero is set to one)
log10trans	(logical, default TRUE) If TRUE, the number of sequences (or ASV if nb_seq = FALSE) is log10 transformed.
tax_level	The taxonomic level to fill ridges
...	Other params passed on to ggridges::geom_density_ridges()

Value

A [ggplot2](#) plot with bar representing the number of sequence en each taxonomic groups

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("ggridges")) {
  ridges_pq(data_fungi_mini, "Time", alpha = 0.5, log10trans = FALSE) + xlim(c(0, 1000))
}

if (requireNamespace("ggridges")) {
  ridges_pq(data_fungi_mini, "Time", alpha = 0.5, scale = 0.9)
  ridges_pq(data_fungi_mini, "Sample_names", log10trans = TRUE) + facet_wrap("~Height")
}
```



```

ridges_pq(data_fungi_mini,
  "Time",
  jittered_points = TRUE,
  position = ggriidges::position_points_jitter(width = 0.05, height = 0),
  point_shape = "|", point_size = 3, point_alpha = 1, alpha = 0.7,
  scale = 0.8
)
}

```

rotl_pq

*rotl wrapper for phyloseq data***Description**

Make a phylogenetic tree using the ASV names of a physeq object and the Open Tree of Life tree.

Usage

```
rotl_pq(physeq, species_colnames = "Genus_species", context_name = "All life")
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`species_colnames` (default: "Genus_species"): the name of the column where the species binominal name is stored in @tax_table slot. Can also be a vector of two column names e.g. c("Genus", "Species")

`context_name` : can be used to select only a part of the Open Tree of Life. See `?rotl::tnrs_contexts()` for available values

Details

This function is mainly a wrapper of the work of others. Please make a reference to rotl package if you use this function.

Value

A plot

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("rotl")) {  
  tr <- rotl_pq(data_fungi_mini, species_colnames = "Genus_species")  
  plot(tr)  
  
  tr_ASCO <- rotl_pq(data_fungi, species_colnames = "Genus_species", context_name = "Ascomycetes")  
  plot(tr_ASCO)  
}
```

sample_data_with_new_names

Load sample data from file and rename samples using names of samples and an optional order

Description

Useful for targets bioinformatic pipeline.

Usage

```
sample_data_with_new_names(  
  file_path,  
  names_of_samples,  
  samples_order = NULL,  
  ...  
)
```

Arguments

`file_path` (required) a path to the sample_data file
`names_of_samples` (required) a vector of sample names
`samples_order` Optional numeric vector to sort sample names
... Other arguments passed on to `utils::read.delim()` function.

Value

A data.frame from `file_path` and new names

Author(s)

Adrien Taudière

See Also[rename_samples\(\)](#)**Examples**

```
sam_file <- system.file("extdata", "sam_data.csv", package = "MiscMetabar")
sample_data_with_new_names(sam_file, paste0("Samples_", seq(1, 185)))
```

sankey_pq *Sankey plot of [phyloseq-class](#) object*

Description

Graphical representation of distribution of taxa across Taxonomy and (optionnaly a factor).

Usage

```
sankey_pq(
  physeq = NULL,
  fact = NULL,
  taxa = 1:4,
  add_nb_seq = FALSE,
  min_prop_tax = 0,
  tax2remove = NULL,
  units = NULL,
  symbol2sub = c("\\.", "-"),
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
taxa	a vector of taxonomic rank to plot
add_nb_seq	Represent the number of sequences or the number of OTUs (add_nb_seq = FALSE). Note that plotting the number of sequences is slower.
min_prop_tax	(default: 0) The minimum proportion for taxon to be plotted. EXPERIMENTAL. For the moment each links below the min.prop. tax is discard from the sankey network resulting in sometimes weird plot.
tax2remove	a vector of taxonomic groups to remove from the analysis (e.g. c('Incertae sedis', 'unidentified'))
units	character string describing physical units (if any) for Value
symbol2sub	(default: c('\.', '-')) vector of symbol to delete in the taxonomy
...	Additional arguments passed on to sankeyNetwork

Value

A [sankeyNetwork](#) plot representing the taxonomic distribution of OTUs or sequences. If `fact` is set, represent the distribution of the last taxonomic level in the modalities of `fact`

Author(s)

Adrien Taudière

See Also

[sankeyNetwork](#), [ggaluv_pq\(\)](#)

Examples

```
data("GlobalPatterns", package = "phyloseq")
GP <- subset_taxa(GlobalPatterns, GlobalPatterns@tax_table[, 1] == "Archaea")
if (requireNamespace("networkD3")) {
  sankey_pq(GP, fact = "SampleType")
}

if (requireNamespace("networkD3")) {
  sankey_pq(GP, taxa = 1:4, min_prop_tax = 0.01)
  sankey_pq(GP, taxa = 1:4, min_prop_tax = 0.01, add_nb_seq = TRUE)
}
```

save_pq

A wrapper of write_pq to save in all three possible formats

Description

A wrapper of `write_pq` to save in all three possible formats

Usage

```
save_pq(physeq, path = NULL, ...)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the `phyloseq` package.

`path` a path to the folder to save the `phyloseq` object

`...` Other arguments passed on to [write_pq\(\)](#) or [utils::write.table\(\)](#) function.

Details

Write :

- 4 separate tables
- 1 table version
- 1 RData file

Value

Build a folder (in path) with four csv tables (refseq.csv, otu_table.csv, tax_table.csv, sam_data.csv) + one table with all tables together + a rdata file (physeq.RData) that can be loaded using `base::load()` function + if present a phylogenetic tree in Newick format (phy_tree.txt)

Author(s)

Adrien Taudière

See Also

[write_pq\(\)](#)

Examples

```
save_pq(data_fungi, path = paste0(tempdir(), "/phyloseq"))
unlink(paste0(tempdir(), "/phyloseq"), recursive = TRUE)
```

search_exact_seq_pq *Search for exact matching of sequences*

Description

Search for exact matching of sequences using complement, reverse and reverse-complement

Usage

```
search_exact_seq_pq(physeq, seq2search)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.
seq2search A DNASTringSet object of sequences to search for.

Value

A list of data-frames for each input sequences with the name, the sequences and the number of occurrences of the original sequence, the complement sequence, the reverse sequence and the reverse-complement sequence.

Author(s)

Adrien Taudière

Examples

```
data("data_fungi")
search_primers <- search_exact_seq_pq(data_fungi,
  seq2search = Biostrings::DNAStringSet(c("TTGAACGCACATTGCGCC", "ATCCCTACCTGATCCGAG"))
)
```

select_one_sample *Select one sample from a physeq object*

Description

Mostly for internal used, for example in function `track_wkflow_samples()`.

Usage

```
select_one_sample(physeq, sam_name, silent = FALSE)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.
sam_name (required) The sample name to select
silent (logical) If true, no message are printing.

Value

A new [phyloseq-class](#) object with one sample

Author(s)

Adrien Taudière

Examples

```
A8_005 <- select_one_sample(data_fungi, "A8-005_S4_MERGED.fastq.gz")
A8_005
```

select_taxa	<i>Select a subset of taxa in a specified order where possible</i>
-------------	--

Description

Select (a subset of) taxa; if x allows taxa to be reordered, then taxa are given in the specified order.

Usage

```
select_taxa(x, taxa, reorder = TRUE)

## S4 method for signature 'sample_data,character'
select_taxa(x, taxa)

## S4 method for signature 'otu_table,character'
select_taxa(x, taxa, reorder = TRUE)

## S4 method for signature 'taxonomyTable,character'
select_taxa(x, taxa, reorder = TRUE)

## S4 method for signature 'XStringSet,character'
select_taxa(x, taxa, reorder = TRUE)

## S4 method for signature 'phylo,character'
select_taxa(x, taxa)

## S4 method for signature 'phyloseq,character'
select_taxa(x, taxa, reorder = TRUE)
```

Arguments

x	A phyloseq object or phyloseq component object
taxa	Character vector of taxa to select, in requested order
reorder	Logical specifying whether to use the order in taxa (TRUE) or keep the order in taxa_names(x) (FALSE)

Details

This is a simple selector function that is like `prune_taxa(taxa, x)` when `taxa` is a character vector but always gives the taxa in the order `taxa` if possible (that is, except for `phy_tree`'s and `phyloseq` objects that contain `phy_tree`'s).

Author(s)

Michael R. McLaren (orcid: [0000-0003-1575-473X](https://orcid.org/0000-0003-1575-473X))

signif_ancombc	<i>Filter ancombc_pq results</i>
----------------	----------------------------------

Description

Internally used in [plot_ancombc_pq\(\)](#).

Usage

```
signif_ancombc(  
  ancombc_res,  
  filter_passed = TRUE,  
  filter_diff = TRUE,  
  min_abs_lfc = 0  
)
```

Arguments

ancombc_res	(required) the result of the ancombc_pq function For the moment only bimodal factors are possible.
filter_passed	(logical, default TRUE) Do we filter using the column passed_ss? The passed_ss value is TRUE if the taxon passed the sensitivity analysis, i.e., adding different pseudo-counts to 0s would not change the results.
filter_diff	(logical, default TRUE) Do we filter using the column diff? The diff value is TRUE if the taxon is significant (has q less than alpha)
min_abs_lfc	(integer, default 0) Minimum absolute value to filter results based on Log Fold Change. For ex. a value of 1 filter out taxa for which the abundance in a given level of the modality is not at least the double of the abundance in the other level.

Details

This function is mainly a wrapper of the work of others. Please make a reference to ANCOMBC: : ancombc2() if you use this function.

Value

A data.frame with the same number of columns than the ancombc_res param but with less (or equal) numbers of rows

See Also

[ancombc_pq\(\)](#), [plot_ancombc_pq\(\)](#)

Examples

```
if (requireNamespace("mia")) {
  data_fungi_mini@tax_table <- phyloseq::tax_table(cbind(
    data_fungi_mini@tax_table,
    "taxon" = taxa_names(data_fungi_mini)
  ))

  res_time <- ancombc_pq(
    data_fungi_mini,
    fact = "Time",
    levels_fact = c("0", "15"),
    tax_level = "taxon",
    verbose = TRUE
  )

  signif_ancombc(res_time)
}
```

simplify_taxo

Simplify taxonomy by removing some unused characters such as "k_"

Description

Internally used in [clean_pq\(\)](#)

Usage

```
simplify_taxo(physeq, remove_space = TRUE)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.
remove_space (logical; default TRUE): do we remove space?

Value

A [phyloseq-class](#) object with simplified taxonomy

Author(s)

Adrien Taudière

SRS_curve_pq	<i>Scaling with ranked subsampling (SRS) curve of phyloseq object</i>
--------------	---

Description

A wrapper of `SRS::SRScurve()` function.

Usage

```
SRS_curve_pq(physeq, clean_pq = FALSE, ...)
```

Arguments

physeq	(required): a <code>phyloseq-class</code> object obtained using the phyloseq package.
clean_pq	(logical): Does the phyloseq object is cleaned using the <code>clean_pq()</code> function?
...	Other arguments passed on to <code>SRS::SRScurve()</code>

Value

A plot

Examples

```
if (requireNamespace("SRS")) {
  SRS_curve_pq(data_fungi_mini,
    max.sample.size = 200,
    rarefy.comparison = TRUE, rarefy.repeats = 3
  )
  SRS_curve_pq(data_fungi_mini, max.sample.size = 500, metric = "shannon")
}
```

subsample_fastq	<i>Subsample a fastq file copying the n_seq first sequences in a given folder</i>
-----------------	---

Description

Useful to test a pipeline on small fastq files.

Usage

```
subsample_fastq(fastq_files, folder_output = "subsample", nb_seq = 1000)
```

Arguments

fastq_files The path to one fastq file or a list of fastq files (see examples)
 folder_output The path to a folder for output files
 nb_seq (int; default 1000) : Number of sequences kept (every sequence spread across 4 lines)

Value

Nothing, create subsampled fastq files in a folder

Author(s)

Adrien Taudière

Examples

```
ex_file <- system.file("extdata", "ex_R1_001.fastq.gz",
  package = "MiscMetabar",
  mustWork = TRUE
)
subsample_fastq(ex_file, paste0(tempdir(), "/output_fastq"))
subsample_fastq(list_fastq_files("extdata"), paste0(tempdir(), "/output_fastq"), n = 10)
unlink(paste0(tempdir(), "/output_fastq"), recursive = TRUE)
```

subset_samples_pq *Subset samples using a conditional boolean vector.*

Description

The main objective of this function is to complete the `phyloseq::subset_samples()` function by propose a more easy (but more prone to error) way of `subset_samples`. It replace the subsetting expression which used the name of the variable in the `sam_data` by a boolean vector.

Warnings: you must verify the result of this function as the boolean condition must match the order of samples in the `sam_data` slot.

This function is robust when you use the `sam_data` slot of the `phyloseq` object used in `physeq` (see examples)

Usage

```
subset_samples_pq(physeq, condition)
```

Arguments

physeq (required): a `phyloseq-class` object obtained using the `phyloseq` package.
 condition A boolean vector to subset samples. Length must fit the number of samples

Value

a new phyloseq object

Examples

```
cond_samp <- grepl("A1", data_fungi@sam_data[["Sample_names"]])
subset_samples_pq(data_fungi, cond_samp)

subset_samples_pq(data_fungi, data_fungi@sam_data[["Height"]] == "Low")
```

subset_taxa_pq	<i>Subset taxa using a conditional named boolean vector.</i>
----------------	--

Description

The main objective of this function is to complete the `phyloseq::subset_taxa()` function by propose a more easy way of subset_taxa using a named boolean vector. Names must match taxa_names.

Usage

```
subset_taxa_pq(
  physeq,
  condition,
  verbose = TRUE,
  clean_pq = TRUE,
  taxa_names_from_physeq = FALSE
)
```

Arguments

physeq	(required): a <code>phyloseq-class</code> object obtained using the phyloseq package.
condition	A named boolean vector to subset taxa. Length must fit the number of taxa and names must match taxa_names. Can also be a condition using a column of the tax_table slot (see examples). If the order of condition is the same as taxa_names(physeq), you can use the parameter taxa_names_from_physeq = TRUE.
verbose	(logical) Informations are printed
clean_pq	(logical) If set to TRUE, empty samples are discarded after subsetting ASV
taxa_names_from_physeq	(logical) If set to TRUE, rename the condition vector using taxa_names(physeq). Carefully check the result of this function if you use this parameter. No effect if the condition is of class tax_table.

Value

a new phyloseq object

Examples

```
subset_taxa_pq(data_fungi, data_fungi@tax_table[, "Phylum"] == "Ascomycota")

cond_taxa <- grepl("Endophyte", data_fungi@tax_table[, "Guild"])
names(cond_taxa) <- taxa_names(data_fungi)
subset_taxa_pq(data_fungi, cond_taxa)

subset_taxa_pq(data_fungi, grepl("mycor", data_fungi@tax_table[, "Guild"]),
  taxa_names_from_physeq = TRUE
)
```

```
subset_taxa_tax_control
```

Subset taxa using a taxa control or distribution based method

Description

There is 3 main methods : discard taxa (i) using a control taxa (e.g. truffle root tips), (ii) using a mixture models to detect bimodality in pseudo-abundance distribution or (iii) using a minimum difference threshold pseudo-abundance. Each cutoff is defined at the sample level.

Usage

```
subset_taxa_tax_control(
  physeq,
  taxa_distri,
  method = "mean",
  min_diff_for_cutoff = NULL
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
taxa_distri	(required) a vector of length equal to the number of samples with the number of sequences per sample for the taxa control
method	(default: "mean") a method to calculate the cut-off value. There are 6 available methods: <ol style="list-style-type: none"> 1. cutoff_seq: discard taxa with less than the number of sequence than taxa control, 2. cutoff_mixt: using mixture models, 3. cutoff_diff: using a minimum difference threshold (need the argument min_diff_for_cutoff) 4. min: the minimum of the three firsts methods 5. max: the maximum of the three firsts methods 6. mean: the mean of the three firsts methods

min_diff_for_cutoff

(int) argument for method cutoff_diff. Required if method is cutoff_diff, min, max or mean

Value

A new [phyloseq-class](#) object.

Author(s)

Adrien Taudière

Examples

```
subset_taxa_tax_control(data_fungi,
  as.numeric(data_fungi@otu_table[, 300]),
  min_diff_for_cutoff = 2
)
```

summary_plot_pq

Summarize a [phyloseq-class](#) object using a plot.

Description

Graphical representation of a phyloseq object.

Usage

```
summary_plot_pq(
  physeq,
  add_info = TRUE,
  min_seq_samples = 500,
  clean_pq = TRUE
)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

add_info Does the bottom down corner contain extra informations?

min_seq_samples (int): Used only when add_info is set to true to print the number of samples with less sequences than this number.

clean_pq (logical): Does the phyloseq object is cleaned using the [clean_pq\(\)](#) function?

Value

A ggplot2 object

Examples

```
summary_plot_pq(data_fungi)
summary_plot_pq(data_fungi, add_info = FALSE) + scale_fill_viridis_d()
```

swarm_clustering	<i>Re-cluster sequences of an object of class phyloseq or cluster a list of DNA sequences using SWARM</i>
------------------	---

Description

A wrapper of SWARM software.

Usage

```
swarm_clustering(
  physeq = NULL,
  dna_seq = NULL,
  d = 1,
  swarmpath = "swarm",
  vsearch_path = "vsearch",
  nproc = 1,
  swarm_args = "--fastidious",
  tax_adjust = 0,
  keep_temporary_files = FALSE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
dna_seq	NOT WORKING FOR THE MOMENT You may directly use a character vector of DNA sequences in place of physeq args. When physeq is set, dna sequences take the value of physeq@refseq
d	(default: 1) maximum number of differences allowed between two amplicons, meaning that two amplicons will be grouped if they have d (or less) differences
swarmpath	(default: swarm) path to swarm
vsearch_path	(default: vsearch) path to vsearch, used only if physeq is NULL and dna_seq is provided.
nproc	(default: 1) Set to number of cpus/processors to use for the clustering
swarm_args	(default : "-fastidious") a one length character element defining other parameters to passed on to swarm See other possible methods in the SWARM pdf manual
tax_adjust	(Default 0) See the man page of merge_taxa_vec() for more details. To conserved the taxonomic rank of the most abundant ASV,
keep_temporary_files	(logical, default: FALSE) Do we keep temporary files ?

- temp.fasta (refseq in fasta or dna_seq sequences)
- temp_output (classical output of SWARM)
- temp_uclust (clusters output of SWARM)

Details

This function use the `merge_taxa_vec` function to merge taxa into clusters. By default `tax_adjust = 0`. See the man page of `merge_taxa_vec()`.

This function is mainly a wrapper of the work of others. Please cite **SWARM**.

Value

A new object of class `physeq` or a list of cluster if `dna_seq` args was used.

References

SWARM can be downloaded from <https://github.com/torognes/swarm/>.

SWARM can be downloaded from <https://github.com/torognes/swarm>. More information in the associated publications [doi:10.1093/bioinformatics/btab493](https://doi.org/10.1093/bioinformatics/btab493) and [doi:10.7717/peerj.593](https://doi.org/10.7717/peerj.593)

See Also

[asv2otu\(\)](#), [vsearch_clustering\(\)](#)

Examples

```
summary_plot_pq(data_fungi)
system2("swarm", "-h")

data_fungi_swarm <- swarm_clustering(data_fungi)
summary_plot_pq(data_fungi_swarm)

sequences_ex <- c(
  "TACCTATGTTGCCTTGGCGGCTAAACCTACCCGGGATTTGATGGGGCGAATTAATAACGAATTCATTGAATCA",
  "TACCTATGTTGCCTTGGCGGCTAAACCTACCCGGGATTTGATGGGGCGAATTACCTGGTAAGGCCCACTT",
  "TACCTATGTTGCCTTGGCGGCTAAACCTACCCGGGATTTGATGGGGCGAATTACCTGGTAGAGGTG",
  "TACCTATGTTGCCTTGGCGGCTAAACCTACC",
  "CGGGATTTGATGGCGAATTACCTGGTATTTAGCCCACTTACCCGGTACCATGAGGTG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACCTGG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACAAAG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACAAAG",
  "GCGGCTAAACCTACCCGGGATTTGATGGCGAATTACAAAG"
)

sequences_ex_swarm <- swarm_clustering(
  dna_seq = sequences_ex
)
```

taxa_as_columns	<i>Force taxa to be in columns in the otu_table of a physeq object</i>
-----------------	--

Description

Mainly for internal use. It is a special case of clean_pq function.

Usage

```
taxa_as_columns(physeq)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

Value

A new [phyloseq-class](#) object

Author(s)

Adrien Taudière

taxa_as_rows	<i>Force taxa to be in columns in the otu_table of a physeq object</i>
--------------	--

Description

Mainly for internal use. It is a special case of clean_pq function.

Usage

```
taxa_as_rows(physeq)
```

Arguments

physeq (required): a [phyloseq-class](#) object obtained using the phyloseq package.

Value

A new [phyloseq-class](#) object

Author(s)

Adrien Taudière

`taxa_only_in_one_level`*Show taxa which are present in only one given level of a modality*

Description

Given one modality name in `sam_data` and one level of the modality, return the taxa strictly specific of this level.

Usage

```
taxa_only_in_one_level(  
  physeq,  
  modality,  
  level,  
  min_nb_seq_taxa = 0,  
  min_nb_samples_taxa = 0  
)
```

```
taxa_only_in_one_level(  
  physeq,  
  modality,  
  level,  
  min_nb_seq_taxa = 0,  
  min_nb_samples_taxa = 0  
)
```

Arguments

<code>physeq</code>	(required): a phyloseq-class object obtained using the phyloseq package.
<code>modality</code>	(required) The name of a column present in the <code>@sam_data</code> slot of the physeq object. Must be a character vector or a factor.
<code>level</code>	(required) The level (must be present in modality) of interest
<code>min_nb_seq_taxa</code>	(default 0 = no filter) The minimum number of sequences per taxa
<code>min_nb_samples_taxa</code>	(default 0 = no filter) The minimum number of samples per taxa

Value

A vector of taxa names

A vector of taxa names

Author(s)

Adrien Taudière

See Also

[ggvenn_pq\(\)](#) and [upset_pq\(\)](#)

Examples

```
data_fungi_mini_woNA4height <- subset_samples(
  data_fungi_mini,
  !is.na(data_fungi_mini@sam_data$Height)
)
taxa_only_in_one_level(data_fungi_mini_woNA4height, "Height", "High")
# Taxa present only in low height samples
suppressMessages(suppressWarnings(taxa_only_in_one_level(data_fungi, "Height", "Low")))
# Number of taxa present only in sample of time equal to 15
suppressMessages(suppressWarnings(length(taxa_only_in_one_level(data_fungi, "Time", "15"))))
```

tax_bar_pq

Plot the distribution of sequences or ASV in one taxonomic levels

Description

Graphical representation of distribution of taxonomy, optionnaly across a factor.

Usage

```
tax_bar_pq(
  physeq,
  fact = "Sample",
  taxa = "Order",
  percent_bar = FALSE,
  nb_seq = TRUE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
taxa	(default: 'Order') Name of the taxonomic rank of interest
percent_bar	(default FALSE) If TRUE, the stacked bar fill all the space between 0 and 1. It just set position = "fill" in the ggplot2::geom_bar() function
nb_seq	(logical; default TRUE) If set to FALSE, only the number of ASV is count. Concretely, physeq otu_table is transformed in a binary otu_table (each value different from zero is set to one)

Value

A [ggplot2](#) plot with bar representing the number of sequence en each taxonomic groups

Author(s)

Adrien Taudière

See Also[plot_tax_pq\(\)](#) and [multitax_bar_pq\(\)](#)**Examples**

```
data_fungi_ab <- subset_taxa_pq(data_fungi, taxa_sums(data_fungi) > 10000)
tax_bar_pq(data_fungi_ab) + theme(legend.position = "none")

tax_bar_pq(data_fungi_ab, taxa = "Class")
tax_bar_pq(data_fungi_ab, taxa = "Class", percent_bar = TRUE)
tax_bar_pq(data_fungi_ab, taxa = "Class", fact = "Time")
```

tax_datatable

*Make a datatable with the taxonomy of a [phyloseq-class](#) object***Description**

An interactive table for phyloseq taxonomy.

Usage

```
tax_datatable(
  physeq,
  abundance = TRUE,
  taxonomic_level = NULL,
  modality = NULL,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
abundance	(default: TRUE) Does the number of sequences is print
taxonomic_level	(default: NULL) a vector of selected taxonomic level using their column numbers (e.g. taxonomic_level = 1:7)
modality	(default: NULL) A sample modality to split OTU abundancy by level of the modality
...	Other argument for the datatable function

Value

A datatable

Author(s)

Adrien Taudière

Examples

```
data("GlobalPatterns", package = "phyloseq")
if (requireNamespace("DT")) {
  tax_datatable(subset_taxa(
    GlobalPatterns,
    rowSums(GlobalPatterns@otu_table) > 10000
  ))

  # Using modality
  tax_datatable(GlobalPatterns,
    modality = GlobalPatterns@sam_data$SampleType
  )
}
```

tbl_sum_samdata	<i>Summarize information from sample data in a table</i>
-----------------	--

Description

A wrapper for the `gtsummary::tbl_summary()` function in the case of phyloseq object.

Usage

```
tbl_sum_samdata(physeq, remove_col_unique_value = TRUE, ...)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`remove_col_unique_value` (logical, default TRUE) Do we remove informative columns (categorical column with one value per samples), e.g. samples names ?

... Other arguments pass on to `gtsummary::tbl_summary()`.

Details

This function is mainly a wrapper of the work of others. Please make a reference to `gtsummary::tbl_summary()` if you use this function.

Value

A new `phyloseq-class` object with a larger slot `tax_table`

Author(s)

Adrien Taudière

Examples

```
if (requireNamespace("gtsummary")) {
  tbl_sum_samdata(data_fungi) %>%
    gtsummary::as_kable()

  summary_samdata <- tbl_sum_samdata(data_fungi,
    include = c("Time", "Height"),
    type = list(Time ~ "continuous2", Height ~ "categorical"),
    statistic = list(Time ~ c("{median} ({p25}, {p75})", "{min}, {max}"))
  )
}

data(enterotype)
if (requireNamespace("gtsummary")) {
  summary_samdata <- tbl_sum_samdata(enterotype)
  summary_samdata <- tbl_sum_samdata(enterotype, include = !contains("SampleId"))
}
```

Tengeler2020_pq	<i>This tutorial explores the dataset from Tengeler et al. (2020) available in the mia package. obtained using</i> <code>mia::makePhyloseqFromTreeSE(Tengeler2020)</code>
-----------------	--

Description

This is a phyloseq version of the Tengeler2020 dataset.

Usage

```
data(Tengeler2020_pq)
```

Format

A phyloseq object

Details

Tengeler2020 includes gut microbiota profiles of 27 persons with ADHD. A standard bioinformatic and statistical analysis done to demonstrate that altered microbial composition could be a driver of altered brain structure and function and concomitant changes in the animals behavior. This was investigated by colonizing young, male, germ-free C57BL/6J0laHsd mice with microbiota from individuals with and without ADHD.

Tengeler, A.C., Dam, S.A., Wiesmann, M. et al. Gut microbiota from persons with attention-deficit/hyperactivity disorder affects the brain in mice. *Microbiome* 8, 44 (2020). <https://microbiomejournal.biomedcentral.com/articles/10.1007/s12287-020-00816-x>

track_wkflow	<i>Track the number of reads (= sequences), samples and cluster (e.g. ASV) from various objects including dada-class and derep-class.</i>
--------------	---

Description

- List of fastq and fastg.gz files -> nb of reads and samples
- List of dada-class -> nb of reads, clusters (ASV) and samples
- List of derep-class -> nb of reads, clusters (unique sequences) and samples
- Matrix of samples x clusters (e.g. otu_table) -> nb of reads, clusters and samples
- Phyloseq-class -> nb of reads, clusters and samples

Usage

```
track_wkflow(
  list_of_objects,
  obj_names = NULL,
  clean_pq = FALSE,
  taxonomy_rank = NULL,
  ...
)
```

Arguments

list_of_objects	(required) a list of objects
obj_names	A list of names corresponding to the list of objects
clean_pq	(logical) If set to TRUE, empty samples and empty ASV are discarded before clustering.
taxonomy_rank	A vector of int. Define the column number of taxonomic rank in physeq@tax_table to compute the number of unique value. Default is NULL and do not compute values for any taxonomic rank
...	Other arguments passed on to <code>clean_pq()</code> function.

Value

The number of sequences, clusters (e.g. OTUs, ASVs) and samples for each object.

See Also

[track_wkflow_samples\(\)](#)

Examples

```
data(enterotype)
if (requireNamespace("pbapply")) {
  track_wkflow(list(data_fungi, enterotype), taxonomy_rank = c(3, 5))
}
```

track_wkflow_samples	<i>Track the number of reads (= sequences), samples and cluster (e.g. ASV) for each sample</i>
----------------------	--

Description

Contrary to [track_wkflow\(\)](#), only phyloseq object are possible. More information are available in the manual of the function [track_wkflow\(\)](#)

Usage

```
track_wkflow_samples(list_pq_obj, ...)
```

Arguments

list_pq_obj	(required): a list of object passed on to track_wkflow() Only phyloseq object will return value because information of sample is needed
...	Other args passed on to track_wkflow()

Value

A list of dataframe. cf [track_wkflow\(\)](#) for more information

Author(s)

Adrien Taudière

Examples

```
tree_A10_005 <- subset_samples(data_fungi, Tree_name == "A10-005")
if (requireNamespace("pbapply")) {
  track_wkflow_samples(tree_A10_005)
}
```

transp	<i>Adds transparency to a vector of colors</i>
--------	--

Description

Adds transparency to a vector of colors

Usage

```
transp(col, alpha = 0.5)
```

Arguments

col	a vector of colors
alpha	(default 0.5) a numeric value between 0 and 1 representing the alpha coefficient; 0: total transparency; 1: no transparency.

Value

a color vector

Author(s)

Thibaut Jombart in adegenet package

See Also

The R package RColorBrewer, proposing a nice selection of color palettes. The viridis package, with many excellent palettes

treemap_pq	<i>Plot treemap of 2 taxonomic levels</i>
------------	---

Description

Note that lvl2 need to be nested in lvl1

Usage

```
treemap_pq(  
  physeq,  
  lvl1,  
  lvl2,  
  nb_seq = TRUE,  
  log10trans = TRUE,  
  plot_legend = FALSE,  
  ...  
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
lv11	(required) Name of the first (higher) taxonomic rank of interest
lv12	(required) Name of the second (lower) taxonomic rank of interest
nb_seq	(logical; default TRUE) If set to FALSE, only the number of ASV is count. Concretely, physeq otu_table is transformed in a binary otu_table (each value different from zero is set to one)
log10trans	(logical, default TRUE) If TRUE, the number of sequences (or ASV if nb_seq = FALSE) is log10 transformed.
plot_legend	(logical, default FALSE) If TRUE, plot the legend of color for lv1 1
...	Other arguments passed on to treemapify::geom_treemap() function.

Value

A ggplot2 graphic

Author(s)

Adrien Taudière

Examples

```
data(data_fungi_sp_known)
if (requireNamespace("treemapify")) {
  treemap_pq(
    clean_pq(subset_taxa(
      data_fungi_sp_known,
      Phylum == "Basidiomycota"
    )),
    "Order", "Class",
    plot_legend = TRUE
  )
}
```

```
if (requireNamespace("treemapify")) {
  treemap_pq(
    clean_pq(subset_taxa(
      data_fungi_sp_known,
      Phylum == "Basidiomycota"
    )),
    "Order", "Class",
    log10trans = FALSE
  )
  treemap_pq(
    clean_pq(subset_taxa(
      data_fungi_sp_known,
      Phylum == "Basidiomycota"
    )),
    "Order", "Class",
```

```

    nb_seq = FALSE, log10trans = FALSE
  )
}

```

tsne_pq

Compute tSNE position of samples from a phyloseq object

Description

Compute tSNE position of samples from a phyloseq object

Usage

```
tsne_pq(physeq, method = "bray", dims = 2, theta = 0, perplexity = 30, ...)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
method	A method to calculate distance using <code>vegan::vegdist()</code> function
dims	(Int) Output dimensionality (default: 2)
theta	(Numeric) Speed/accuracy trade-off (increase for less accuracy), set to 0.0 for exact TSNE (default: 0.0 see details in the man page of <code>Rtsne::Rtsne</code>).
perplexity	(Numeric) Perplexity parameter (should not be bigger than $3 * \text{perplexity} < \text{nrow}(X) - 1$, see details in the man page of <code>Rtsne::Rtsne</code>)
...	Other arguments passed on to <code>Rtsne::Rtsne()</code>

Value

A list of element including the matrix Y containing the new representations for the objects. See `?Rtsne::Rtsne()` for more information

Examples

```

if (requireNamespace("Rtsne")) {
  res_tsne <- tsne_pq(data_fungi)
}

```

unique_or_na

Get the unique value in x or NA if none

Description

If `unique(x)` is a single value, return it; otherwise, return an NA of the same type as `x`. If `x` is a factor, then the levels and ordered status will be kept in either case. If `x` is a non-atomic vector (i.e. a list), then the logical NA will be used.

Usage

```
unique_or_na(x)
```

Arguments

`x` A vector

Value

Either a single value (if `unique(x)` return a single value) or a NA

Author(s)

Michael R. McLaren (orcid: [0000-0003-1575-473X](https://orcid.org/0000-0003-1575-473X))

Examples

```
f <- factor(c("a", "a", "b", "c"), ordered = TRUE)
unique_or_na(f)
unique_or_na(f[1:2])

x <- c("a", "b", "a")
unique_or_na(x[c(1, 3)])
unique_or_na(x)
unique_or_na(x) %>% typeof()
```

upset_pq*Make upset plot for phyloseq object.*

Description

Alternative to venn plot.

Usage

```
upset_pq(
  physeq,
  fact,
  taxa_fill = NULL,
  min_nb_seq = 0,
  na_remove = TRUE,
  numeric_fonction = sum,
  rarefy_after_merging = FALSE,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required): Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
taxa_fill	(default NULL) fill the ASV upset using a column in tax_table slot.
min_nb_seq	minimum number of sequences by OTUs by samples to take into count this OTUs in this sample. For example, if min_nb_seq=2, each value of 2 or less in the OTU table will not count in the venn diagram
na_remove	: if TRUE (the default), NA values in fact are removed if FALSE, NA values are set to "NA"
numeric_fonction	(default : sum) the function for numeric vector useful only for complex plot (see examples)
rarefy_after_merging	Rarefy each sample after merging by the modalities of fact parameter
...	Other arguments passed on to the ComplexUpset::upset()

Value

A [ggplot2](#) plot

Author(s)

Adrien Taudière

See Also

[ggvenn_pq\(\)](#)

Examples

```
if (requireNamespace("ComplexUpset")) {
  upset_pq(data_fungi_mini,
    fact = "Height", width_ratio = 0.2,
```

```

    taxa_fill = "Class"
  )
}

if (requireNamespace("ComplexUpset")) {
  upset_pq(data_fungi_mini, fact = "Height", min_nb_seq = 1000)
  upset_pq(data_fungi_mini, fact = "Height", na_remove = FALSE)

  upset_pq(data_fungi_mini, fact = "Time", width_ratio = 0.2, rarefy_after_merging = TRUE)

  upset_pq(
    data_fungi_mini,
    fact = "Time",
    width_ratio = 0.2,
    annotations = list(
      "Sequences per ASV \n (log10)" = (
        ggplot(mapping = aes(y = log10(Abundance)))
        +
          geom_jitter(aes(
            color =
              Abundance
          ), na.rm = TRUE)
        +
          geom_violin(alpha = 0.5, na.rm = TRUE) +
          theme(legend.key.size = unit(0.2, "cm")) +
          theme(axis.text = element_text(size = 12))
        ),
      "ASV per phylum" = (
        ggplot(mapping = aes(fill = Phylum))
        +
          geom_bar() +
          ylab("ASV per phylum") +
          theme(legend.key.size = unit(0.2, "cm")) +
          theme(axis.text = element_text(size = 12))
        )
      )
    )
  )

  upset_pq(
    data_fungi_mini,
    fact = "Time",
    width_ratio = 0.2,
    numeric_fonction = mean,
    annotations = list(
      "Sequences per ASV \n (log10)" = (
        ggplot(mapping = aes(y = log10(Abundance)))
        +
          geom_jitter(aes(
            color =
              Abundance
          ), na.rm = TRUE)
        +
          geom_violin(alpha = 0.5, na.rm = TRUE) +

```

```

        theme(legend.key.size = unit(0.2, "cm")) +
        theme(axis.text = element_text(size = 12))
    ),
    "ASV per phylum" = (
      ggplot(mapping = aes(fill = Phylum))
      +
      geom_bar() +
      ylab("ASV per phylum") +
      theme(legend.key.size = unit(0.2, "cm")) +
      theme(axis.text = element_text(size = 12))
    )
  )
)

upset_pq(
  subset_taxa(data_fungi_mini, Phylum == "Basidiomycota"),
  fact = "Time",
  width_ratio = 0.2,
  base_annotations = list(),
  annotations = list(
    "Sequences per ASV \n (log10)" = (
      ggplot(mapping = aes(y = log10(Abundance)))
      +
      geom_jitter(aes(
        color =
          Abundance
      )), na.rm = TRUE)
      +
      geom_violin(alpha = 0.5, na.rm = TRUE) +
      theme(legend.key.size = unit(0.2, "cm")) +
      theme(axis.text = element_text(size = 12))
    ),
    "ASV per phylum" = (
      ggplot(mapping = aes(fill = Class))
      +
      geom_bar() +
      ylab("ASV per Class") +
      theme(legend.key.size = unit(0.2, "cm")) +
      theme(axis.text = element_text(size = 12))
    )
  )
)

data_fungi2 <- data_fungi_mini
data_fungi2@sam_data[["Time_0"]] <- data_fungi2@sam_data$Time == 0
data_fungi2@sam_data[["Height__Time_0"]] <-
  paste0(data_fungi2@sam_data[["Height"]], "__", data_fungi2@sam_data[["Time_0"]])
data_fungi2@sam_data[["Height__Time_0"]][grepl("NA", data_fungi2@sam_data[["Height__Time_0"]])] <-
  NA
upset_pq(data_fungi2, fact = "Height__Time_0", width_ratio = 0.2, min_size = 2)
}

```

upset_test_pq *Test for differences between intersections*

Description

See [upset_pq\(\)](#) to plot upset.

Usage

```
upset_test_pq(
  physeq,
  fact,
  var_to_test = "OTU",
  min_nb_seq = 0,
  na_remove = TRUE,
  numeric_fonction = sum,
  ...
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required): Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
var_to_test	(default c("OTU")) : a vector of column present in the tax_table slot from the physeq object
min_nb_seq	minimum number of sequences by OTUs by samples to take into count this OTUs in this sample. For example, if min_nb_seq=2,each value of 2 or less in the OTU table will not count in the venn diagram
na_remove	: if TRUE (the default), NA values in fact are removed if FALSE, NA values are set to "NA"
numeric_fonction	(default : sum) the function for numeric vector useful only for complex plot (see examples)
...	Other arguments passed on to the ComplexUpset::upset_test()

Value

A [ggplot2](#) plot

Author(s)

Adrien Taudière

See Also

[upset_pq\(\)](#)

Examples

```
data(data_fungi)
if (requireNamespace("ComplexUpset")) {
  upset_test_pq(data_fungi, "Height", var_to_test = c("OTU", "Class", "Guild"))
  upset_test_pq(data_fungi, "Time")
}
```

var_par_pq	<i>Partition the Variation of a phyloseq object by 2, 3, or 4 Explanatory Matrices</i>
------------	--

Description

The function partitions the variation in `otu_table` using distance (Bray per default) with respect to two, three, or four explanatory tables, using adjusted R^2 in redundancy analysis ordination (RDA) or distance-based redundancy analysis. If response is a single vector, partitioning is by partial regression. Collinear variables in the explanatory tables do NOT have to be removed prior to partitioning. See [`vegan::varpart\(\)`](#) for more information.

Usage

```
var_par_pq(
  physeq,
  list_component,
  dist_method = "bray",
  dbrda_computation = TRUE
)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`list_component` (required) A named list of 2, 3 or four vectors with names from the `@sam_data` slot.

`dist_method` (default "bray") the distance used. See [`phyloseq::distance\(\)`](#) for all available distances or run [`phyloseq::distanceMethodList\(\)`](#). For "aitchison" and "robust.aitchison" distance, [`vegan::vegdist\(\)`](#) function is directly used.

`dbrda_computation` (logical) Do dbrda computations are runned for each individual component (each name of the list component) ?

Details

This function is mainly a wrapper of the work of others. Please make a reference to [`vegan::varpart\(\)`](#) if you use this function.

Value

an object of class "varpart", see [`vegan::varpart\(\)`](#)

Author(s)

Adrien Taudière

See Also[var_par_rarperm_pq\(\)](#), [vegan::varpart\(\)](#), [plot_var_part_pq\(\)](#)**Examples**

```

if (requireNamespace("vegan")) {
  data_fungi_woNA <-
    subset_samples(data_fungi, !is.na(Time) & !is.na(Height))
  res_var <- var_par_pq(data_fungi_woNA,
    list_component = list(
      "Time" = c("Time"),
      "Size" = c("Height", "Diameter")
    ),
    dbrda_computation = TRUE
  )
}

```

var_par_rarperm_pq	<i>Partition the Variation of a phyloseq object with rarefaction permutations</i>
--------------------	---

Description

This is an extension of the function [var_par_pq\(\)](#). The main addition is the computation of nperm permutations with rarefaction even depth by sample. The return object

Usage

```

var_par_rarperm_pq(
  physeq,
  list_component,
  dist_method = "bray",
  nperm = 99,
  quantile_prob = 0.975,
  dbrda_computation = FALSE,
  dbrda_signif_pval = 0.05,
  sample.size = min(sample_sums(physeq)),
  verbose = FALSE,
  progress_bar = TRUE
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
list_component	(required) A named list of 2, 3 or four vectors with names from the @sam_data slot.
dist_method	(default "bray") the distance used. See phyloseq::distance() for all available distances or run phyloseq::distanceMethodList() . For aitchison and robust.aitchison distance, vegan::vegdist() function is directly used.#' @param fill_bg
nperm	(int) The number of permutations to perform.
quantile_prob	(float, [0:1]) the value to compute the quantile. Minimum quantile is compute using 1-quantile_prob.
dbrda_computation	(logical) Do dbrda computations are runned for each individual component (each name of the list component) ?
dbrda_signif_pval	(float, [0:1]) The value under which the dbrda is considered significant.
sample.size	(int) A single integer value equal to the number of reads being simulated, also known as the depth. See phyloseq::rarefy_even_depth() .
verbose	(logical). If TRUE, print additional informations.
progress_bar	(logical, default TRUE) Do we print progress during the calculation?

Details

This function is mainly a wrapper of the work of others. Please make a reference to [vegan::varpart\(\)](#) if you use this function.

Value

A list of class varpart with additional information in the \$part\$indfract part. Adj.R.square is the mean across permutation. Adj.R.squared_quantil_min and Adj.R.squared_quantil_max represent the quantile values of adjuste R squared

Author(s)

Adrien Taudière

See Also

[var_par_pq\(\)](#), [vegan::varpart\(\)](#), [plot_var_part_pq\(\)](#)

Examples

```
if (requireNamespace("vegan")) {
  data_fungi_woNA <- subset_samples(data_fungi, !is.na(Time) & !is.na(Height))
  res_var_9 <- var_par_rarperm_pq(
    data_fungi_woNA,
    list_component = list(
```

```

      "Time" = c("Time"),
      "Size" = c("Height", "Diameter")
    ),
    nperm = 9,
    dbrda_computation = TRUE
  )
  res_var_2 <- var_par_rarperm_pq(
    data_fungi_woNA,
    list_component = list(
      "Time" = c("Time"),
      "Size" = c("Height", "Diameter")
    ),
    nperm = 2,
    dbrda_computation = TRUE
  )
}

```

 venn_pq

Venn diagram of [phyloseq-class](#) object

Description

Graphical representation of distribution of taxa across combined modality of a factor.

Usage

```
venn_pq(physeq, fact, min_nb_seq = 0, print_values = TRUE)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
fact	(required): Name of the factor to cluster samples by modalities. Need to be in physeq@sam_data.
min_nb_seq	(default: 0): minimum number of sequences by OTUs by samples to take into count this OTUs in this sample. For example, if min_nb_seq=2, each value of 2 or less in the OTU table will be change into 0 for the analysis
print_values	(logical) Print (or not) the table of number of OTUs for each combination. If print_values is TRUE the object is not a ggplot object. Please use print_values = FALSE if you want to add ggplot function (cf example).

Value

A [ggplot2](#) plot representing Venn diagram of modalities of the argument factor

Author(s)

Adrien Taudière

See Also[venneuler](#)**Examples**

```
if (requireNamespace("venneuler")) {
  data("enterotype")
  venn_pq(enterotype, fact = "SeqTech")
}

if (requireNamespace("venneuler")) {
  venn_pq(enterotype, fact = "ClinicalStatus")
  venn_pq(enterotype, fact = "Nationality", print_values = FALSE)
  venn_pq(enterotype, fact = "ClinicalStatus", print_values = FALSE) +
    scale_fill_hue()
  venn_pq(enterotype, fact = "ClinicalStatus", print_values = FALSE) +
    scale_fill_hue()
}
```

`verify_pq`*Verify the validity of a phyloseq object*

Description

Mostly for internal use in MiscMetabar functions.

Usage

```
verify_pq(
  physeq,
  verbose = FALSE,
  min_nb_seq_sample = 500,
  min_nb_seq_taxa = 1
)
```

Arguments

`physeq` (required): a [phyloseq-class](#) object obtained using the phyloseq package.

`verbose` (logical, default FALSE) If TRUE, prompt some warnings.

`min_nb_seq_sample` (numeric) Only used if `verbose = TRUE`. Minimum number of sequences per samples to not show warning.

`min_nb_seq_taxa` (numeric) Only used if `verbose = TRUE`. Minimum number of sequences per taxa to not show warning.

Value

Nothing if the phyloseq object is valid. An error in the other case. Warnings if verbose = TRUE

vsearch_clustering	<i>Recluster sequences of an object of class phyloseq or cluster a list of DNA sequences using vsearch software</i>
--------------------	---

Description

A wrapper of VSEARCH software.

Usage

```
vsearch_clustering(
  physeq = NULL,
  dna_seq = NULL,
  nproc = 1,
  id = 0.97,
  vsearchpath = "vsearch",
  tax_adjust = 0,
  vsearch_cluster_method = "--cluster_size",
  vsearch_args = "--strand both",
  keep_temporary_files = FALSE
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
dna_seq	You may directly use a character vector of DNA sequences in place of physeq args. When physeq is set, dna sequences take the value of physeq@refseq
nproc	(default: 1) Set to number of cpus/processors to use for the clustering
id	(default: 0.97) level of identity to cluster
vsearchpath	(default: vsearch) path to vsearch
tax_adjust	(Default 0) See the man page of merge_taxa_vec() for more details. To conserved the taxonomic rank of the most abundant ASV, set tax_adjust to 0 (default). For the moment only tax_adjust = 0 is robust
vsearch_cluster_method	(default: "--cluster_size) See other possible methods in the vsearch manual (e.g. --cluster_size or --cluster_smallmem) <ul style="list-style-type: none"> • --cluster_fast : Clusterize the fasta sequences in filename, automatically sort by decreasing sequence length beforehand. • --cluster_size : Clusterize the fasta sequences in filename, automatically sort by decreasing sequence abundance beforehand.

- `--cluster_smallmem` : Clusterize the fasta sequences in filename without automatically modifying their order beforehand. Sequence are expected to be sorted by decreasing sequence length, unless `--usersort` is used. In that case you may set `vsearch_args` to `vsearch_args = "--strand both --usersort"`
- `vsearch_args` (default : "--strand both") a one length character element defining other parameters to be passed on to `vsearch`.
- `keep_temporary_files`
(logical, default: FALSE) Do we keep temporary files ?
- `temp.fasta` (refseq in `fasta` or `dna_seq` sequences)
 - `cluster.fasta` (centroid if `method = "vsearch"`)
 - `temp.uc` (clusters if `method = "vsearch"`)

Details

This function use the `merge_taxa_vec()` function to merge taxa into clusters. By default `tax_adjust = 0`. See the man page of `merge_taxa_vec()`.

This function is mainly a wrapper of the work of others. Please cite [vsearch](#).

Value

A new object of class `physeq` or a list of cluster if `dna_seq` args was used.

Author(s)

Adrien Taudière

References

VSEARCH can be downloaded from <https://github.com/torognes/vsearch>. More information in the associated publication <https://pubmed.ncbi.nlm.nih.gov/27781170>.

See Also

[asv2otu\(\)](#), [swarm_clustering\(\)](#)

Examples

```
summary_plot_pq(data_fungi)
d_vs <- vsearch_clustering(data_fungi)
summary_plot_pq(d_vs)
```

vs_search_global	<i>Search for a list of sequence in a fasta file against physeq reference sequences using R</i> hrefhttps://github.com/torognes/vsearch/vsearch
------------------	---

Description

Use of VSEARCH software.

Usage

```
vs_search_global(  
  physeq,  
  seq2search = NULL,  
  path_to_fasta = NULL,  
  vsearchpath = "vsearch",  
  id = 0.8,  
  iddef = 0,  
  keep_temporary_files = FALSE  
)
```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
seq2search	(required if path_to_fasta is NULL) Either (i) a DNAstringSet object or (ii) a character vector that will be convert to DNAstringSet using Biostrings::DNAstringSet()
path_to_fasta	(required if seq2search is NULL) a path to fasta file if seq2search is est to NULL.
vsearchpath	(default: vsearch) path to vsearch
id	(default: 0.8) id for the option --usearch_global of the vsearch software
iddef	(default: 0) iddef for the option --usearch_global of the vsearch software
keep_temporary_files	(logical, default: FALSE) Do we keep temporary files <ul style="list-style-type: none"> • temp.fasta (refseq in fasta) • cluster.fasta (centroid) • temp.uc (clusters)

Details

This function is mainly a wrapper of the work of others. Please cite [vsearch](#).

Value

A dataframe with uc results (invisible)

Author(s)

Adrien Taudière

Examples

```

if (requireNamespace("seqinr")) {
  file_dna <- tempfile("dna.fa")
  seqinr::write.fasta("GCCCATAGTATTCTAGTGGGCATGCCTGTTCGAGCGTCATTTTCAACC",
    file = file_dna, names = "seq1"
  )

  res <- vs_search_global(data_fungi, path_to_fasta = file_dna)
  unlink(file_dna)

  res[res$identity != "*", ]

  clean_pq(subset_taxa(data_fungi, res$identity != "*"))
}

```

write_pq

Save phyloseq object in the form of multiple csv tables.

Description

This is the reverse function of [read_pq\(\)](#).

Usage

```

write_pq(
  physeq,
  path = NULL,
  rdata = FALSE,
  one_file = FALSE,
  write_sam_data = TRUE,
  sam_data_first = FALSE,
  clean_pq = TRUE,
  reorder_asv = FALSE,
  rename_asv = FALSE,
  remove_empty_samples = TRUE,
  remove_empty_taxa = TRUE,
  clean_samples_names = TRUE,
  silent = FALSE,
  verbose = FALSE,
  quote = FALSE,
  sep_csv = "\t",
  ...
)

```

Arguments

physeq	(required): a phyloseq-class object obtained using the phyloseq package.
path	a path to the folder to save the phyloseq object
rdata	(logical) does the phyloseq object is also saved in Rdata format?
one_file	(logical) if TRUE, combine all data in one file only
write_sam_data	(logical) does the samples data are add to the file. Only used if one_file is TRUE. Note that these option result in a lot of NA values.
sam_data_first	(logical) if TRUE, put the sample data at the top of the table Only used if one_file and write_sam_data are both TRUE.
clean_pq	(logical) If set to TRUE, empty samples are discarded after subsetting ASV
reorder_asv	(logical) if TRUE the otu_table is ordered by the number of sequences of ASV (descending order). Default to TRUE. Only possible if clean_pq is set to TRUE.
rename_asv	reorder_asv (logical) if TRUE, ASV are renamed by their position in the OTU_table (asv_1, asv_2, ...). Default to FALSE. Only possible if clean_pq is set to TRUE.
remove_empty_samples	(logical) Do you want to remove samples without sequences (this is done after removing empty taxa)
remove_empty_taxa	(logical) Do you want to remove taxa without sequences (this is done before removing empty samples)
clean_samples_names	(logical) Do you want to clean samples names?
silent	(logical) If true, no message are printing.
verbose	(logical) Additional informations in the message the verbose parameter overwrite the silent parameter.
quote	a logical value (default FALSE) or a numeric vector. If TRUE, any character or factor columns will be surrounded by double quotes. If a numeric vector, its elements are taken as the indices of columns to quote. In both cases, row and column names are quoted if they are written. If FALSE nothing is quoted.
sep_csv	(default tabulation) separator for column
...	Other arguments passed on to utils::write.table() function.

Value

Build a folder (path) containing one to four csv tables (refseq.csv, otu_table.csv, tax_table.csv, sam_data.csv) and if present a phy_tree in Newick format

Author(s)

Adrien Taudière

See Also

[save_pq\(\)](#)

Examples

```
write_pq(data_fungi, path = paste0(tempdir(), "/phyloseq"))  
write_pq(data_fungi, path = paste0(tempdir(), "/phyloseq"), one_file = TRUE)  
unlink(paste0(tempdir(), "/phyloseq"), recursive = TRUE)
```

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